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Evaluation of doxycycline sensitivity *in vitro* against *Babesia gibsoni* by Real Time-PCR

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Abstract

Doxycycline sensitivity against *in vitro* cultured *Babesia gibsoni* was evaluated by real-time PCR and parasitemia. The culture of *B. gibsoni* was successfully continued and mean parasitemia was 4.63% when *in vitro* drug sensitivity test was started. The drug sensitivity test by the real-time PCR calculated the gene copy number from cultured sample. Even if the complete growth inhibition was at certain concentration, the genomic DNA might remain in existence. The results revealed that the doses of doxycycline 8 µM, 16 µM, 32 µM and 64 µM of both real-time PCR and parasitemia inhibited the growth of *B. gibsoni* in a dose-dependent manner at 96 h and 144 h. We determined the IC₅₀ of doxycycline 17.9 µM at 96 h and 13.8 µM at 144 h after treatment using real-time PCR. Morphological observation revealed that the number of parasites decreased per erythrocyte and line shaped parasites increased in erythrocytes with the increased concentration of doxycycline. Doxycycline effectively inhibits the growth of *B. gibsoni in vitro*. Therefore, doxycycline can be used to treat *B. gibsoni* infection. Further studies are important to know the doxycycline resistance in *B. gibsoni*.

Keywords: Doxycycline, *Babesia gibsoni*, RT-PCR, Growth inhibition

Introduction

B. gibsoni is a tick-borne intra-erythrocytic protozoan parasite. It is one of the most common causal agents of severe canine haemolytic anemia. Dogs typically present with the acute form of babesiosis characterized by fever, pallor mucous membrane, lymphadenopathy, splenomegaly, and eventual death (Boozar and Macintire 2003). Chronic infection is common. Definitive diagnosis is based on the clinical signs, history and the identification of *Babesia* spp. within the erythrocytes, positive serologic results and amplification of nucleic acid extracted from blood or tissues (Birkenheuer *et al.*, 2004). Several drugs have been used for the clinical management of canine babesiosis including diminazene aceturate, imidocarb dipropionate (Fowler *et al.*, 1972), pentamidine isethionate phenamidine isethionate (Farwell *et al.*, 1982). Although, these drugs show clinical effectiveness slowly (Suzuki *et al.*, 2007; Lin and Huang, 2010) but these drugs have some adverse effects such as pain at the injection site and nervous symptom due to cerebral haemorrhage (Wulansari *et al.*, 2003). However, no report has discussed the effectiveness of other clinically usable drugs that are available. Some new therapies with antibiotics like clindamycin, metronidazole, doxycycline enrofloxacin have been reported to reduce parasitemia in experimentally infected and in clinical patients (Suzuki *et al.*, 2007; Lin and Huang 2010). Tetracycline antibiotics have been recognized to exhibit activity against protozoa such as *Ehrlichia canis* and *P. falciparum*. In *Babesia* parasites, the activity against *B. divergens* (Losson and Patz, 1989) and *Babesia canis* has been recognized only *in vivo* (Vercammen *et al.* 1966). Most dogs recovered from the acute stage of the disease are at risk of recurrence and may become a reservoir for tick-borne infections (Conrad *et al.*, 1991). Although these therapeutic modalities are beneficial and seem to cure partially, the treatment failed for some of the dogs. Few studies have evaluated the effect of extracts of traditional Indonesian medicinal plants against *B. gibsoni* by using *in vitro* culture and the results of these studies are promising from the view point of future anti-babesial drug development (Groves *et al.*, 1972; Murnigsih *et al.*, 2005; Suzuki *et al.*, 2007). To evaluate the effect of potential drugs against this pathogen, an *in vitro* susceptibility test would be important by Real Time-PCR (RT-PCR) but limited information is available. Because *in vitro* culturing of this parasite is difficult and it is successfully conducted only in a limited number of research institutes (Sunaga *et al.*, 2002; Murase *et al.*, 1991). *B. gibsoni* infected dogs either clinical patient or experimental are required for this method. Our laboratory established a continuous culture of *B. gibsoni* isolated from Tosa dogs (Matsuu *et al.*, 2004a). There is

no precise report emphasizing the efficacy of doxycycline sensitivity alone *in vitro* against *B. gibsoni* by RT-PCR. In this study, the growth inhibitory effect of different concentrations of doxycycline against *B. gibsoni* tested in a continued culture and evaluated by RT-PCR at different time points. Also the IC₅₀ of doxycycline was determined.

