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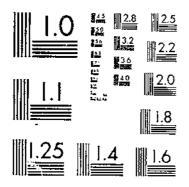
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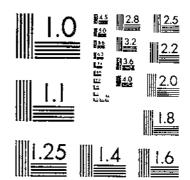
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T6 1428 (1971). USBA TECHNICAL BULLETINS: * UPDATA
NEBRCTIC SAWFLIES: 111 HETERARTHRINAE, ADULTS AND LARVALA
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# **NEARCTIC SAWFLIFS**

III. Heterarthrinae: Adults and Larvae (Hymenoptera: Tenthredinidae)

By DAVID R. SMITH

Technical Bulletin No. 1420

Agricultural Research Service
U.S. DEPARTMENT OF AGRICULTURE

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#### NEARCTIC SAWFLIES.

# III. Heterarthrinae: Adults and Larvae (Hymenoptera: Tenthredinidae)

By DAVID R. SMITH, entomologist, Systematic Entomology Laboratory, Agricultural Research Service

The Heterarthrinae are represented in America north of Mexico by 37 species in 12 genera and three tribes. This subfamily includes some of the most economically important sawfiles, such as the birch leaf miner (Fenusa pusilla (Lepeletier)), the elm leaf miner (Fenusa ulmi Sundevall), the European alder leaf miner (Fenusa dohrnii (Tischbein)), the rose-slug (Endelomyia aethiops (Fabricius)), and the pear-slug (Caliroa cerasi (Linnaeus)). Members of the tribes Heterarthrini and Fenusini all are leaf miners in the larval stage and may be destructive to Alnus, Betula, Crataegus, Platanus, Populus, Potentilla, Prunus, Quercus, Rubus, Salix, Ulmus, and Viola. Members of the tribe Caliroini are external feeders in the larval stage and skeletonize the leaves of Castanea, Nyssa, Prunus, Pyrus, Quercus, Rosa, and Salix.

This bulletin is the first comprehensive revision of the Heterarthrinae and is intended to provide a means for identifying the Nearctic genera and species of this subfamily. Other information may be found in the literature cited, which includes most of the

pertinent literature for each taxon.

₫.

The subfamily limits accepted here are those proposed by Ross (1937, 1951). Specimens may be keyed to this subfamily by using Ross' 1937 key to the subfamilies of Tenthredinidae. The name "Phyllotominae" was used for this subfamily at that time. Benson (1952) treated the components of this group in a different manner. His concept of the subfamily Heterarthrinae was more limited and included only the genus Heterarthrus. Caliroa and Endelomyia were included in the tribe Caliroini of the Blennocampinae, and the Fenusini were also considered a tribe in the Blennocampinae.

The adults of the Heterarthrinae may be characterized as follows: Vein M and 1m-cu of forewing divergent; vein M meets Sc+R at or near junction of Rs+M and Sc+R; vein 2A and 3A complete for entire length, connected to 1A by oblique crossvein in Caliroini and Heterarthrini, or vein 2A and 3A partially atrophied, leaving anal cell petiolate and vein 2A and 3A present as stub, which may be straight, curved up, or curved up and meeting

<sup>&#</sup>x27;The year in italic after the author's name is the key to the reference in Literature Cited, p. 69.

1A to form small basal anal cell in the Fenusini. The wing venation of various species is shown in plates I, 3-9; II, 10-18; and III, 19-20. In general, these sawflies are small black robust species.

The sawflies of the Heterarthrinae are divided into three groups, each composed of closely related genera, and here treated as the

tribes Caliroini, Heterarthrini, and Fenusini.

The Caliroini include two genera, Endelomyia and Caliroa, which are separated from the other tribes by the complete 2A and 3A vein of the forewing and the tarsal claw with a single outer tooth and large acute basal lobe.

Heterarthrus is the only genus in the Heterarthrini, which is a Palaearctic group with only one introduced species found in North America. This tribe is separated from the others by the complete 2A and 3A vein of the forewing, the 10- or 11-segmented antenna, and the tarsal claw with two subequal outer teeth and a large acute basal lobe.

The Fenusini, represented in North America by nine genera, are separated from the other tribes by the petiolate anal cell of the forewing and with only the basal stub of vein 2A and 3A present. The genera in this tribe may be divided further into three groups: (1) Those genera with cell  $R_1$  closed in the hindwing—Messa, Metallus, and Setabara; (2) those genera with cell  $R_1$  open in the hindwing, with a simple tarsal claw, and with the stub of vein 2A and 3A of the forewing curved up and meeting 1A to form a small basal anal cell—Fenusa, Fenella, and Prolatus; (3) those genera with cell  $R_1$  open in the hindwing, with a tarsal claw having a large acute basal lobe, and with the stub of vein 2A and 3A of the forewing either straight or curved up at its apex—Profenusa, Bidigitus, and Nefusa.

The larvae of the Heterarthrinae fall into three distinct groups: (1) The typically sawflylike larva of *Endelomyia aethiops*, an external feeder (pl. XI, 130-136). The body is neither flattened nor sluglike (or tadpolelike) in appearance. The larva of this species may be separated from other sawfly larvae by the mandibles, which have four truncate teeth on the ventral margin and

by the presence of tubercles on the thorax, on annulets 2 and 4 of abdominal segments 1 to 8, and on terga 9 and 10.

(2) The sluglike or tadpolelike larvae of Caliroa, all external feeders (pl. XI, 137-141). These larvae have an enlarged thorax with the body sharply narrowing at the base and toward the apex of the abdomen; the head is hypognathous; the thoracic legs are reduced in size; and a large fleshy protuberance stems anteriorly from each prothoracic leg.

(3) The small dorsoventrally flattened larvae of the Heterarthrini and Fenusini (pls. XII-XVIII), all leaf miners. The head is usually dorsoventrally flattened and prognathous; the thoracic legs are usually reduced, sometimes lacking tarsal claws; various numbers of sclerotized plates are present on the venter of the body; and the prolegs are reduced or wanting.

The larvae are not well known for the Caliroa species, but they are well known for most of the leaf-mining species. Many of the leaf-mining species have been reared and the larvae have been described in the literature.

The terminology used in this revision is that of Ross (1937) and Smith (1969) for adults and that of Yuasa (1922), Lorenz and Kraus (1957), and Smith (1969) for larvae. I have examined the types of all species discussed in this bulletin except those located in Europe.

#### Hosts

The known host plants for the North American species of Heterarthrinae are as follows:

Plant family and gen Salicaceae:		Insect species
Salix	{	(Caliroa ladrata MacGillivray (Messa wuestneii (Konow)
Populus	,	( Messa hortulana (Kiug) Messa leucostoma (Rohwer) Messa populifoliella (Townsend)
Betulaceae:		
Betula		(Fenusa pusilla (Lepeletier) Heterarthrus nemoratus (Fallén) Messa nana (Klug) Profenusa thomsoni (Konow)
Alnus Fagaceae:	*** * *** -	Fenusa dohrnii (Tischbein)
Quercus		(Calivoa fasciata (Norton) Caliroa lobata MacGillivray Caliroa obsoleta (Norton) Caliroa petiolata, new species Caliroa quarcuscoccineae (Dyar) Profenusa alumna (MacGillivray) Profenusa inspirata (MacGillivray) Profenusa lucifex (Ross)
Castanea		Caliroa lorata MacGillivray
Ulmaceae:		
Ulmus		Fenusa ulmi Sundevall
Platanaceae:		
Platanus	****	Bidigitus platani (Burks)
Cornaceae:		
Nyssa		Caliroa nyssae, new species
Violaceae:		
Viola		Nefusa ambigua (Norton)
Rosaceae:		
Rosa		Endelomyia aethiops (Fabricius)
Rubus	····· {	Metallus capitalis (Norton) Metallus rohweri MacGillivray
Crataegus		Profenusa canadensis (Marlatt)
Potentilla		Fenella nigrita Westwood

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Prunus	(Caliroa cerasi (Linnaeus)   Caliroa liturata MacGillivray   Profenusa canadensis (Marlatt)   Setabara histrionica (MacGillivray) (?)
Pyrus	** * * *

The Calire species recorded on Ceanothus, Cotoneaster, and Acer by Smith (1967) are larvae only. They cannot be ascribed to a particular species.

# Systematic Arrangement

#### Family TENTHREDINIDAE

#### Subfamily HETERARTHRINAE

#### Tribe CALIROINI

#### Genus Endelomyia Ashmead

(1) Endelomyia aethiops (Fabricius); British Columbia, Colorado, Connecticut, Delaware, District of Columbia, Idaho, Illinois, Kansas, Kentucky, Maine, Massachusetts, Michigan, Minnesota, Missouri, Montana, New Jersey, New York, Ohio, Oregon, Pennsylvania, Utah, Virginia, Washington; Palaearctic; on Rosa.

#### Genus Caliroa O. Costa

(2) Caliroa cerasi (Linnaeus); widespread in North America; also in Australia, New Zealand, South Africa, South America; Palaearctic; on Prunus, Pyrus, other Rosaceae.

(3) Caliroa distincta, new species; British Columbia, California, Nevada, Oregon.

(4) Caliroa fasciata (Norton); Connecticut, Illinois, Iowa, Maine, Maryland, New Jersey, New York, Ohio, Ontario, Virginia; on Quercus.

(5) Caliroa foridana, new species; Florida.
(6) Caliroa hyalina, new species; Oregon.
(7) Caliroa labrata MacGillivray; Alberta, British Columbia, California,

Nevada, Oregon; on Salix.

(8) Caliroa liturata MacGillivray; Colorado, Connecticut, Florida, Georgia, Illinois, Louisiana, Missouri, Montana, New York, Rhode Island, South Caro-

Hina, Tennessee, Virginia; on Prunus.

(9) Catiroa lobata MacGillivray; Illinois, Iowa, Maryland, Michigan, New York, Texas, Vermont, Virginia; on Quercus.

(10) Caliroa lorata MacGillivray; Alabama, Maine, Maryland, Massachusetts, New York, North Carolina, Pennsylvania, Virginia, Wisconsin; on Castanea.

(11) Caliroa lunata MacGillivray; Illinois, Michigan, Newfoundland, New Hampshire, New York, Ohio, Pennsylvania, Texas, Virginia, West Virginia. (12) Caliroa nyssae, new species; Georgia, Maryland, Mississippi, Pennsylvania, Virginia; on Nyssa.

(13) Caliroa obsoleta (Norton); Connecticut, Georgia, Illinois, Iowa, Massachusetts, Michigan, Minnesota, New Jersey, New York, Ontario, Virginia, Wisconsin; on Quercus.
(14) Caliroa petiolata, new species; Maryland, Pennsylvania, Virginia;

on Quercus.

(15) Caliroa quercuscoccineae (Dyar); Connecticut, Delaware, District of Columbia, Illinois, Louisiana, Maine, Maryland, Michigan, Minnesota, Missouri, New Hampshire, New Jersey, North Carolina, Pennsylvania, Virginia, Wisconsin; on Quercus.

#### Tribe HETERARTHRINI

#### Genus Heterarthrus Stephens

(16) Heterarthrus nemoratus (Fallén); Maine, Massachusetts, New Brunswick, Newfoundland, New Hampshire, New York, Nova Scotia, Ontario, Quebec; Palaearctic; leaf miner of Betula.

