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Estimation of sensitivity and specificity arising from validity studies with incomplete designs

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Abstract. This insert introduces `valides`, `validesi`, and `validesu`, a set of commands that allow the calculations of point and precision estimates of sensitivity and specificity obtained in validity studies with incomplete designs.

Keywords: st0019, validity, sensitivity, specificity, incomplete design

1 Description

`valides` estimates sensitivity (Se) and specificity (Sp) arising from multi-stage (phase) study designs, whereby a *new test* (NT) under scrutiny for its concurrent validity is applied first to all subjects and the *reference test* (RT) thereafter on only a subsample of those. This may be suitable, for instance, when the prevalence of an event is relatively rare and the RT is costly, invasive, potentially risky, etc. Very often, the RT is applied to all $NT+$, but to just a fraction of $NT-$, say, on a sample size up to five times that of $NT+$.

`validesi` is the immediate form of `valides`. Here, cell values $\#a$, $\#b$, $\#c$, and $\#d$ comprising a 2×2 contingency table of NT by RT are directly typed as arguments instead of read from a dataset stored in memory. The program is intended for use in the same setting described for `valides`, but is also meant for data originated from multi-phase designs such as, for instance, when the reference procedure is evaluated on (new) test-positives collected in one setting (dataset), whereas independently applied to another (external) sample of test-negatives. Such study designs may be contemplated for logistic, financial, or even convenience reasons.

The population quantities to be estimated in validity studies with complete designs are in Table 1, where $Se = 100(P_A/P_{M_1})$ and $Sp = 100(P_D/P_{M_0})$. Table 2 portrays the structure of the data in classic one-stage designs when all subjects are simultaneously measured through NT and RT , following a random or systematic sampling procedure. Here, direct estimation of sensitivity— $Se = 100(p_a/p_{m_1})$ —and specificity— $Sp = 100(p_d/p_{m_0})$ —are possible (Streiner and Norman 1995).

Table 1: Population parameters to be estimated.

	NT+	NT-	
RT+	P_A	P_B	P_{M_1}
RT-	P_C	P_D	P_{M_0}
	P_{N_1}	P_{N_0}	1

Table 2: Classic one-stage/phase sampling design.

	(a) Absolute values				(b) Proportions		
	NT+	NT-			NT+	NT-	
RT+	a	b	m_1	\longleftrightarrow	p_a	p_b	p_{m_1}
RT-	c	d	m_0	\longleftrightarrow	p_c	p_d	p_{m_0}
	n_1	n_0	n		p_{n_1}	p_{n_0}	1

Tables 3(a), (b), and (c) show a generic situation in which there is an incomplete design due to the breaking up of the measurement procedure. Table 3(a) pictures a step whereby NT is first applied to all subjects. Tables 3(b) and (c) illustrate the sequel of the process, a second step where RT is given to sub-groups of NT+ and NT-. Usually, n'_1 and n'_0 are, respectively, subsamples of n_1 and n_0 , but sometimes $n_1 = n'_1$.

Table 3: Diagram referring to the incomplete, multi-stage/phase sampling design—(a) step for collecting information for the estimation of $p_{n_1} = n_1/n$; (b) step for collecting information for the estimation of $p_\alpha = a'/n'_1$ and $p_\delta = d'/n'_0$: absolute values; and (c) ditto: proportions.

	(a)				(b)				(c)	
	NT+	NT-			NT+	NT-			NT+	NT-
RT+	—	—	\longrightarrow	RT+	a'	b'	\longleftrightarrow	p_a	p_b	
RT-	—	—	\longrightarrow	RT-	c'	d'	\longleftrightarrow	p_c	p_d	
	n_1	n_0	n		n'_1	n'_0				