Materials and Methods

Test compound

Doxycycline hydrochloride was purchased from Sigma-Aldrich (Tokyo, Japan). Different doses of doxycycline (DXC) 2 μ M, 4 μ M, 8 μ M, 16 μ M, 32 μ M and 64 μ M were used to evaluate the growth inhibitory effects.

Packed cell volume and thrombocyte count

Measurements of packed cell volume (PCV) and thrombocyte counts were performed using an automated blood cell counter (Celltaq MEK6400, Nihon Kohden Co, Tokyo, Japan).

Culture of *B. gibsoni*

B. gibsoni parasites were isolated from a naturally infected Tosa dog in Aomori Prefecture, Japan (Matsuu *et al.*, 2004a). The 18S rDNA sequences of this parasite exhibit 100% homology with the genotype of *B. gibsoni* isolated from dogs in Asia, Oklahoma, North Carolina and Okinawa (Matsuu *et al.*, 2004b). The parasite was maintained in the laboratory by passage in beagle dogs.

Preparation of erythrocytes from the infected blood sample for culture

The blood sample was washed 3 times with phosphate-buffered saline (PBS) by centrifugation at 4 × *g* for 10 min at 4 °C. The culture medium was RPMI-1640 (Invitrogen, Carlsbad, CA, USA) supplemented with 25 mM pyruvic acid, 2 mM L-glutamine, 100 units/mL penicillin G, 100 μ g/mL streptomycin (Invitrogen) and 10% fetal bovine serum (FBS; Funakoshi, Tokyo, Japan). Canine erythrocytes sufficient to yield a packed cell volume (PCV) of 10% were obtained from a healthy Beagle. Packed infected RBCs (200 μ l) were dispensed into 1800 μ l of culture medium in each well of a 12-well plate. This plate was incubated at 37 °C in a humidified atmosphere containing 5% CO₂ and 5% O₂ for 7 days. The supernatant medium (800 μ l) was replaced daily by a fresh medium. Sub-culturing was performed 7 days after primary culturing was initiated. Normal fresh erythrocytes were sampled from normal beagles by using sodium citrate as the anticoagulant and immediately washed 3 times with PBS. We transferred 200 μ l of the cultured and infected erythrocyte suspension to a new well containing 1620 μ l of fresh medium and 180 μ l of a normal dog erythrocyte suspension. Subsequent continuous culturing was performed every 5–7 days. The percent parasitemia at subculture was calculated by enumeration of 2000 total infected and uninfected RBC on Giemsa-stained smears under microscope. The ensuing parasitemia was monitored daily as above.

Growth inhibitory test of doxycycline

For the *in vitro* sensitivity test, *B. gibsoni* cultures that had reached parasitemia approximately 5% (parasitemia before adding drug = Pre) after the last subculture for 2-3 days were collected in a collection tube. Two hundred mL of these parasitized erythrocyte suspension was distributed per well in 96-well plates in three replicates of each drug concentration. Doxycycline hydrochloride was dissolved in ultra pure water and aliquot was prepared and then added to the culture medium to give the final concentrations of 2 μ M, 4 μ M, 8 μ M, 16 μ M, 32 μ M and 64 μ M. This range of drug concentrations were based on the results of preliminary assays conducted at the laboratory. From the aliquots drug was added into each well of the serially diluted doxycycline in culture medium and one well without drug (ND) as control for normal growth of parasite. The plates were incubated in a 5% CO₂ incubator at 37°C. Every 24 h the medium (80 μ L) in each well was aspirated and a fresh solution containing the appropriate concentration of the test drug was added. After 48, 72, 96 and 144 h of incubation from each well 1 μ L sample was taken for parasitemia and 1 μ L for RT-PCR.

Real-Time PCR

DNA was extracted using DNA extraction kit (QIAamp DNA Mini Kit; Qiagen, USA) and Real-Time PCR was performed using a standard protocol recommended by the manufacturer (iQ SYBR Green Supermix, Bio-Rad, Japan). One μL of template DNA was added to 19 μL of reaction mixture containing 0.5 μL of each primer, 8 μL ultra pure water, and 10 μL iQ SYBR Green Supermix. The reaction was performed under the following conditions (Mini Opticon, Bio-Rad) 95°C for 30s. 95°C for 3min, 55°C for 5s. A thermal melt profile was built following PCR to identify amplicons as specified, and analysis was performed with analysis software (CFX-Manager, Bio-Rad). The IC_{50} of doxycycline was defined as the concentration required for the 50% reduction in the mean number of parasitized erythrocytes from that of control culture after 96 and 144 h of incubation. Each test was performed in triplicates. IC_{50} values were expressed as the mean concentration $\pm\text{SD}$.