#### Tribe FENUSINI

#### Genus Metallus Forbes

(17) Metallus bensoni, new species; British Columbia, New York.
(18) Metallus capitalis (Norton); British Columbia, Illinois, Maine, New Brunswick, Newfoundland, New Hampshire, New York, Oregon, Quebec; leaf miner of Rubus.

(19) Metallus rohweri MacGillivray; Connecticut, Delaware, Florida, Illinois, Maine, Maryland, Massachusetts, Michigan, Missouri, New Jersey, New York, North Carolina, Ohio, Ontario, Pennsylvania, Rhode Island, Virginia, Wisconsin; leaf miner of Rubus.

#### Genus Messa Leach

(20) Messa hortulana (Klug); Massachusetts; Palaearctic; leaf miner of Populus.

(21) Messa leucostoma (Rohwer); Alberta, Galifornia, Colorado, Iowa, Minnesota, New Brunswick, Oregon, Washington; leaf miner of Populus.
(22) Messa nana (Klug); Maine, New York; Palaearctic; leaf miner of

Betula.

(23) Messa populifoliella (Townsend); Arizona, Golorado, Connecticut, Michigan, New Brunswick, New Mexico, Ontario, South Dakota; leaf miner

(24) Messa wiestneii (Konow); Alaska, British Columbia, California, Colorado, Nevada; Palaearctic; leaf miner of Saliz.

#### Genus Setabara Ross

(25) Setabara histrionica (MacGillivray); California, Colorado, Idaho, Nevada, Oregon, Washington; leaf miner of Prunus (?).

#### Genus Profenusa MacGillivray

(26) Profesusa alumna (MacGillivray); Illinois, Maine, Maryland, New York, Pennsylvania, Virginia; leaf miner of Querous.

(27) Profesusa canadensis (Marlatt); Arkansas, District of Columbia, Illinois, Iowa, Kentucky, Maine, Massachusetts, Michigan, Missouri, New York, Ontario, Pennsylvania, Quebec, Texas; leaf miner of Crataegus and Prunus.

(28) Profesusa inspirata (MacGillivray); California, Nevada, Oregon;

leaf miner of Quercus.

(29) Profesusa lucifex (Ross); Illinois, Maine, New York, Ontario; leaf

miner of Quercus.

(30) Profenusa thomsoni (Konow); Connecticut, Maine, Ontario, Quebec, Vermont; Palaearctic; leaf miner of Betula.

#### Genus Bidiaitus Smith

(31) Bidigitus platani (Burks); California; leaf miner of Platanus.

#### Genus Nefusa Ross

(32) Nefusa ambigua (Norton); Maine, Maryland, Massachusetts, Michigan, New York, Ohio, Pennsylvania, Quebec, Tennessee, Wisconsin; leaf miner of Viola.

#### Genus Prolatus Smith

(33) Prolatus artus Smith; Oregon.

#### Genus Fenusa Leach

(34) Fenusa dohrnii (Tischbein); Alberta, British Columbia, Colorado, Connecticut, Illinois, Maine, Massachusetts, Michigan, New Brunswick, Newfoundland, New Hampshire, New Mexico, New York, Nova Scotia, Ontario, Oregon, Pennsylvania, Saskatchewan, Utah, Washington; Palaearctic; leaf miner of Alnus.

(35) Fenusa pusilla (Lepeletier); Connecticut, Maine, Massachusetts, New Brunswick, Newfoundland, New Hampshire, New Jersey, New York, Nova Scotia, Ontario, Oregon, Quebec, Vermont; Palaearctic; leaf miner of Betula.

(36) Fenusa ulmi Sundevall; Massachusetts, Michigan, New York, Ontario,

Quebec; Palaearctic; leaf miner of Ulmus.

#### Genus Fenella Westwood

(37) Fenella nigrita Westwood; Connecticut, Maine, Michigan, Ontario; Palaearctic; leaf miner of Potentilla.

# Keys to Heterarthrinae Genera

#### ADDITOR

	ADULTS
	Forewing with vein 2A and 3A complete for its entire length, connected to 1A by oblique crossvein (pl. I, 1, 3, 8)
2,	Antenna with 10 or more segments (pl. III, 31); tarsal claw with two subequal outer teeth and large acute basal lobe (pl. III, 23); head markedly depressed between eyes (pl. III, 30); Heterarthrini
3.	Antenna nine-segmented (pl. IV, 40-42); tarsal claw with one outer tooth and large acute basal lobe (pl. III, 22); head convex between eyes (pl. III, 29); Caliroini
u.	gradually decreasing in length (pl. IV, 42); basal emargination of anal cell of forewing with short basally projecting spur (pl. I, 1); anal cell of hindwing petiolate (pl. I, 2)  Endelowing Ashmed
	Apical four antennal segments reduced in length, together only slightly longer than third segment (pl. IV, 40, 41); basal emargination of anal cell of forewing without spur (pl. I, 3); anal cell of hindwing sessile or petiolate (pl. I, 4, 5). Caliroa O. Costa
4.	Cell & of findwing open at apex (nl. 11, 15, 16, 19)
	Tarsal claw with one or two outer teeth and large acute basal lobe (pl. II, 12, 23); stub of 2A and 3A of forewing straight or curved up at apex (pl. II, 14, 17); hindwing with anal cell present or
	absent (pl. II, 15, 16)
6.	cell absent (pl. III, 20)  Tarsal claw with two subequal outer teeth (pl. III, 23) Bidigitus Smith
7.	Tarsai claw with single outer tooth (pl. III, 22)
	Prepectus present

3.	Antenna long and slender, third and fourth segments subequal in length (pl. III, 37); foretarsus 1½ times or more longer than foretibia (pl. III, 25)  Prolatus Smith
9.	Antenna stout, third segment longer than fourth segment (pl. III, 32-34); foretarsus subequal in length to foretibia (pl. III, 26)
10.	Antenna nine-segmented (pl. III, 32, 33); hindtarsus with fourth segment conspicuously produced apically (pl. III, 27). Fenusa Leach Tarsal claw with small indistinct lobe at base, appearing simple (pl. III, 24). Setabara Ross
11.	Tarsal claw with large acute basal lobe (pl. III, 22). 11 Second antennal segment broader than long (pl. III, 33, 39); third and fourth antennal segments usually subequal in length  Metallus Forbes
	Second antennal segment longer than broad (pl. III, 35, 36); third antennal segment usually longer than fourth segment: Messa Leach
	LARVAE
	(based on final feeding stage)
1.	Sluglike or typically sawflylike in appearance; body cylindrical, not dorsoventrally flattened; head round, not dorsoventrally flattened; prolegs distinct; external feeders (pl. XI)  Modified; body dorsoventrally flattened; head usually dorsoventrally flattened; prolegs reduced or absent; leaf miners (pls. XII-XVIII)
2.	Head higher than wide; membranous process stemming anteriorly from each prothoracic leg; body largest in region of thorax, distinctly narrowing toward apex of abdomen; anal prolegs united (pl. KI, 137)  Head round; prothoracic legs without membranous processes; body cylindrical, not sharply tapering toward apex; anal prolegs sepa-
3.	rated (pl. XI, 130-132)  Prolegs absent; each coxn of thoracic legs with three small tubercles; on Quercus, Betula (pl. XIV, 179; pl. XV, 181, 184, 189, 190; pl. XVI, 200)  Profesusa MacGillivray (pt.)  Prolegs present, although reduced; coxne without tubercles
4.	Prolegs present on abdominal segments 2 to 7 and 10; on Rubus (pl. XIII, 159-161)  Metallus Forbes
5.	Prolegs present on abdominal segments 2 to 8 and 10 Each proleg surrounded by well-defined crescentlike dark area (pl. XII, 152, 153; pl. XIV, 164)
6.	Prolegs without dark areas, or present only around anal proleg Mesosternum, metasternum, and sternum of first abdominal segment with dark plates; on Populus, Betula, Salix (pl. XII, 152, 153; pl. XIV, 164)  Mesosternum, metasternum, and sternum of first abdominal segment
	without plates; on Crataegus (pl. XVI, 201, 202)  Profenusa MacGillivray (pt.)
**	Don't proceentlike plate suprounding and prology on Patula Int. VII

7. Dark crescentlike plate surrounding anal proleg; on Betula (pl. XII,
143)

Heterarthrus Stephens
Anal proleg without dark plate

8

8. Mesosternum and metasternum without dark plates; on Platanus (pl. XVII, 212)

Bidigitus Smith Mesosternum and metasternum with light or dark plates; not on Platanus

9

9. First abdominal sternum without dark plate; on Alnus
Fenusa Leach (pt.)
First abdominal sternum with dark plate; not on Alnus
10

Ninth abdominal sternum with cluster of small spines; small spines about prolegs on segments 6 to 8 and at center of sterna 1 to 6; on Ulmus (pl. XVIII, 220-222) \_\_\_\_\_\_\_Fenusa Leach (pt.) Abdomen and prolegs smooth, without spines \_\_\_\_\_\_\_11

11. Plates on thoracic sterna and first abdominal sternum very dark, rectangular in shape, longer than wide; on Betula (pl. XVIII, 232, 233) Fenusa Leach (pt

232, 233) Fenusa Leach (pt.)
Plates lighter, that of prosternum wider than long, those of mesosternum, metasternum, and first abdominal sternum small; on
Potentilla (pl. XVII, 213, 214) Fenella Westwood

# Keys to Leaf-Mining Larvae of Betula and Quercus

#### BETULA

Prolegs absent; each coxa with three small tubercles on inner anterior surface; thoracic legs short, tarsal claw absent (pl. XV, 181, 182, 184)
 Profenusa thomsoni (Konow)
 Prolegs present, reduced; coxac without small tubercles; thoracic legs longer, distinctly segmented, tarsal claw present, sometimes reduced

2. Anal proleg without dark ring (pl. XVIII, 232)

Fenusa pusilla (Lepeletier)

Anal proleg with dark ring (pl. XII, 143, 153)
3. Each proleg with dark ring (pl. XII, 152, 153)
Only anal proleg with dark ring (pl. XII, 142, 143)
Heterarthrus nemoratus (Fallén)

Hering (1937) compared the birch leaf-mining species of Europe, and Lindquist (1959) gave a key to three birch leaf-mining species of Ontario.

#### QUERCUS

Mesosternum with dark plate (pl. XIV, 179)
 Profenusa inspirata (MacGillivray)
 Mesosternum without dark plate (pl. XV, 189)

2. Thoracic legs each with minute tarsal claw; antenna longer with distinct apical segment (pl. XVI, 199, 200) Profenusa lucifex (Ross) Tarsal claws absent; antenna without small apical segment (pl. XV, 190, 192) Profenusa alumna (MacGillivray)

# Tribe CALIROINI

The tribe Caliroini includes two genera, *Endelomyia* and *Caliroa*, the larvae of which are external feeders on the leaves of various trees and shrubs. The adults are recognized by the complete vein 2A and 3A of the forewing, connected to 1A by an oblique crossvein, the nine-segmented antenna, and the tarsal claw with a single outer tooth and a large acute basal lobe. The larvae may be distinguished as outlined in the introductory discussion on larvae.