Since sensitivity and specificity cannot be assessed directly from incomplete study designs, *valides/validesi* indirectly calculates these quantities by means of

$$Se = \frac{p_{n_1} p_\alpha}{p_\alpha p_{n_1} + (1 - p_{n_1}) (1 - p_\delta)} \tag{1}$$

$$Sp = \frac{p_\delta (1 - p_{n_1})}{p_{n_1} + p_\delta - p_{n_1} p_\alpha - p_{n_1} p_\delta} \tag{2}$$

where p_{n_1} is the proportion of positive subjects according to the *test*; p_α is the proportion

of positive subjects detected by the *reference test* among the $NT+$ previously sampled; and p_δ is the proportion of negative subjects detected by the *reference test* among the $NT-$ previously sampled. Equations (1) and (2) are derived from those presented by Choi (1992) a few years ago and are adapted for the purpose of this program.

`valides` and `validesi` also calculate confidence intervals, allowing for the variability occurring in both stages (phases) of the sampling procedure to be taken into account. These can be estimated via maximum likelihood or parametric bootstrap methods. The former is the default. Companion ado-files `_vld_lf.ado` and `_sim_lf.ado` relating, respectively, to each method are needed.

The maximum-likelihood estimation routine uses `ml`'s `lf` method on an internally reshaped data structure and assumes that the number of $NT+$ in the first stage and both $RT+ | NT+$ and $RT- | NT-$ in the second are binomially distributed. The log-likelihood function is written in terms of sensitivity (Se), specificity (Sp), and the prevalence of the event (P), the three quantities of interest.

Given the equalities

$$p_{n_1} = Se Sp + (1 - Sp) (1 - P) \quad (3)$$

$$q_{n_1} = 1 - p_{n_1} \quad (4)$$

$$p_\alpha = \frac{Se P}{Se P + (1 - Sp) (1 - P)} \quad (5)$$

$$q_\alpha = 1 - p_\alpha \quad (6)$$

$$p_\delta = \frac{Sp (1 - P)}{Sp + P - P (Se + Sp)} \quad (7)$$

$$q_\delta = 1 - p_\delta \quad (8)$$

and the reshaped data being

(Continued on next page)

$$\begin{aligned}
 X_1 &= \begin{cases} 1, & NT+ \\ 0, & NT- \end{cases} \\
 X_2 &= \begin{cases} 1, & NT+, RT+ \\ 0, & NT+, RT- \\ \cdot, & NT+, RT_{\text{missing}} \end{cases} \\
 X_3 &= \begin{cases} 1, & NT-, RT+ \\ 0, & NT-, RT- \\ \cdot, & NT-, RT_{\text{missing}} \end{cases}
 \end{aligned}$$

the log-likelihood function is

$$\ln f = \begin{cases} \ln(q_{n_1}) + X_1 * \ln(p_{n_1}/q_{n_1}), \\ \quad \text{for } X_2 = \cdot, X_3 = \cdot \\ \\ \ln(q_{n_1}) + X_1 * \ln(p_{n_1}/q_{n_1}) + \ln(q_\alpha) + X_2 * \ln(p_\alpha/q_\alpha), \\ \quad \text{for } X_2 \neq \cdot, X_3 = \cdot \\ \\ \ln(q_{n_1}) + X_1 * \ln(p_{n_1}/q_{n_1}) + \ln(q_\delta) + X_3 * \ln(p_\delta/q_\delta), \\ \quad \text{for } X_2 = \cdot, X_3 \neq \cdot \end{cases} \quad (9)$$

In the optional parametric bootstrap procedure, the means and distributions of the quantities of interest are iteratively calculated in 6 steps and are detailed in Table 4. First, r simulated sensitivity and specificity values are obtained— $Se^{(i)}$ and $Sp^{(i)}$ —using, at each i iteration, the simulated proportions $p_{n_1}^{(i)}$, $p_\alpha^{(i)}$, and $p_\delta^{(i)}$ (steps 1 to 5). The desired statistics are then calculated from the r simulated Se and Sp values (step 6). Centiles are of particular interest for the confidence intervals. For example, a 95% CI can be specified by detecting the 2.5 and 97.5 simulated centiles. Alternatively, standard deviates are estimated from the collection of simulated values and are used to parametrically calculate the CI. The two procedures tend to converge as the number of replications increase. The projected prevalence and respective confidence intervals may be directly obtained since $P = (p_{n_1} \cdot p_\alpha) / Se$. The procedure needs `rndbin` to be installed (Hilbe and Linde-Zwirble 1996). Note that if there are any zero-cells arising from the data in memory (or from inputted values when using `validesi`), estimations are not possible and an appropriate warning is issued.