Parasitemia

A thin smear was prepared, fixed with methanol and stained with Giemsa solution to count the number of parasitized erythrocytes. Parasitemia was monitored by counting the number of parasitized cells among 2000 erythrocytes and expressing this as a percentage.

The sensitivity of doxycycline was evaluated by measuring the rate of growth inhibition. This was calculated by counting the number of parasitized erythrocytes per 2000 erythrocytes in each of the wells containing the drug and that in the control well without drug (ND).

Results and Discussion

B. gibsoni culture was successfully continued. Parasitemia reduced upto 0.5% in few phases but recovered and cultured parasite grew steadily and gained considerable size as a result of continuous culturing (Fig. 1).

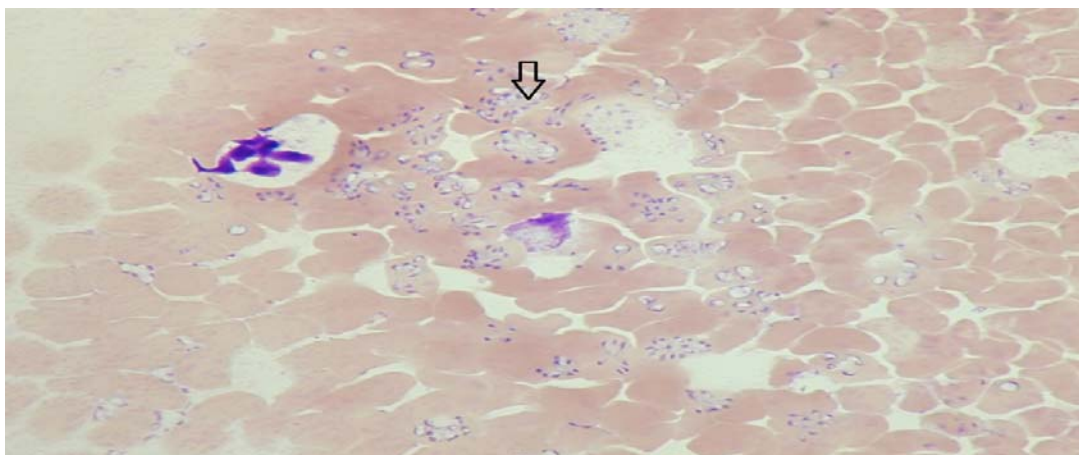


Fig 1. The arrow showing the growth of *Babesia gibsoni* in the sub-culture under microscope (10x)

The rate of growth inhibition of *B. gibsoni* culture has been shown in a tabular form (Table 1).

Table 1. The percentages (%) of parasitemia are shown as *in vitro* doxycycline sensitivity against *Babesia gibsoni*

Drug groups	48 h (Mean±SD)	72 h (Mean±SD)	96 h (Mean±SD)	144 h (Mean±SD)
Pre	4.633±1.528			
ND	4.80±0.778	5.066±1.189	6.433±1.247	7.766±0.286
2 µM	4.30±1.042	4.333±0.784	6.533±1.265	6.821±0.153
4µM	4.20±0.668	4.533±1.225	6.30±1.555	7.066±1.020
8µM	3.833±0.974	3.0±1.042	3.233±0.188	2.466±0.377
16µM	2.466±0.880	1.70±0.883	2.433±0.659	1.90±0.374
32µM	1.266±0.094	1.033±0.249	0.666±0.094	0.50±0.163
64µM	0.650±0.150	0.50±0.141	0.433±0.124	0.266±0.047

The mean parasitemia was 4.6±1.5% when *in vitro* drug sensitivity test started and mean parasitemia have been counted at 48 h, 72h, 96 h and 144 h after incubation. The sensitivity was evaluated by growth inhibition, determined from the RT-PCR and parasitemia at 96 h and 144 h in different concentrations of doxycycline treated groups Figs. 2 and 3).