#### Genus ENDELOMYIA Ashmead

Endelomyia Ashmead, 1898, p. 256; Konow, 1905, p. 74 (= Eriocampoides Konow); MacGillivray, 1909b, p. 346; Rohwer, 1911b, p. 100 (= Caliroa O. Costa); Ross, 1937, p. 70 (= Caliroa O. Costa); Benson, 1938, p. 368; Benson, 1940, p. 214; Ross, 1951, p. 28; Benson, 1952, p. 96; Malaise, 1957, p. 18; Malaise, 1963, p. 176; Smith, 1967, p. 279.
Type-species: "M. rosae Harris." Original designation (see discussion be-

low).

Description.—Second antennal segment longer than broad; third segment longer than fourth segment; segments beyond third gradually decreasing in length, apical four segments not reduced (pl. IV, 42). Clypeus truncate; malar space less than diameter of front ocellus. Prepectus absent. Tarsal claw with single outer tooth and large acute basal lobe (pl. III, 22). Forewing with vein 2A and 3A complete, connected to 1A by oblique crossvein; basal angle of vein 2A and 3A with short basally projecting spur (pl. I, 1). Hindwing with anal cell petiolate; cell M present, cell Rs absent (pl. I, 2).

A single Holarctic species is in this genus, aethiops (Fabricius), which is an external feeder in the larval stage on the foliage of

Rosa.

The type-species was designated as "M. rosae Harris" by Ashmead (1898). This species was originally described by Harris (1841) as Selandria rosae and is a synonym of aethiops.

#### Description of Endelomyia Species

Endelomyia aethiops (Fabricius)

Tenthredo aethiops Fabricius, 1781, p. 416; Fabricius, 1787, p. 256; Gmelin, 1790, p. 2662; Fabricius, 1793, p. 121; Fabricius, 1804, p. 39; Klug, 1814, p. 66; Klug, 1819, p. 84; Lepeletier, 1823, p. 112; Hartig, 1837, p. 268; Zetterstedt, 1838, p. 339; Ratzeburg, 1844, p. 130; Gorski, 1852, p. 191.

Allantus acthiops: Jurine, 1807, p. 56.

Hylotoma acthiops: Fallén, 1807, p. 209.

Nematus acthiops: Spinola, 1808, p. 155.

Phyllotoma acthiops: Fallén, 1829, p. 33.

Selandria acthiops: Stephens, 1835, p. 51; Westwood, 1850, p. 207; Kalten-

bach, 1867, p. 92; Kaltenbach, 1874, p. 221.

Blennocampa acthiops: Costa, 1859, p. 48; Kaltenbach, 1874, p. 200. Eriocampa aethiops: Cameron, 1876h, p. 192; Konow, 1886a, p. 109. Eriocampoides aethiops: Konow, 1890, p. 248; Dalla Torre, 1894, p. 192;

Eriocampoides aethiops: Konow, 1890, p. 248; Dana Torre, 1894, p. 182, Konow, 1905, p. 74.

Caliroa aethiops: Enslin, 1914, p. 253; Middleton, 1922, p. 12; Dovnar-Zapolsky, 1929, p. 15; Johnson, 1930, p. 93; Dovnar-Zapolsky, 1931, p. 55; Balachowsky and Mesnil, 1935, p. 234; Smith, Kelley, Dean, et al., 1943, p. 384; Hardouin, 1945, p. 161; Berland, 1947, p. 270; Schuh and Mote, 1948, p. 125; Dominguez Garcia-Tejero, 1950, p. 185; Tadic, 1956, p. 18.

Endelomyia aethiops: MacGillivray, 1909b, p. 346; Yuasa, 1922, p. 58; Miles, 1935, p. 126; Benson, 1940, p. 214; Ross, 1951, p. 28; Benson, 1952, p. 96; Maxwell, 1955, p. 54; Peterson, 1956, p. 268; Malaise, 1957, p. 18; Lorenz and Kraus, 1957, p. 113; Benson, 1962, p. 391; Smith, 1967, p. 279; Benson, 1968, p. 148.

Selandria rosae Harris, 1841, p. 380; Norton, 1861, p. 223; Norton, 1867, p. 256; Scudder, 1869, p. 268; LeBaron, 1871, p. 79; Riley, 1875, p. 27; Riley, 1877, p. 19; Provancher, 1878, p. 100; Provancher, 1880, p. 127; LeBaron, 1880, p. 66; Provancher, 1883, p. 202; MacGillivray, 1909b, p. 346 (= aethiops Fabricius).

Eriocampa rosae: Cameron, 1882, p. 227; Dalla Torre, 1894, p. 132.

Monostegia rosae: Provancher, 1888, p. 351; Riley, 1892, p. 6; Dyar, 1895a, p. 194; Dyar, 1895b, p. 338.

Endelomyia rosae: Ashmead, 1898, p. 256; Chittenden, 1908, p. 1. Eriocampoides rosae: Konow, 1905, p. 74; Gillette and List, 1915, p. 2.

The European synonymy for aethiops was given by Enslin (1914) and Benson (1952).

Female.—Average length, 4.6 mm. Black; extreme apex of each femur and all of each tibia and tarsus white to infuscate; back tibia and tarsus occasionally black. Wings lightly infuscate.

Sheath straight above, rounded below. Serrulae of lancet each lobelike, rounded at apex, with one anterior and one posterior subbasal tooth located near ventral margin of lancet (pl. IV, 62).

Male.—Unknown in North America. They are very rare in

Europe (Benson, 1952).

Larva.—Final feeding stage (pl. XI, 130-136). Length, 13 mm. Body cylindrical, typically sawflylike, not distinctly narrowing toward apex. Yellowish green when alive; tubercles concolorous

with rest of body.

Head rounded, hypognathous. Antenna conical, five-segmented. Clypeus with four setae; labrum truncate, with six setae; epipharynx with about 12 spines on each side; each mandible with ventral row of four long rectangular teeth; in addition, right mandible with one large truncate middle tooth and two sharp dorsal teeth and left mandible with one pointed middle tooth and three sharp dorsal teeth; one seta on basal angle of each mandible; maxillary palpus four-segmented, second segment with one seta, palpifer with three setae, galea conical, lacinia with 12 to 14 spines; labial palpus four-segmented.

Thoracic legs normal, five-segmented; tarsal claw present; each tibia with lateral pad; membranous process of prothoracic leg

absent. Tubercles of thorax arranged as in plate XI, 130.

Prolegs on abdominal segments 2 to 8 and 10. Abdominal segments 1 to 8 each with six annulets; annulets 2 and 4 each with two small conical tubercles on each side of body; ninth segment with row of six to eight tubercles dorsally on fourth annulet; dorsum of 10th segment with eight to 10 tubercles arranged approximately in three transverse rows. Suranal and subanal areas with setae. Spiracles not winged.

The larva was described by Dyar (1895a), Yuasa (1922), Miles (1935), Peterson (1956), and Lorenz and Kraus (1957), and the internal larval anatomy was described by Maxwell (1955). The distinct mandibles and tubercle arrangement of the body will

separate it from most other sawfly larvae.

Holotupes.—The location of the type of aethiops Fabricius is not known. Harris' type of rosae cannot be found.

Distribution.—Widespread in North America; Europe. The following data are limited to the State or Province and city or county of those specimens I have examined.2 British Columbia: Agassiz; Vancouver. Colorado: Boulder; Ft. Collins. Connecticut: Lyme; Branford. Delaware: Newark. District of Columbia: Washington. Idaho: Lenore. Illinois: Algonquin; Urbana; Champaigne; Rock Island; Alto Pass; Oakwood, Kansas: Yates Center; Douglas Co.; Manhattan. Kentucky: Lexington. Maine: Belfast. Massachusetts: Wellesley. Michigan: Detroit; East Lansing; LaPeer Co. Minnesota: Olmstead Co. Missouri: Jackson Co.; Kansas City; Columbia. Montana: Bozeman. New Jersey: N. Brunswick. New York: Oswego; Ithaca; Fulton Co. Ohio: Sugar Grove. Oregon: Troutdale; Salem; Forest Grove; Langdon Lake; Corvallis; Oakville. Pennsylvania: Philadelphia; Montgomery Co. Utah: Providence. Virginia: Falls Church; Vienna; Arlington; Bluemont. Washington: Colton: Woodland.

**Host.**—The larva is an external feeder on the foliage of wild and cultivated roses (*Rosa* spp.).

Biology.—Numerous papers have appeared on the life history of this species. The more noteworthy are those of Harris (1841), Riley (1892), Chittenden (1908), Middleton (1922) in North

America, and Miles (1935) in England.

The adults fly in May and June and oviposit in the edge of a leaf, usually in serrations. There may be from two to five eggs per leaf. Incubation is from 9 to 14 days. The young larvae feed on either side of the leaf, but the older larvae prefer the lower surface. The feeding period is from 20 to 27 days, and, on maturing, the larvae enter the soil to overwinter. There is one generation a year.

This is one of the three common sawflies that feed on the foliage of roses. The others—Cladius isomerus Norton and Allantus cinctus (L.)—are not treated in this bulletin. C. isomerus is univoltine and oviposits in the stems. The larva is distinctly hairy. A. cinctus has one or two generations a year. The larva has fewer and pointed mandibular teeth and smaller less conspicuous tubercles on the body than the larva of acthiops.

Discussion.—Rose-slug is the approved common name for this species. It has received much attention in the literature and was recognized as a pest of roses as early as 1841 by Harris. In Massachusetts in the 1840's it was such a pest that \$100 was offered for the most successful way to destroy it (Chittenden, 1908).

E. aethiops has sometimes been associated with the genus Caliroa, but the antenna and wing venation of the adult and the nonsluglike appearance of the larva separate it from species of Caliroa.

<sup>&</sup>lt;sup>2</sup> Throughout this bulletin all information pertaining to distribution records is given essentially as it appeared on the insect labels.

#### Genus CALIROA O. Costa

Caliroa O. Costa, 1859, p. 59; Dalla Torre, 1894, p. 192 (= Eriocampoides Konow); Ashmead, 1898, p. 256; Konow, 1905, p. 74 (= Eriocampoides Konow); MacGillivray, 1909b, p. 347; Rohwer, 1911a, p. 119; Rohwer, 1911b, p. 100; Enslin, 1914, p. 252; Yuasa, 1922, p. 58; Ross, 1937, p. 70; Benson, 1938, p. 368; Crevecoeur and Maréchal, 1938, p. 493; Benson, 1940, p. 214; Berland, 1947, p. 269; Ross, 1951, p. 28; Benson, 1952, p. 96; Takeuchi, 1952, p. 56; Malaise, 1957, p. 18; Lorenz and Kraus, 1957, p. 111; Malaise, 1961, p. 240; Malaise, 1963, p. 176; Okutani, 1965, p. 29; Smith, 1967, p. 279.

Type-species: Caliroa sebetia O. Costa. Monotypic.

Eriocampoides Konow, 1890, p. 239; Dalla Torre, 1894, p. 192; Ghigi, 1904, p. 4; Konow, 1905, p. 74; MacGillivray, 1909b, p. 347 (= Caliroa O. Costa).