Table 4: Iterative algorithm for the calculation of confidence intervals for sensitivity and specificity obtained from validity studies with incomplete designs.

Step	Procedure
1	a Simulating n Bernoullis with parameter $p_{n_1} = n_1/n$ b Calculating the proportion of $NT+$ in iteration $i \rightarrow p_{n_1}^{(i)}$
2	a Simulating n'_1 Bernoullis with parameter $p_\alpha = a'/n'_1$ b Calculating the proportion of $(RT+ T+)$ in iteration $i \rightarrow p_\alpha^{(i)}$
3	a Simulating n'_0 Bernoullis with parameter $p_\delta = d'/n'_0$ b Calculating the proportion of $(RT- T-)$ in iteration $i \rightarrow p_\delta^{(i)}$
4	Calculating Se and Sp in iteration i using, respectively, equations (1) and (2), but substituting the terms for $p_{n_1}^{(i)}$, $p_\alpha^{(i)}$ and $p_\delta^{(i)} \rightarrow Se^{(i)}$ and $Sp^{(i)}$
5	Exporting $Se^{(i)}$ and $Sp^{(i)}$ to an “external” reception dataset ↓ ↓
6	Detecting the centiles corresponding to the limits of the respective $100(1 - \alpha)\%$ confidence intervals

The package also includes `validesu`, which allows a re-analysis of the simulated data previously generated and saved by `valides` or `validesi`. These saved values may also be used to further explore the results, for instance, by visually plotting distributions obtained for different strata of interest.

2 Syntax

```
valides refvar testvar [if exp] [in range] [ , prv notab dec2 level(#)
  simul reps(#) saving(filename) using(filename) ]
```

testvar is the variable that identifies the result of the new diagnostic test, and *refvar* is the variable containing the real status of subjects. Note that the lower category must identify the non-exposed or the negative result of NT and RT or the false status of the patient, and must assume value 0. Positive results assume value 1.

```
validesi #a #b #c #d [ , ttot(#) tpos(#) prv notab dec2 level(#)
  simul reps(#) saving(filename) using(filename) ]
```

Cells *#a* and *#b* relate to the $NT+$ and $NT-$ among the $RT+$. Conversely, cells *#c* and *#d* relate to the NT status among the $RT-$. Unless the study design is complete, `validesi` needs to be informed of the total NT sample using `ttot()`, as well as the number of detected $NT+$ using `tpos()` (see details below).

```
validesu [ , using(filename) prv dec2 level(#) ]
```

3 Options

`prv` requests the display of the projected prevalence of the event.

`notab` suppresses the display of the 2×2 table containing the *testvar* by *refvar* cross-tabulation.

`dec2` requests 2-digit displays.

`level(#)` specifies the confidence level (%) for the confidence intervals for sensitivity, specificity, and prevalence (if option `prv` is requested). The default is 95%.

`simul` specifies that confidence intervals are to be calculated via simulation (parametric bootstrap) instead of via the default maximum likelihood method.

`reps(#)` specifies the number of simulations to be performed. The default value of 100 is used when the option is not requested. The minimum is 2. `reps()` may only be active if `simul` is also specified.

`saving(filename)` requests that simulated values be dumped to a file called *filename* located in the `pwd` for further use (e.g., by `validesu`). `saving()` may only be active if `simul` is also specified.