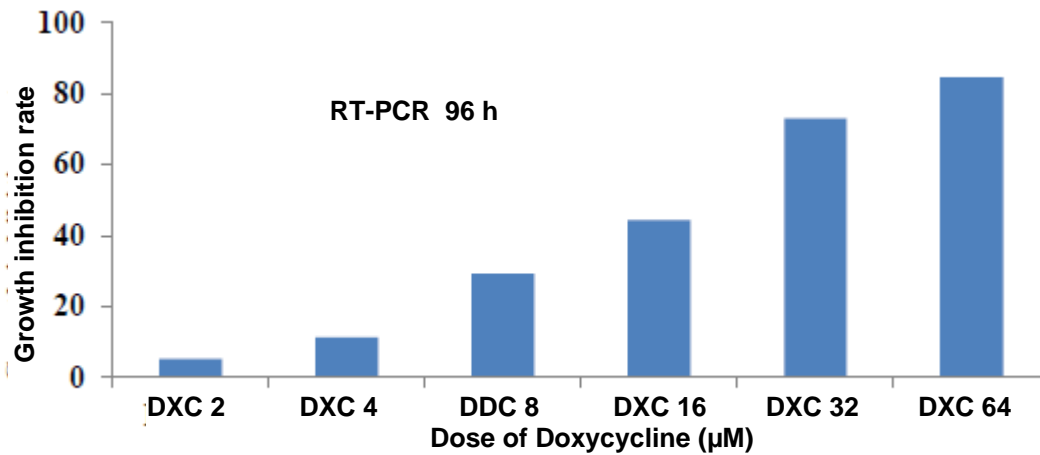


Fig. 2. Growth inhibitory effects of different doses of doxycycline evaluated by RT-PCR at 96 h

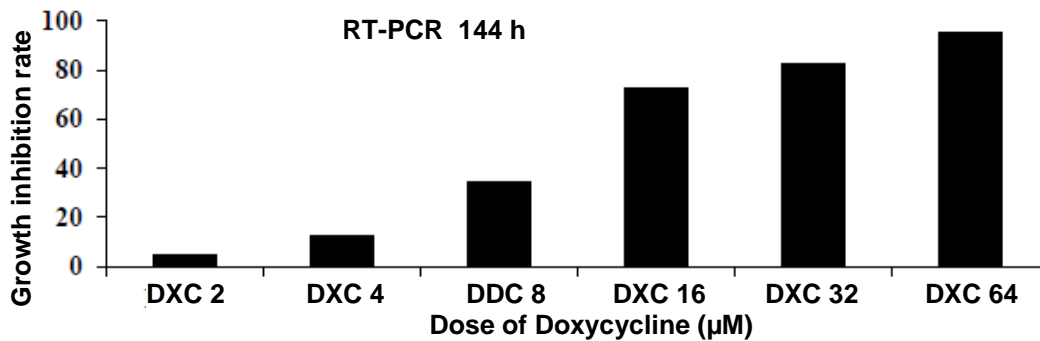


Fig 3. Growth inhibitory effects of different doses of doxycycline evaluated by RT-PCR at 144 h.

The IC₅₀ of doxycycline was found 17.9 µM and 13.8 µM at 96 h and 144 h, respectively. The RT-PCR results revealed that the doses of doxycycline 8µM, 16 µM, 32µM and 64 µM inhibited the growth of *B. gibsoni* in a dose-dependent manner at 96 h (Fig. 2) and at 144 h (Fig. 3). Morphological observation revealed that the number of parasites decreased per erythrocyte and line shaped parasites increased in erythrocytes with the increased concentrations of doxycycline. It was recognized that doxycycline effectively inhibit the growth of *B. gibsoni* *in vitro*.

Previous studies demonstrated the growth inhibitory effect of different antibiotics including macrolide and lincosamides against protozoa. These antibiotics probably exhibit inhibitory effects against the apicomplex protozoa by targeting the apicoplast (Vercammen *et al.*, 1996). For a long time, tetracycline antibiotics have been recognized to exhibit activity against protozoa such as *Ehrlichia canis* and *P. falciparum*. In *Babesia* parasites, the activity against *B. divergens* (Losson and Patz, 1989) and *Babesia canis* has been recognized only *in vivo* (Vercammen *et al.*, 1996). The efficacy of treatment of babesiosis caused by *B. gibsoni* with doxycycline has been evaluated *in vivo* but the ability of this drug alone was controversial. This study will help to understand the growth inhibitory effect of doxycycline by RT-PCR in a better way. Doxycycline provided satisfactory prophylaxis against experimental infection with a highly pathogenic strain of *Babesia canis* but asymptomatic infection could not be ruled out. Imidocarb provides a rather short protection period (Vercammen *et al.*, 1996) and has some practical inconvenience regarding availability and administration mode. In addition, tetracyclines are also useful against canine ehrlichiosis. The IC₅₀ value for doxycycline was slightly higher for *P. falciparum* than for *B. gibsoni*, which was previously reported to be 10.2–17.0 and 7.7–14.9 µM, respectively (Pradines *et al.*, 2001).

It is concluded that treatment of canine babesiosis by oral administration of doxycycline can be of practical importance. Further studies are important to evaluate the doxycycline resistance gene in *B. gibsoni*.

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