Caliroa subgenus Eriocampoides: Rohwer, 1911b, p. 100; Enslin, 1914, p. 252. Type-species: Tenthredo limacina Retzius. Designated by MacGillivray, 1909b.

Perialistoptera Ashmead, 1898, p. 255; Konow, 1905, p. 74 (= Eriocampoides Konow); Enslin, 1914, p. 252 (= Caliroa O. Costa).

Type-species: Monostegia alba Norton. Original designation.

Description.—Second antennal segment longer than broad; third segment longer than fourth segment; apical four segments reduced, together subequal in length to or only slightly longer than third segment (pl. IV, 40, 41). Clypeus shallowly emarginate; malar space less than diameter of front ocellus. Prepectus absent. Tarsal claw with one outer tooth and large acute basal lobe (pl. III, 22). Forewing with vein 2A and 3A present for entire length, connected to 1A by oblique crossvein; basal angle of vein 2A and 3A without basally projecting spur. Hindwing with anal cell petiolate or sessile; cells Rs and M both present, both absent, or either one or the other present (pl. I, 3-5). Hindwing of male with peripheral vein (except cerasi) (pl. I, 6-7).

MacGillivray (1909b) treated 15 species of Caliroa from North America and gave a key to species that is based on variable and ambiguous characters and is impossible to use. Ross (1951) listed 15 species, but he did not express most of the synonymy. The present revision treats 14 species, five of which are new. Benson (1952) reported four species from England and Okutani (1965) seven species from Japan. There are probably about 30 world

species.

This revision is based largely on the female genitalia and hind-wing venation. Males have not been associated with some species, are less commonly collected, and their recognition characters are not so obvious as those of the female, although slight differences do exist in their genitalia, venation of the hindwing, and coloration. The relative lengths of the antennal segments and coloration of the tibiae and tarsi of the females sometimes have been used in the taxonomy of *Caliroa*; however, these characters are variable, vague, and difficult to see and describe, and they are not relied upon in this revision.

On the basis of the females, two groups of Caliroa may be recognized: (1) Those species that have the anal cell of the hindwing petiolate and cells Rs and M usually absent and (2) those species

that have the anal cell of the hindwing sessile and cells Rs and M either both present or at least one or the other present. These venational characters are one of the major divisions in the key to species. Occasionally abnormal specimens appear, and these should be taken through both parts of the key for correct determination.

Caliroa larvae are easily distinguished from larvae of other sawflies by their sluglike or tadpolelike appearance and by the presence of a fleshy protuberance extending anteriorly from each prothoracic leg (pl. XI, 137). They are usually covered with a dark or transparent slime. The head is hypognathous and the thorax is enlarged, with the body distinctly narrowing toward the posterior.

Larvae have not been associated with enough species of *Caliroa* to permit a key for their separation. Larval material is available for study from various hosts, most of it from *Prunus* and *Quercus*. However, at least two species have been associated with *Prunus* and five species with *Quercus*, and it is impossible to determine to which species the larvae belong. Also, a study of the larval material available indicates that there are few obvious characters for species separation. The most obvious character seems to be the color of the head capsule, but this color may change from amber to black in the various instars.

Dyar (1895b) attempted to separate the larvae of some species by using size and head capsule coloration, and Yuasa (1922) gave a key to five species, largely based on the same characters. However, the keys of these authors cannot be relied upon because the taxonomy of the adults was not well known at that time. Lorenz and Kraus (1957) gave a key to the larvae of four European species, which is also based on coloration. Consequently, I have not attempted a key because of the lack of associated specimens, lack of adequate characters, and the impracticality of a key leading only, for example, to "species A" or to "species B."

Caliroa includes the common pear-slug (C. cerasi), which feeds on a wide variety of Rosaceae in the larval stage but seems to prefer Prunus and Pyrus. Other Caliroa species are associated with Castanea, Nyssa, Prunus, Quercus, and Salix. Some adults have been collected from Betula, but this host is not yet authenticated. Some larvae have been taken while feeding on Acer, Ceanothus, and Cotoneaster, but the species are not known.

# Key to Caliroa Species

1.	Female Male	2 15
2.	Pronotum and mesonotum rufous floridana, n.	
	Thorax entirely black	3
3.	Hindlegs entirely black cerasi (Linnaeu	ıs)
	Tibia and tarsus of hindlegs partly white	4
4.	Anal cell of hindwing with short petiole; cells Rs and M usually ab-	
	sent (pl. I, 5)	5
	Anal cell of hindwing sessile; cells Rs and M both present or either	
	one or the other present (pl. I, 4)	10
5.	Serrulae of lancet deep, lobelike, rounded at apex (pl. V, 63, 67)	6
	Serrulae of lancet shallow or pointed at apex (pl. V, 64-66, 68)	7

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6.	Serrulae broad, close together (pl. V, 63); western distincta, n. sp. Serrulae slender, far apart (pl. V, 67); eastern lunata MacGillivray
~	Serrulae slender, far apart (pl. V, 67); eastern lunata MacGillivray
7.	Definition Standar, nointed at above $(n) = V - GA) \cdot weatown = -harding - n - nn$
	Serrulae broad, usually flattened at apex (pl. V, 65-66, 68); eastern
8.	Larget with 10 to 20 generals (-1 V 20)
ο.	species 8 Lancet with 19 to 20 serrulae (pl. V, 68) lorata MacGillivray
9.	Lancet with 15 to 17 serrulae 9 Serrulae flattened at apex, each with two anterior and two posterior
•	subbasal teeth (pl. V, 65) httrata MacGillivray
	Derruise rounded at above each with three or four small enterior and
	two or three large posterior subbasal teeth (pl. V. 66) veticlata, n sp
10.	Serrulae each with large distinct lobelike subbasal teeth (pl. V, 69;
	DL VI, 74) 11
	Serrulae each with small less conspicuous subbasal teeth (pl. VI. 72)
٠,	73; pt. VII, 75, 76)
ĭ ).	Serzulae usually with two anterior subbasal teeth (pl. V, 69);
	western labrata MacGillivray Serrulae usually with one anterior subbasal tooth (pl. VI, 74);
	eastern (Norton)
12.	eastern obsoleta (Norton) Lancet with 17 to 18 serrulae 13
13.	Serrulae of lancet evenly rounded with subbasal teeth extending to
	ventral margin of lancet (p), VII, 76); hindwing usually with calle
	Rs and M present (pl. I. 4) unercuscoccineae (Duan)
	Serrulae of lancet with anterior and posterior basal margins lacking
	subbasal teeth, subbasal teeth confined to rounded apex (pl. VI,
14.	72); hindwing usually with cell Rs present, cell M absent nyssae, n. sp.
14.	Serrulae deep, lobelike, rounded at apex (pl. VI, 73) lobata MacGillivray
	Serrulae small, shallow, more pointed at apex (pl. VII, 75)
15.	Hindwing without peripheral vein (males not seen; based on Benson
	Mindwing with peripheral vein (pl. I. G. 7)
16,	Abdomen rufous with central part of dorsum usually black
	lobata MacGillivray
17.	Abdomen entirely black
<b>1</b> 1.	Apex of anal cell of hindwing removed some distance from margin
	of wing, usually by at least half the width of anal cell at its widest point (pl. I, 7)
	Apex of anal cell close to or touching margin of wing (pl. I, 6)  Legs beyond coxae entirely whitish At least hindfemur black Wings hyaline; western Wings infuscated; eastern species  18 21 petiolata, n. sp. 19 hyalina, n. sp. 20
18.	Legs beyond coxae entirely whitish
	Legs beyond coxae entirely whitish At least hindfemur black  19
19.	Wings hyaline; western hyalina, n. sp.
20	Wings infuscated; eastern species 20
20.	Front and middle femora at least half white; harpe short, as broad
	AS 10DF UDL 1A. 1011! Smaller species literate MeaCillings.
	Front and middle femora black; harpe longer than broad (pl. IX, 98); larger species lorata MacGillivray
21.	98); larger species lorata MacGillivray Wings infuscated fasciata (Norton) and quarcuscoccinege (Dyor)
21,	Wings bending
22.	
	All formore with at least ! !
23.	Hindibia mostly black with only extreme apex white; harpe long,
	Stender (DL 1X, 99)
	Hindtibia with apical half white; harne short and broad (p) IX
	105) nyssae, n. sp.
	the second secon

# Descriptions of Caliroa Species

Caliroa cerasi (Linnaeus)

Tenthredo cerasi Linnacus, 1758, p. 557; Dalla Torre, 1894, p. 194 (lists

numerous references to this species under names "Tenthredo cerasi, Hylotoma cerasi, Phyllotoma cerasi, and Sclandria cerasi," all of which are questionable synonyms of limacina Retzius).

Taliroa cerasi: Rohwer, 1911a, p. 119; Webster, 1912a, p. 167; Webster, 1912b, p. 125; Wilson, 1913, p. 118; Ewing, 1917, p. 330; Britton, 1921, p. 199; Wellhouse, 1922, p. 1086; Yuasa, 1922, p. 59; Severin, 1923, p. 13; Zappe, 1926, p. 322; Obarski, 1933, p. 156; Benson, 1940, p. 214; Russo, 1943, p. 396; Hardouin, 1943, p. 160; Smith, 1943, p. 384; Schuh and Mote, 1948, p. 124; Ross, 1951, p. 28; Benson, 1952, p. 96; Maxwell, 1955, p. 55; Tadic, 1956, p. 18; Peterson, 1956, p. 268; Lorenz and Kraus, 1957, p. 112; Raizenne, 1957, p. 25; Benson, 1962, p. 391; Benson, 1968,

Eriocampoides cerasi: Cook, 1914, p. 40. Tenthredo limacina Retzius, 1783, p. 73.

Eriocampa limacina: André, 1881, p. 322; Cameron, 1882, p. 224; Froggatt,

Eriocampoides limacina: Konow, 1890, p. 248; Dalla Torre, 1894, p. 194; Marlatt, 1897, p. 1; Konow, 1905, p. 74; MacGillivray, 1909b, p. 347; Tullgren, 1910, p. 294; Rohwer, 1911a, p. 121; Jörgensen, 1913, p. 266; Camacho, 1917, p. 1; Tillyard, 1921, p. 2; Izquierdo, 1921, p. 43; Porter, 1928, p. 3; Porter, 1930a, p. 370; Porter, 1930b, p. 9; Forsius, 1931, p. 14; Twinn, 1934, p. 72; Jourdan and Rungs, 1935, p. 205; Talhouk, 1941, p. 128; Kotte, 1941, p. 165; Trujillo Peluffo, 1942, p. 275.