`using(filename)` requests that the analysis be based on the simulated values previously dumped to an external file by `saving()`. When this option is requested in `valides`, only `level()`, `notab`, and `dec2` remain active. `validesu` should be used for re-analysing data generated by `validesi`. `using()` may only be active if `simul` is also specified.

`ttot(#)` is specific to `validesi`. It provides the total number of subjects undergoing the New Test. If not stated, this is the sum of *#a*, *#b*, *#c*, and *#d* values specified by the user, implicitly assuming an analysis for a complete study design.

`tpos(#)` is specific to `validesi`. It provides the total number of positive subjects according to the *new test*. If not stated, this is the sum of values *#a* and *#b*, implicitly assuming that no subsampling of *NT+* was carried out.

4 Examples

Consider a (fictitious) study to assess the concurrent validity of a new data collection instrument (*test*) developed from an accepted *reference test*, with the aim to gain operational efficiency with minimal loss in validity. For that purpose, the new instrument *test* has been thoroughly reduced in item numbers and has been submitted to a field test.

Referring back to the notation presented in Table 1, assume that the population parameters are $P_A = 0.016$, $P_B = 0.098$, $P_C = 0.004$, and $P_D = 0.882$, with $Se = 0.80$ and $Sp = 0.90$ to be estimated. If the researcher had used a complete study design, the formal analysis using `diagt` (Seed and Tobias 2001) would have been

```
. diagt RT NT
```

Reference Test	New Test		Total
	Pos.	Neg.	
Abnormal	80	20	100
Normal	490	4410	4900
Total	570	4430	5000

True abnormal diagnosis defined as RT = 1 (labelled Positive)

[95% Conf. Inter.]

Sensitivity	Pr(+ D)	80.00%	70.82%	87.33%
Specificity	Pr(- ~D)	90.00%	89.13%	90.83%
Positive predictive value	Pr(D +)	14.04%	11.29%	17.16%
Negative predictive value	Pr(~D -)	99.55%	99.30%	99.72%

Prevalence	Pr(D)	2.00%	1.63%	2.43%
------------	-------	-------	-------	-------

Alternatively, ci could have been employed:

```
. ci NT if RT==1
```

Variable	Obs	Mean	Std. Err.	[95% Conf. Interval]	
NT	100	.8	.0402015	.7202315	.8797685

```
. gen NT_i=1-NT
. ci NT_i if RT==0
```

Variable	Obs	Mean	Std. Err.	[95% Conf. Interval]	
NT_i	4900	.9	.0042862	.8915972	.9084028

The analysis with `valides` using the default ml-based estimations yields quite similar results, as required:

```
. valides RT NT, prv dec2
```

Reference Test	New Test		Total
	Positive	Negative	
Positive	80	20	100
Negative	490	4410	4900
Total	570	4430	5000

Note: The data and specifications assume a complete study design.

True D defined by RT [95% Conf. Inter.]
ml based

Sensitivity	Pr(+ D)	80.00%	72.16%	87.84%
			(s_err = 4.00)	
Specificity	Pr(- ~D)	90.00%	89.16%	90.84%
			(s_err = 0.43)	
Prevalence (projected)		2.00%	1.61%	2.39%
			(s_err = 0.20)	

Note that `valides` “understands” that the study design is complete. Also note that the projected prevalence has been requested, yielding a result quite consistent with the population parameter.