Caliroa limacina: Enslin, 1914, p. 254; Enslin, 1924, p. 37; Wolff, 1924, p. 45; Forsius, 1927, p. 6; Henn, 1929, p. 253; Dovynar, Zapolsky, 1929, p. 8

Tota limacina: Enstin, 1914, p. 254; Enstin, 1924, p. 37; Wolff, 1924, p. 45; Forsius, 1927, p. 6; Hepp, 1929, p. 253; Dovnar-Zapolsky, 1929, p. 83; Forsius, 1929, p. 83; Dovnar-Zapolsky, 1931, p. 55; Conde, 1934, p. 180; Beffa, 1934, p. 583; Balachowsky and Mesnil, 1935, p. 234; Miles, 1935, p. 117; Grandi, 1936, p. 224; Golfari, 1937, p. 241; Poluzzi, 1939, p. 525; Benson, 1940, p. 214; Chiesa Molinari, 1942, p. 463; Berland, 1947, p. 271; Dominguez Garcia-Tejero, 1950, p. 183; Scognamiglio, 1954, p. 96; Spirchez, 1956, p. 314; Iukhnevich, 1960, p. 18; Dadurian, 1962, p. 81.

Tenthredo cerasi Peck, 1799, p. 9.

Selandria cerasi: Harris, 1841, p. 383; Norton, 1861, p. 222; Winchell, 1865, p. 321; Norton, 1867, p. 254; Riley, 1875, p. 27; Thomas, 1881, p. 67; Forbes, 1883, p. 98; Packard, 1890, p. 522; MacGillivray, 1909b, p. 348 (= limacina Retzius).

Eriocampa cerasi: Dalla Torre, 1894, p. 130; Dyar, 1895a, p. 195; Dyar,

1895b, p. 338.

Eriocampoides cerasi: Konow, 1905, p. 74.

Caliroa landata MacGillivray, 1909b, p. 356, 9; Frison, 1927, p. 238; Ross, 1951, p. 29 (= cerasi Linnaeus).

Caliroa lacinata MacGillivray, 1909b, p. 357, 9; Frison, 1927, p. 238; Ross, 1951, p. 29 (= cere i Linnaeus).

Female.—Average length, 4.8 mm. Black with front and middle tibiae brownish. Wings lightly, uniformly infuscated.

Antenna with third segment subequal in length to segments 4 plus 5; apical four segments subequal in length to third segment. Clypeus shallowly, circularly emarginated. Hindwing with anal cell sessile; cells Rs and M usually both present, sometimes both absent or one or the other present. Lancet with about 17 serrulae, each serrula long, pointed at apex, with one anterior and three posterior subbasal teeth (pl. VI, 70).

Male.—Parthenogenetic, unknown in North America. Males are rare in Europe but apparently common in Turkey (Benson, 1968). Benson (1952) stated that they lack a peripheral vein in the hindwing.

Larra.—Final feeding stage. Length, 11 mm. May be separated from other known Caliroa larvae by the following combination of characters: (1) Host: Pyrus, Prunus, or other Rosaceae; (2) darkbrown head capsule in late feeding stages; and (3) black, rather than transparent, slime covering body. The general shape of the body, mandibles, epipharynx, and maxilla are as illustrated in

plate XI, 137-141.

The larva of cerasi has been described numerous times in the literature. Dyar (1895a, 1895b) and Yuasa (1922) attempted to separate it from other North American Caliroa larvae. Miles (1935), Peterson (1956), and Lorenz and Kraus (1957) all described and illustrated the larva, but Scognamiglio (1954) has given the most complete descriptive and morphological account of the larva. Maxwell (1955) described the internal larval anatomy.

Holotypes.—Malaise and Benson (1934) stated that Linnaeus' type of cerasi is lost. The location of Retzius' and Peck's types are not known. MacGillivray's types are at the Illinois Natural History Survey: C. landata, a female, is labeled "Vancouver, B.C., 19-VI-03," and C. lacinata, a female, is labeled "Algonquin, Ill., 6-8-94."

Distribution.—Widespread in temperate Eurasia and North America; also, Africa, Argentina, Australia, Chile, China, New Zealand, Tasmania, Uruguay. I have seen specimens from the following North American States and Provinces: British Columbia, California, Colorado, Connecticut, Delaware, District of Columbia, Georgia. Idaho, Illinois. Iowa, Maine, Massachusetts, Michigan, Minnesota, Montana, Nebraska, New Hampshire, New Jersey, New Mexico, New York, Ontario, Oregon, Pennsylvania, Quebec, Rhode Island, South Dakota, Utah, Virginia, Washington.

Hosts.—The larva is an external feeder on the foliage of many Rosaceae, preferably pear (Pyrus) and cherry (Prunus). Dyar (1895a) and Wellhouse (1922) reported that this species feeds on Crataegus, and labels on adult specimens indicate that Sorbus may be a host. Benson (1952) recorded Amygdalus, Crataegus, Cydonia, Mespilus, Quercus, Rosa, Rubas, Salix, and Sorbus as hosts of this species in England.

Biology.—The first reference to the life history of cerasi was as early as 1799 when Peck was awarded \$50 and a gold medal from the Massachusetts Agricultural Society for his printed account "Natural History of the Slug Worm." Since that time numerous papers have appeared, such as those by Harris (1841), Winchell (1865). Riley (1870), Thomas (1881), Forbes (1883), Marlatt (1897). Webster (1912a, 1912b), Wilson (1913), Ewing (1917), and Britton (1921). The insect has always received attention in reports of State entomologists. Outside of North America biological work has been carried on wherever this pest exists, such as in England (Miles, 1935), Lebanon (Talhouck, 1941), Morocco (Jourdan and Rungs, 1935), Chile (Porter, 1928, 1930a, 1930b; Izquierdo, 1921), Argentina (Jörgensen, 1913; Chiesa Molinari, 1942), Uruguay (Trujillo Peluffo, 1942), Australia (Froggatt, 1901), and New Zealand (Tillyard, 1921).

In North America two generations a year are usual, the first in May or June, the second in July or August. The eggs are inserted on the underside of the leaf near a midrib or larger vein. The feeding period may extend to 3 weeks, after which the larva enters the ground to pupate for the next generation or to overwinter. Slight differences are found in the biology of *cerasi* and *liturata*, another species associated with *Prunus*. (See section on biology under *liturata*.)

**Discussion.**—C. cerasi is easily separated from all other Caliroa species by the entirely black legs. All other species, except floridana, which has a rufous pronotum and mesonotum, have some white markings on the hindtibia and hindbasitarsus. The lancet with the relatively slender serrulae, each with one anterior and three posterior subbasal teeth, is also distinctive for this species.

Pear-slug is the approved common name.

#### Caliroa distincta, new species

Female.—Length, 4.8 mm. Black; front and middle tarsi, front and middle tibiae, and extreme apex of front and middle femora brownish to white; basal one-third of hindtibia and basal one-third of hindbasitarsus white. Wings very lightly, uniformly, infuscated.

Antenna with third segment subequal in length to segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus circularly emarginated. Hindwing with cells Rs and M absent; anal cell with short petiole. Lancet with 16 to 17 serrulae; serrulae deep, close together, and lobelike, each rounded at apex and with three or four anterior and three or four posterior subbasal teeth located near ventral margin of lancet (pl. V, 63).

Male.—Unknown.

Larva.---Unknown.

Holotype.—Female, 7 mi. N.W. Roseburg, Oregon, Douglas Co., May 8, 1964, J. D. Vertrees, U.S.N.M. (U.S. National Museum) type No. 70212.

Paratypes.—British Columbia: Cultus Lake, 21-VII-1948, H. R. Foxlee (1 9); Salmon Arm, 10-VII-1935, A. A. Dennys (1 9); Robson, 9-V-1947, H. R. Foxlee (1 9). California: Sta. Cruz Mts. (2 99); nr. Stanford U., Bear Creek Gulch, March 25, 1915, Harold Morrison (19). Nevada: "Nev." (19). Oregon: "Oregon," Koebele (19). Talent, May 2, 1936, L. G. Gentner (19); Douglas Co., 4 mi. S. Canyonville, May 21, 1964, D. R. Smith (19). Deposited in the Canadian National Collection, U.S. National Museum, and at Oregon State University.

**Distribution.**—Pacific coastal region from British Columbia to California and Nevada.

Host .-- Unknown.

Biology.-Unknown.

**Discussion.**—This western species is recognized by the petiolate anal cell and absence of cells Rs and M in the hindwing and the large lobelike serrulae of the lancet. The lancet is similar to that of lobata of eastern North America, but in distincta the serrulae are larger, closer together, and fewer.

#### Caliroa fasciata (Norton)

Selandria fasciata Norton, 1864, p. 9, 9; Norton, 1867, p. 256; Cresson, 1928, p. 6.

Eriocampa fasciata: Dalla Torre, 1894, p. 130. Eriocampoides fasciata: Konow, 1905, p. 74.

Caliroa fasciata: MacGillivray, 1909b, p. 362; MacGillivray, 1916, p. 80; Ross, 1951, p. 29; Maxwell, 1955, p. 55 (?).

Female.—Average length, 4.8 mm. Black; front and middle tibiae and tarsi white; basal two-thirds of hindtibia and basal two-thirds of hindbasitarsus white. Wings moderately to darkly infuscated on basal two-thirds, nearly hyaline on apical one-third.

Antenna with third segment shorter than segments 4 plus 5; apical four segments together slightly shorter than third segment. Clypeus shallowly, circularly emarginated. Hindwing with cells Rs and M both present; anal cell sessile. Lancet with 20 to 22 serrulae; each serrula shallow and broad, rounded at apex, with one or two anterior and three or four posterior subbasal teeth (pl. VII, 75).

Male.—Average length, 4.5 mm. Color similar to that of female except for extreme apex of each femur, which is white, and hind-tibia, which is only one-half white. Hindwing with peripheral vein; anal cell with apex close to margin of wing (pl. I, 6). Genitalia as in plate IX, 103 and 104; harpe longer than broad, pointed at apex.

Larva.—Unknown. Maxwell (1955) described the internal anatomy of a species of Caliroa on Crataegus, which she called fasciata.

*Holotype.*—S. fasciata Norton, a female labeled "Mass.," is type No. 10340 at the Academy of Natural Sciences of Philadelphia.

Distribution.—Eastern North America. Connecticut: Union, June 24, 1932; Poquonock, June 27, 1905; E. Hartford, Aug. 25, 1950. Illinois: Hardin, June 5–9, 1932; Parker, June 4, 1913; Jeff, June 12, 1934. Iowa: Mt. Pleasant, May 1, 1933, on pin oak. Maine: Brunswick, Mare Pt., 1–VII–65, ex w. birch; Augusta, VIII–10, 1941; Woolwich, emgd. VI–6, 7, VII–10, 14, 16, 1958, ex red oak. Maryland: Plummers Id., July 6, 1921, June 18, 1916, VIII–3–1912; Glen Echo. New Jersey: Ateo; Manahawkin, V–30–11. New York: Ardsley, Westchester Co., May 30, 1957. Ohio: Put-in-Bay, June 20–30, 1924. Ontario: Little Rapids, 3–V–1955, red oak; Nestorville, 18–IV–1955, red oak. Virginia: Veitch, July 7, 1914, Quercus velutina: Falls Church, July 23, 1913, reared VII–7–1916, Quercus palustris.

Host.—Quercus spp. Series of adults have been reared from larvae feeding on Quercus velutina Lam., Q. palustris Muenchh., and Q. rubra L.

Biology.-No information is available.

**Discussion.**—This species is most closely related to quercuscoccineae; however, quercuscoccineae has only 17 or 18 serrulae on the lancet and each serrula has several more anterior subbasal teeth. I could not separate the males of these two species. The presence of cells Rs and M and sessile anal cell of the hindwing and the rela-

tively shallow serrulae of the lancet will separate the females from those of other *Caliroa* species. The wings, which are only partly infuscate, will also help to differentiate this species. The males are more difficult to separate from those of other species. The closeness of the anal cell to the margin of the hindwing, entirely black body, and partly infuscate wings will help to separate them.

#### Caliroa floridana, new species

Female.—Length, 4.8 mm. Head and antenna black; labrum brownish. Thorax black with pronotum, tegula, and mesonotum rufous. Legs black with front tibia and tarsus whitish. Abdomen

black. Wings uniformly, darkly infuscated.

Antenna with third segment shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus shallowly, circularly emarginated. Hindwing with cells Rs and M both absent; and cell with very short petiole. Lancet with 20 to 22 serrulae; each serrula relatively deep, rounded at apex, and with two or three anterior and two or three posterior subbasal teeth (pl. VI, 71).

Male.—Unknown. Larva.—Unknown.

Holotype.--Female, Hillsboro Co., Fla., 3-24-30, C. L. Rabb,

Florida Fruit Fly Trap Survey. U.S.N.M. type No. 70213.

Paratypes.—Florida: Sumter Co., 4-24-30, W. A. Bryant, Florida Fruit Fly Trap Survey (1 ?); LaBelle, IV-21-21, J. N. Knull (1 ?). Deposited in the U.S. National Museum.

Distribution.—Known only from Florida.

Host .-- Unknown.

Biology.--Unknown.

Discussion.—The red pronotum and mesonotum and black hindlegs will separate this species from all other Caliroa species. The lancet is very similar to that of fasciata except for several more anterior subbasal teeth on the serrulae of the lancet of floridana. I have not seen specimens that may indicate intermediate color variations between this and other species and am therefore considering this species distinct.

# Caliroa hyalina, new species

Female.—Length, 4.8 mm. Black; front and middle tibiae and tarsi, basal one-third of hindtibia, and basal one-third of hind-

basitarsus white. Wings hyaline.

Antenna with third segment subequal in length to segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus circularly emarginated. Hindwing with cells Rs and M absent; anal cell with very short petiole. Lancet with 16 to 17 serrulae; serrulae far apart, each serrula slender, pointed at apex, with one anterior and two posterior subbasal teeth (pl. V, 64).

Male.—Length, 4.6 mm. In color and structure similar to female. Hindwing with peripheral vein; anal cell with apex removed from

margin of wing (pl. I, 7). Genitalia as in plate IX, 96; harpe only slightly longer than broad.

Larva.—Unknown.

Holotype.—Female, Salem, Oreg., VII-10-18, on cherry, E. J. Newcomer, collector, Quaintance No. 14085. U.S.N.M. type No. 70214.

Paratypes.—Oregon: Mt. Hood (1 9, 3); same data as for holotype (1 9); "Oregon," Koebele (2 9 9, 1 3); Klamath I'alls, June 6, 1956, Joe Schuh, coll. (1 9). Deposited in the U.S. National Museum and at Oregon State University.

Distribution.—Known only from Oregon.
Host.—Adults were collected from cherry.

Biology.—No information is available.

Discussion.—This species is similar to cerasi, but may be separated by the more hyaline wings, lack of cells Rs and M in the hindwing, the petiolate anal cell of the hindwing, and the widely separated serrulae of the lancet, each serrula with fewer subbasal teeth. The venation of the hindwing and lancet characters will also separate hyalina from other Caliroa species.

#### Caliroa labrata MacGillivray

Caliroa labrata MacGillivray, 1909b, p. 360, &; Frison, 1927, p. 238; Ross, 1951, p. 29.

Female.—Average length, 4.8 mm. Black; front tibia and tarsus, basal one-half of middle tibia, and basal one-third of hindtibia white. Wings uniformly, lightly infuscated.

Antenna with third sigment subequal in length to or slightly shorter than segments 4 plus 5; apical four segments together longer than segment 3. Clypeus circularly emarginated. Hindwing with anal cell sessile; cells Rs and M both present. Lancet with 16 to 17 serrulae; each serrula shallow, broad, with large lobate subbasal teeth, usually two large posterior and two large anterior subbasal teeth (pl. V, 69).

Male.—Average length, 4.5 mm. Black with extreme apex of front and middle femora, all of front and middle tibiae and tarsi, and basal one-half of hindtibia white. Hindwing with peripheral vein; apex of anal cell close to margin of wing (pl. I, 6). Genitalia as in plate IX, 97; harpe longer than broad, apex and inner margin rounded.

Larva.—I have seen one small larva, probably an early instar, taken from Salix. The head capsule and thoracic legs are dark brown.

Holotype.—At the Illinois Natural History Survey, a male labeled "Mts. near Claremont, Calif., Baker."

Distribution.—Western North America. Alberta: Jumping Pd. Cr., 20 mi. W. Calgary, July 3, 1962. British Columbia: Remo, June 13, 1960. California: Mt. Shasta, Siskiyou Co., 3561', em. 9 Oct. 1965, 30 Sept. 1965, leaf slug, no cocoon, Salix lemmonii; 8 mi. S. Bieber, Lassen Co., VIII-3-1965. Nevada: Galena Cr., Campground, 7 mi. S.W. Reno, Washoe Co., 6100', 27-VI-1965, 11-

VII-1965, Salix lasiolepis; Douglas Co., east end Kingsbury Grade, 3 mi. S. Genoa, 4800', 10-VII-1965, em. 7, 8-VIII-1965, Salix lasiolepis. Oregon: Corvallis, August 8, 1962.

Host.—Adults have been bred from larvae feeding on Salix

spp.

Biology.—No information is available.

Discussion.—The sessile anal cell and presence of cells Rs and M in the hindwing and the large lobate subbasal teeth of the serrulae of the lancet distinguish labrata from other Caliroa species. It is most closely related to obsoleta, but labrata may be separated by the more darkly infuscated wings and the serrulae, which usually have an extra anterior and posterior subbasal lobe. Also, the host of obsoleta is Quercus and that of labrata is Salix. This is the only known North American Caliroa species associated with Salix.

#### Caliroa liturata MacGillivray

"3S" (in Key), Dyar, 1895b, p. 338. Caliroa obsoleta Dyar, nee Norton, 1896, p. 237.

Caliron liturata MacGillivray, 1909b, p. 349, 9; Frison, 1927, p. 238; Ross, 1951, p. 29.

Caliroa lineata MacGillivray, 1909b, p. 350, 9; Frison, 1927, p. 238; Ross, 1951, p. 29. New synonymy.

Caliroa loricata MacGillvray, 1909b, p. 351, Q; Frison, 1927, p. 239; Ross,

1951, p. 29. New synonymy.
Caliroa (Eriocampoides) amygdalina Rohwer, 1911c, p. 263, &. &; Cushman, 1911, p. 91; Ross, 1951, p. 28; Burks, 1967, p. 16. New synonymy.

Female.—Average length, 4.5 mm. Black; front and middle tibiae and tarsi, basal two-thirds of hindtibia, and basal one-half of hindbasitarsus white. Wings uniformly, lightly infuscated,

sometimes slightly darker basally.

Antenna with third segment subequal in length to or only slightly shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus circularly emarginated. Hindwing with cells Rs and M absent; anal cell with short petiole. Lancet with 17 to 18 serrulae; each serrula low, flattened at apex, and with two anterior and two posterior subbasal teeth, flattened apex sometimes with several teeth, and apical serrulae usually with only one anterior and one posterior subbasal tooth (pl. V, 65).

Male .-- Average length, 4.3 mm. Black; front and middle tibiae and tarsi, apical half of each femur, and basal half of hindtibia and hindbasitarsus whitish. Hindwing with peripheral vein; apex of anal cell removed some distance from margin of wing (pl. I, 7). Genitalia as in plate IX, 101 and 102; harpe slightly longer than broad.

Larva.—The larva was described by Dyar (1896) under the name "obsoleta" and by Cushman (1911) under the name "amygdalina." The following may serve to distinguish the larva: (1) Feeds on *Prunus* spp. and (2) the head is pale brown, as opposed to dark brown for cerasi, which also feeds on Prunus.

Holotypes.-MacGillivray's types, all females, are at the Illinois Natural History Survey and are labeled as follows: C. liturata. "Florida"; C. lineata, "Columbia, Mo., 15 July 1905"; C. loricata,

"Columbia, Mo., 2 Sept. 1905." C. amygdalina Rohwer, a female, is U.S.N.M. type No. 13371, labeled "Hunter No. 1936, II-15, 7-24-

10, bred peach, Tallulah, La., 7-13-10."

Distribution.—Widespread in eastern North America. Colorado: Boulder, 5500', June 9, 11, 19, 21, 1961, 6000', June 4, 1961. Connecticut: Lyme, VI-19-18; Windsor, VIII-20-1951. Florida: "Florida:" Georgia: Austell, VIII-27-10. Illinois: McLean, June 25, 1940; Dubois, May 21, 1917. Louisiana: Baton Rouge, May 1, 1896, Sept. 9, 1922, bred from peach leaves, adult, Sept. 28; Tallulah, 8-22-10, 8-12-10, on peach, August 18, 1909. on peach. Missouri: Columbia, July 15, 1905, Sept. 2, 1905. Montana: "Montana." New York: Ithaca, July 4, 1918; Buffalo, 6-25-10; Keene Valley, on plum (Dyar's "3S"). Rhode Island: Kingston, June 20, 1905. South Carolina: Clemson Co., May 31, 1895. Tennessee: Rhea Co., VI-14-1939, peach orchard. Virginia: Falls Church, June 9, 1921, July 20, 1917; Suffolk, June 11, 1895; Vienna, VI-12-1915, bred from plum.

Hosts.—This species has been bred from peach and plum (Prunus spp.). Dyar (1896) bred it from "Prunus serotina" and

"P. pennsylvanica."

Biology.—Cushman (1911) reported on the biology of this species. The first adults were found in April in Louisiana and were present until cold weather in the fall, usually after six or seven generations. Oviposition is on the upper side of the leaf close to the midrib or a larger vein. This is opposed to cerasi, which usually oviposits on the underside of the leaf. The liturata larvae feed for about 3 weeks and drop to the ground to pupate. The total life cycle takes about 20 to 30 days. Another difference between the two species is that the liturata larvae do not eat the exuvium whereas the cerasi larvae do.

Discussion.—Attention was given to this species in 1910 when it was discovered damaging peach trees in Louisiana. Rohwer (1911c), believing it was a new species, described it as amygdalina; however. MacGillivrav (1909b) had described it as liturata 2 years before. The petiolate anal ceil and absence of cells Rs and M in the hindwing and short lancet with the servulae shallow and truncate at their apices will separate the females from those of other Caliroa species. The males are more difficult to identify, but the anal cell of the hindwing does not reach the margin of the wing, the wings are infuscated, and the front and middle femora are partly white. Generally this is a smaller species than most other Caliroa species.