Minding the costs and/or inconvenience involved in applying the *RT* to all subjects, the researcher decides to do this for all *NT+*, but only to a fraction of *NT-*. The researcher opts for a 1:2 (sub)sample-size ratio, which still implies reducing to 1/3 the number of subjects to whom the *RT* needs to be offered. A plausible dataset and the ensuing analysis with `valides` could be as follows:

```
. list id NT RT
      id      NT      RT
  1.     1 Negative Negative
  2.     2 Negative Negative
(output omitted)
1134.  1134 Positive Negative
1135.  1135 Positive Negative
(output omitted)
1625.  1625 Negative Positive
1626.  1626 Negative Positive
(output omitted)
1631.  1631 Positive Positive
1632.  1632 Positive Positive
(output omitted)
1711.  1711 Negative      .
(output omitted)
5000.  5000 Negative      .
```

```
. valides RT NT, prv dec2
Reference Test |      New Test      | Total
                | Positive Negative |
-----|-----|-----
Positive       |      80      6    |     86
Negative       |     490    1134   |    1624
-----|-----|-----
Total          |     570    1140   |    1710
```

```
Reference Test applied to T+ and T-
Total sample size for Test = 5000
Test positives = 570
```

```
True D defined by RT          [95% Conf. Inter.]
                               ml based
-----|-----|-----
Sensitivity Pr( +| D)  77.43%  62.96%  91.90%
                               (s_err = 7.38)

Specificity Pr( -|-D)  89.99%  89.15%  90.83%
                               (s_err = 0.43)

Prevalence (projected)      2.07%   1.56%  2.57%
                               (s_err = 0.26)
-----|-----|-----
```

Inevitably, there was a loss in statistical efficiency since there is now less data available in the incomplete study design. In the case of this example, perhaps the researcher would still be willing to accept the amount of imprecision in the estimates, but a final de-

cision would need to be made in the light of how much the gains in operational efficiency would outweigh this relative deficiency. Note that, as before, `valides` “understood” the data, this time as originating from an incomplete study design. In passing, the same output would have been obtained using

```
. validesi 80 6 490 1134, ttot(5000) tpos(570) prv dec2
```

A sub-group analysis could also be requested, for instance, according to severity of the underlying event/disease. The estimations using a 90% CI would be

```
. valides RT NT if D==1, le(90)
(2500 observations deleted)
```

Reference Test	New Test		Total
	Positive	Negative	
Positive	45	2	47
Negative	245	567	812
Total	290	569	859

Reference Test applied to T+ and T-
 Total sample size for Test = 2500
 Test positives = 290

True D defined by RT [90% Conf. Inter.]
 ml based

Sensitivity	Pr(+ D)	85.3%	70.4%	100.0%
			(s_err = 9.1)	
Specificity	Pr(- -D)	90.0%	89.0%	91.0%
			(s_err = 0.6)	

```
. valides RT NT if D==0, le(90)
(2500 observations deleted)
```

Reference Test	New Test		Total
	Positive	Negative	
Positive	35	4	39
Negative	245	567	812
Total	280	571	851

Reference Test applied to T+ and T-
 Total sample size for Test = 2500
 Test positives = 280

True D defined by RT [90% Conf. Inter.]
 ml based

Sensitivity	Pr(+ D)	69.2%	50.8%	87.7%
			(s_err = 11.2)	
Specificity	Pr(- -D)	90.0%	89.0%	91.0%
			(s_err = 0.6)	

Once again `valides` acknowledges the data, correctly using the appropriate subsets in the analysis.

Similar results would have been produced when using the parametric bootstrap method. Although admittedly less efficient from a computational stand, an immediate bonus of this option would be the ability to further explore the data using the simulated data dumped to an external file. Perhaps the researcher would be interested in a visual comparison of the *new test*'s properties according to the severity of disease. The following simple do-file would produce Figure 1:

```
use valides.dta, clear
valides RT NT if D=1, sim reps(10000) saving(simfile1)
valides RT NT if D=0, sim reps(10000) saving(simfile0)

use simfile0.dta, clear
rename s_sens s_sens0
rename sens sens0
keep s_num s_sens0 sens0
sort s_num
save simf0tmp, replace

use simfile1.dta, clear
rename s_sens s_sens1
rename sens sens1
keep s_num s_sens1 sens1
sort s_num
merge using simf0tmp
erase simf0tmp.dta

label var s_sens1 "BS Sensitivity, Disease=1"
label var s_sens0 "BS Sensitivity, Disease=0"
label var sens1 "Sensitivity, Disease=1"
label var sens0 "Sensitivity, Disease=0"

gr s_sens0 s_sens1 s_num, xlab ylab s(!!!) c(1[_]1)
```