# Caliroa lobata MacGillivray

Caliron lobata MacGillivray, 1909b, p. 355, 9; Frison, 1927, p. 239; Ross, 1951, p. 29.

Female.—Average length. 4.8 mm. Black; extreme apex of each femur, all middle and front tibiae and tarsi, basal three-fourths of hindbasitarsus white.

Wings uniformly, moderately infuscated, sometimes lighter on

apical one-third.

Antenna with third segment shorter than segments 4 plus 5; apical four segments subequal in length to third segment. Clypeus shallowly, circularly emarginated. Hindwing with cells Rs and M both present; anal cell sessile. Lancet with 20 to 21 serrulae; each serrula deep, symmetrical, lobelike, with apex rounded and three or four anterior and three or four posterior subbasal teeth located near ventral margin of lancet (pl. VI, 73).

Male.—Average length, 4.5 mm. Antenna, head, and thorax black. Each leg entirely whitish beyond coxa except apical one-third of hindtibia and hindbasitarsus, which are black. Abdomen dark rufous with central part of each terga and hypandrium black. Hindwing with peripheral vein; apex of anal cell near margin of

vein (pl. I, 6). Genitalia as in plate IX, 107.

Larra.-Unknown.

Holotype.—At the Illinois Natural History Survey, a female,

labeled "Oswego, N.Y., July 25, 1895."

Distribution.—Eastern North America. Illinois: "N. Ill."; Muncie, IX-16; Havana, July 28, 1910. Iowa: County No. 16. June 19, 1918. Maryland: Plummers Id., Aug. 12, 1914, IX-13-1958, VIII-5-1914. Michigan: Marquette. New York: L.I.; Oswego, July 25, 1895; Ithaca, June 19, 1908. Texas: College Station, April 19, 1932. Vermont: Rutland, Aug. 1-15, 1916. Virginia: Falls Church, July 25, Aug. 13, 16, 22, Sept. 12, 1927, reared, Quercus, Aug. 28, 1912. Quercus velntina, VII-25-1916, Quercus palustris, May 20, 1918, IX-11-20, IX-19, 1917.

Host.—Quereus spp. Adults have been bred from larvae feeding

on Quercus palustris Muenchh, and Q. velutina Lam.

Biology.—No information is available.

Discussion.—The sessile anal cell and presence of cells Rs and M of the hindwing and the large lobate serrulae of the lancet will distinguish the female of the species. The males may be separated by the rufous abdomen, which is black in all other known Caliroa males.

# Caliroa lorata MacGillivray

Caliroa larata MacGillivray, 1909b, p. 355, 9; Frison, 1927, p. 239; Ross, 1951, p. 29.

Eriocampoides castaneae Rohwer, 1917, p. 152, 9. New synonymy. Caliroa castaneae: Ross, 1951, p. 29.

Cuttion custaneae. Ross, 1991, p. 29.

Female.—Average length, 4.8 mm. Black; front and middle tibiae and tarsi, basal one-half of hindtibia, and basal one-half of hindbasitarsus white. Wings uniformly, lightly, infuscated.

Antenna with third segment slightly shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus circularly emarginated. Hindwing with cells Rs and M both absent; anal cell with short petiole. Lancet with 19 to 21 serrulae; serrulae shallow, not extending far below margin of lancet, each serrula with anterior margin more or less straight,

with one anterior subbasal tooth, posterior margin oblique, with two or three subbasal teeth (pl. V, 68).

Male.—Average length, 4.5 mm. Color similar to that of female. Hindwing with peripheral vein; apex of anal cell removed short distance from margin of wing (pl. I, 7). Genitalia similar to plate IX, 97; harpe at least twice as long as broad.

Larva.—Unknown.

Holotypes.-C. lorata MacGillivray, a female, is at the Illinois Natural History Survey with the data "Mt. Tom, Mass., July 16, '98." E. castaneae Rohwer, a female, is U.S.N.M. type No. 15486 with the data "11307a Hopk, U.S., reared, Aug. 5, '12, Castanea

dentata, Falls Church, Va., Wm. Middleton."

Distribution.—Eastern North America. Alabama: Pyziton, Clay Co. Maine: Winslow, Kennebec Co., June 30, 1957. Maryland: Plummers Id., 20-VI-1912, 8-26-1960; Glen Echo; 3 mi. S.E. Beltsville, May 30, 1966. Massachusetts: Mt. Tom, July 16, '98. New York: Ithaca. North Carolina: "N.C." Pennsylvania: Wernersville, VIII-5-1913; 5 mi. W. Davidsburg, VI-16-1958; Harrisburg, VI-17. Virginia: Falls Church, reared Aug. 5, 1912, Castanea dentata, VI-19-13, Castanea dentata, July 16, 1913, VIII-14-1913, Castanca pumila; Black Pond, Fairfax Co., June 19, 1919; Glencarlyn, V-26; Barcroft, VI-5-11; Vienna, reared, Aug. 13, 1912, Castanca dentata, Wisconsin: Gays Mills, 7-2-1930.

Host .- Adults have been bred from larvae feeding on Castanea spp.

Biology.—No information is available.

Discussion.—The petiolate anal cell and absence of cells Rs and M in the hindwing and the low serrulae of the lancet will distinguish the females of this species. The males may be separated by the apex of the anal cell of the hindwing, which is removed from the margin of the wing, the infuscated wings, and the black femora.

# Caliroa lunata MacGillivray

Caliroa lunata MacGillivray, 1909b, p. 353, 9; Frison, 1927, p. 239; Ross, 1951, p. 29. Caliron lata MacGillivray, 1909b, p. 361, 9; Frison, 1927, p. 238; Ross, 1951,

p. 29. New synonymy.

Female.—Average length, 4.5 mm. Black; front and middle tibiae and tarsi, basal one-half of hindtibia, and basal one-half of hindbasitarsus white. Wings lightly infuscated.

Antenna with third segment slightly shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus circularly emarginated. Hindwing with cells Rs and M both absent; anal cell with short petiole. Lancet with 17 to 19 serrulae; serrulae far apart, each long, slender, symmetrical, with apex rounded and two or three anterior and two or three posterior subbasal teeth located near ventral margin of lancet (pl. V, 67).

Male.--Unknown.

Larva.---Unknown.

Holotypes.—MacGillivray's types are at the Illinois Natural History Survey. C. lunata, a female, is labeled "Ithaca, N.Y., 27 May '93," and C. lata, a female, is labeled "Ithaca, N.Y., 22 July '90."

Distribution.—Eastern North America. Illinois: Shawneetown, June 14, 1934; Kampsville, June 10, 1932; Vienna, July 10, 1935; Oakdale, 8-3-35; Riverside Woods, Cook Co., 9-3-1949. Michigan: "Mich." Newfoundland: 3 mi. S.E. St. George's, June 27, 1966. New Hampshire: Notchland, Carroll Co., June 12, 1967. New York: Ithaca, May 27, '93, July 22, '90; Babylon, L.I., VII-13-1937; New Balt., 8-11-93. Ohio: Columbus, 05-10-1902. Pennsylvania: Harrisburg, VI-22. Texas: Liberty, III-18-08. Virginia: Rosslyn. West Virginia: Lost River St. Pk., Hardy Co., Aug. 1-14, 1960.

Host .- Unknown.

Biology.—Unknown.

**Discussion.**—This species may be recognized by the absence of cells Rs and M and the petiolate anal cell of the hindwing and the long slender lobelike serrulae of the lancet. The lancet resembles that of *lobata* and *distincta*, but in *lunata* the serrulae are more slender and farther apart. Also, there are fewer serrulae in the lancet of *lunata* than in that of *lobata*.

#### Caliroa nyssae, new species

Female.—Length, 4.8 mm. Black; extreme apices of front and middle femora, all front and middle tibiae and tarsi, basal one-half of hindtibia, and basal two-thirds of hindbasitarsus white. Wings hyaline.

Antenna with third segment slightly shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus circularly emarginated. Hindwing with anal cell sessile; cell Rs present, cell M absent. Lancet with 17 to 18 serrulae; each serrula low, broad, anterior and posterior margins smooth, without subbasal teeth, about five or six teeth present on apical rounded part (pl. VI, 72).

*Male.*—Length, 4.5 mm. In color and structure similar to female. Hindwing with peripheral vein; apex of anal cell close to margin of wing (pl. I, 6). Genitalia as in plate IX, 105 and 106; harpe slightly longer than broad.

Larva.--- Unknown.

Holotype.—Female, Falls Church, Va., reared, Aug. 5, 1912, Nyssa sylvatica, Wm. Middleton, collector, Hopk. U.S. 11308a. U.S.N.M. type No. 70215.

Allotype.—Male, same data as for holotype. Deposited with holotype.

Paratypes.—Georgia: Dalton, Aug. 29, 1909 (1 ?); Atlanta, 8-6-41, P. W. Fattig (1 ?). Maryland: Glen Echo, R. M. Fouts (1 ?). Mississippi: Biloxi, Sept. 29, 1908, black gum, J. Brodei (1 ?): Biloxi, VIII-31, 1908, issued IX-14-1908, on black gum (1 ?). Pennsylvania: Lakemont Park, Blair Co., 8-8-1947 (1 ?). Virginia: Data as for holotype (1 &); Great Falls, reared May 26,

1914, Nyssa, Heinrich, colr., Hopk. U.S. 11398e (2 9 9); Kearney, reared, Sept. 9, 1912, Nyssa sp. ?, Wm. Middleton, colr., Hopk. U.S. 11317d (1 9); Falls Church, IX-5-17, G. M. Greene (1 9); Falls Church, July 24, 1920, Wm. Middleton, colr. (1 2). Deposited in the U.S. National Museum and at the Illinois Natural History Survey.

Distribution.—Eastern North America.

Host.—Adults have been reared from larvae feeding on Nyssa sulvatica Marsh.

Biology.—No information is available. Collection dates on specimens indicate this species flies in late summer, usually in August.

Discussion.—The sessile anal cell, presence of cell Rs and absence of cell M in the hindwing, hyaline wings, and lancet will separate females of this species. The hindwing venation is unique and is very consistent in this species; however, the lancet should always be checked before final determination. The closeness of the anal cell to the margin of the hindwing, the hyaline wings, the partly white femora, and short broad harpe of the genitalia will distinguish males of this species.

#### Caliroa obsoleta (Norton)

Selandria obsoleta Norton, 1867, p. 254; Provancher, 1878, p. 100; Provancher,

1883, p. 202; Cresson, 1928, p. 8. Caliroa obsoleta: Kirby, 1882, p. 182; Provancher, 1888, p. 351 (?); Yuasa, 1922, p. 59; Ross, 1951, p. 29.

Eriocampoides obsoleta: Dalla Torre, 1894, p. 196; Konow, 1905, p. 74. Selandria quercus alba Norton, 1867, p. 258, 9; Norton, 1872, p. 85; Packard,

1890, p. 205. New synonymy.

Monostegia querous alba: Kirby, 1882, p. 186; Dyar, 1894, p. 43. Eriocampa querous alba: Dalla Torre, 1894, p. 132.

Selandria quercus-alba Cresson, 1880, p. 60. Correction of quercus alba Norton.