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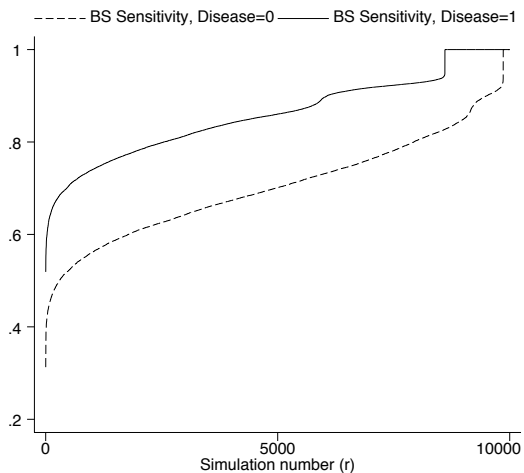


Figure 1: Distribution of BS simulated values of sensitivity according to disease status

For completeness, the output using `valides` with `simul` and `saving()` options on the disease positives and `validesu` applied to previously saved data on the disease negatives is shown next. As expected, estimations are indeed consistent with those obtained with the `ml` method.

```
. valides RT NT if D==1, sim reps(10000) saving(simfile1) notab
(2500 observations deleted)
```

```
Reference Test applied to T+ and T-
Total sample size for Test = 2500
Test positives = 290
```

Please wait ... calculating confidence intervals

True D defined by RT		[95% Conf. Inter.]				
		s_err based		centile based		
Sensitivity	Pr(+ D)	85.3%	67.9%	100.0%	67.6%	100.0%
			(s_err= 8.9)			
Specificity	Pr(- -D)	90.0%	88.8%	91.2%	88.8%	91.2%
			(s_err= 0.6)			

(reps = 10000)

```
Values dumped on 22 Apr 2002 15:39
to simfile1.dta on the pwd
```

```
. validesu, using(simfile0)

Using data in simfile0
True D defined by RT
```

		[95% Conf. Inter.]				
		s_err based		centile based		
Sensitivity	Pr(+ D)	69.2%	46.8%	91.7%	49.0%	92.0%
			(s_err= 0.1)			
Specificity	Pr(- -D)	90.0%	88.8%	91.2%	88.8%	91.2%
			(s_err= 0.0)			

```
-----
(reps in simulation data = 10000)
```

5 Saved Results

`valides`, `validesi`, and `validesu` save in `r()`:

Scalars

```
r(sens)      sensitivity
r(se.ci.l)   lower standard-error-based confidence interval for sensitivity
r(se.ci.u)   upper standard-error-based confidence interval for sensitivity
r(se.c.l)    lower centile-based confidence interval for sensitivity *
r(se.c.u)    upper centile-based confidence interval for sensitivity *
r(se.s_err)  standard error for sensitivity
r(spec)      specificity
r(sp.ci.l)   lower standard-error-based confidence interval for specificity
r(sp.ci.u)   upper standard-error-based confidence interval for specificity
r(sp.c.l)    lower centile-based confidence interval for specificity *
r(sp.c.u)    upper centile-based confidence interval for specificity *
r(sp.s_err)  standard error for specificity
r(s_prev)    prevalence (projected)
r(pr.ci.l)   lower standard-error-based confidence interval for prevalence
r(pr.ci.u)   upper standard-error-based confidence interval for prevalence
r(pr.c.l)    lower centile-based confidence interval for prevalence *
r(pr.c.u)    upper centile-based confidence interval for prevalence *
r(pr.s_err)  standard error for prevalence
r(z)         z score
r(p.alpha)   alpha probability *
r(p.delta)   delta probability *
```

Note: those marked with an asterisk are only obtained when option `simul` is used. The others are always provided but relate to the applied estimation method.

6 References

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