Caliroa quercus-alba: Yuasa, 1922, p. 59; Ross, 1951, p. 29; Raizenne, 1957, p. 25 (?).

Eriocampoides quercus Konow, 1905, p. 74. Correction of quercus alba Norton.

Female.—Average length, 4.5 mm. Black; extreme apex of each femur, all front and middle tibiae and tarsi, basal two-thirds of hindtibia, and basal two-thirds of hindbasitarsus white. Wings uniformly hyaline.

Antenna with third segment slightly shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus shallowly, circularly emarginated. Hindwing with anal cell sessile; cells Rs and M usually both present, but either one or both may be absent. Lancet with 18 to 20 serrulae; each serrula shallow, with large lobelike subbasal teeth, one on anterior margin and two on posterior margin, all distinct and subequal in size (pl. VI, 74).

Male.—Average length, 4.3 mm. Color similar to that of female except for each femur, which is whitish. Hindwing with peripheral

vein; apex of anal cell close to margin of wing (pl. I, 6). Genitalia

as in plate IX, 99; harpe longer than broad.

Larva.—Dyar (1894) described the larva and stated that the head capsule is brownish black. I have not seen the larvae, and Dyar's description does not adequately characterize the larva of this species.

Holotypes.—S. obsoleta Norton, a female, labeled "Mass." is type No. 10717 at the Academy of Natural Sciences of Philadelphia.

I could not find the type of quercus alba Norton.

Distribution.—Eastern North America. Connecticut: Lyme, reared, VI-6-16, Quercus alba, July 8, 1918; Stamford, Aug. 17, 1931, reared, Aug. 4, 1931, swamp white oak; East River, 8-20-23, 1914, white oak leaf, reared Aug. 12, 1916, white oak; E. Hartford, Aug. 25, 1950; Stafford, VIII-24-1950. Georgia: Athens, 5-23-37. Illinois: Oakwood, June 2, 1927; Perry Springs, Aug. 11, 1948, on white oak; White Pines St. Pk., May 30, 1936; Fox Lake, June 10, 1936; White Pine Forest, Ogle Co., July 4, 1932; Grand Tower, June 2, 1913; Ziegler, Aug. 21, 1933; Herod, June 23, 1927. Iowa: Mt. Pleasant, May 20, 1930. Massachusetts: Southwick, VIII-29-1916; Woods Holl, on white oak (Dyar's "J"), Michigan: Douglas Lake, July, 1914. Minnesota: "Minn." New Jersey: Ateo. New York: Napeague, L.I., VII-2-1954; Bedford. Westchester Co., July 6, 1907; Ithaca, Aug. 12, 1918; Ft. Montgomery, Aug. 6, 1923. Ontario: One Sided Lake, June 30, 1960. Virginia: Falls Church, reared, Aug. 28, 1912, Quercus alba, June 27, 1913, reared July 29, 1912, Quercus prinus, reared July 29, 1912, Quercus alba, July 31, 1916, Quercus minor; Glencarlyn, June 5, 1918, Wisconsin: Madison, 8-10-1929; Colona, June 19, 1934.

Host.—Reared from Quercus spp. Apparently most common on white oak (Quercus alba L.) but also reared from Q. stellata Warg. (=Q. minor Sarg.) and Q. prinus L. Dyar (1894) reared this

species from white oak; the reared adults are labeled "J."

Biology.—The only information on the biology is that by Dyar (1894). The adults oviposit on the underside of the leaf "near the tip." After feeding, the larvae drop to the ground and "form a

cavity in the earth, lined with a brown secretion."

Discussion.—The large distinct subbasal teeth of the lancet and the sessile anal cell of the hindwing will separate this species. Cells Rs and M of the hindwing are normally present, but some specimens, including the type of obsoleta, lack both cells: in other specimens either one or the other cell may be absent. The wings are usually hyaline, but some specimens from more northern localities have lightly infuscated wings. The lancet is close to that of labrata, but there are more subbasal teeth on the serrulae of labrata and its host is Salix.

Norton's type of *quercus alba* is apparently lost. Its synonymy with *obsoleta*, however, seems justified, since it was apparently bred from white oak, a common host for this species, and, according to Norton's (1867) original description, the wings of *quercus* 

alba are hyaline and cells Rs and M are both present in the hind-wing, both of which are usual characters for this species.

#### Caliroa petiolata, new species

Caliroa lineata of auctt., nec MacGillivray, Burks, 1958, p. 12; Burks, 1967, p. 16.

Female.—Length, 4.5 mm. Black; each front and middle tibia and tarsus, basal two-thirds of hindtibia, and basal two-thirds of hindbasitarsus white. Wings lightly infuscated on basal two-thirds, hyaline on apical one-third.

Antenna with third segment slightly shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus shallowly, circularly emarginated. Hindwing with cells Rs and M absent; anal cell with short petiole. Lancet with 15 to 16 serrulae; each serrula shallow, not extending far below ventral margin of lancet, slightly asymmetrical with two or three anterior and four or five posterior subbasal teeth (pl. V, 66).

Male.—Length, 4.3 mm. Color similar to that of female except for each femur, which is whitish. Hindwing with peripheral vein; anal cell with apex removed some distance from margin of wing (pl. I, 7). Genitalia as in plate IX, 100; harpe about as long as broad.

Larva .-- Unknown.

Holotype.—Female, Caroline Co., Va., 7-15-1941, larva on Quercus palustris. L. A. Hetrick. U.S.N.M. type No. 70216.

Allotype.—Male. Same data as for holotype. Deposited with holotype.

Paratypes.—Maryland: College Park, Aug. 9, 1919, Quercus palustris, E. N. Cory (3 9 9). Pennsylvania: 6 mi. S. E. Meadville, Crawford Co., VII-1-61, on oak, F. B. Negley, collector (2 9 9). Virginia: Same data as for holotype (2 9 9, 1 8); data as for holotype, V-16-1941 (2 9 9); data as for holotype, V-17-1941 (1 8). Deposited in the U.S. National Museum.

Distribution.—Eastern North America.

Host.—The type series was reared from larvae on Quercus palustris Muenchh.

Biology.--Unknown.

Discussion.—The petiolate anal cell and absence of cells Rs and M of the hindwing, short lancet, and shallow serrulae of the lancet will separate the females of this species. The anal cell with the apex removed from the margin of the hindwing, white femora, and genitalia will separate the males.

# Caliroa quercuscoccineae (Dyar)

Monostegia querous-coccineae Dyar, 1894, p. 42, Q. larva; Dyar, 1895b, p. 338; Taylor, 1931, p. 459.

Caliroa quercus-coccuneae: MacGillivray, 1909b, p. 359; MacGillivray, 1916, p. 80; Yuasa, 1922, p. 59; Ross, 1951, p. 29.

Female.—Average length, 4.7 mm. Black; extreme apex of each femur, all front and middle tibiae and tarsi, basal two-thirds of hindtibia, and basal two-thirds of hindbasitarsus white. Wings with basal two-thirds moderately infuscated, apical one-third hyaline.

Antenna with third segment shorter than segments 4 plus 5; apical four segments together subequal in length to third segment. Clypeus shallowly, circularly emarginated. Hindwing with cells Rs and M present; anal cell sessile. Lancet with 17 to 18 serrulae; each serrula moderately deep, rounded at apex, nearly symmetrical, and with three or four anterior and three or four posterior subbasal teeth (pl. VII, 76).

Male.—Average length, 4.5 mm. Color similar to that of female. Hindwing with peripheral vein; anal cell with apex close to margin of wing (pl. I, 6). Genitalia similar to plate IX, 103 and 104; harpe

longer than broad, narrowing toward apex.

Larva.—Dyar (1894) described the larva and stated that the head capsule is "orange-yellowish." The characters given by Dyar do not permit an adequate characterization of this species.

Holotype.—Female, U.S.N.M. type No. 4127, labeled "T" and "Collection, H. G. Dyar." Dyar's "T" is from Woods Holl, Mass.

Distribution.—Eastern North America. Connecticut: Lyme, 7-8-18; Stonington, VII-5-32, Quercus velutina. Delaware: "Delaware," Sept. 23, 1960, on oak. District of Columbia: Sept. 10, 1913, coll. on pin oak. Illinois: Urbana, June 20, 1932; Carbondale, July 22, 1909. Louisiana: Shreveport, III-27-07. Maine: Augusta, VII-21-1944. Maryland: Glen Echo, 4-97. Michigan: "Mich." Minnesota: Northfield. VII-3-17. Missouri: Warrenton, VII-8-1963. New Hampshire: Hanover. New Jersey: Westfield, July 15, 1956. North Carolina: Durham, Oct. 8, 1942. Quercus palustris. Pennsylvania: North East, VI-18-16; Inglenook, VI-27-09. Virginia: Falls Church, reared VII-29-16, Quercus alba, reared, Aug. 26, 1912, Quercus velutina. Wisconsin: Trout Lake, July 21-23, 1937; Minong, Kimball Lake, Aug. 25, 1945.

Host.—Quercus spp. Dyar (1894) reared the type series from "Quercus coccinea." Other specimens examined have been reared from larvae on Quercus velutina Lam., Q. palustris Muenchh., and Q. alba L.

**Biology.**—The only information on the biology is that by Dyar (1894). The eggs are inserted parallel to the midrib of the leaf. The larvae feed gregariously, and, after maturing, drop to the ground and "form little elliptical cells in the ground for pupation."

Discussion.—The sessile anal cell and presence of cells Rs and M in the hindwing, the partially infuscated wings, and rounded serrulae of the lancet will serve to distinguish this species. It is closest to fasciata but may be separated by the fewer serrulae of the lancet: fasciata has more than 20, quercuscoccineae has less than 20. There are apparently no distinguishing characters between the males of the two species. If the distinguishing characters between fasciata and quercuscoccineae are found to be variable, it may be necessary to synonymize the two species.

# Tribe HETERARTHRINI

The single genus in the tribe Heterarthrini may be separated by the complete vein 2A and 3A of the forewing, the 10- or 11-segmented antenna, and the bifid tarsal claw with an acute basal lobe. The larvae are dorsoventrally flattened, typical of the leafmining larvae of the Fenusini, and may be separated in the key to larvae. Only one species is in North America, introduced from Europe.

# Genus HETERARTHRUS Stephens

Phyllotoma Fallén, 1829, p. 25; Gameron. 1882, p. 282; Dalla Torre, 1894, p. 197; Ashmead, 1898, p. 255; Konow, 1905, p. 72; Enslin, 1914, p. 257; Forsius, 1930, p. 103; Ross, 1937, p. 70; Weber, 1939, p. 541; Berland, 1947, p. 273. Preoccupied.

Type-species: Phyllotoma vagans Fallen. Designated by Rohwer, 1911b. Heterarthrus Stephens, 1835, p. 94; Westwood, 1840, p. 54; Dalla Torre, 1894, p. 197 (=Phyllotoma Fallen); Ross, 1951, p. 29; Benson, 1952, p. 77; Malaise, 1961, p. 236.

Type-species: Decatria fuscipennis Stephens (=Tenthredo (Emphytus) ochropoda Klug). Original designation.

Heterarthus Cameron, 1882, p. 282. Emendation for Heterarthrus Stephens.

Decatria Stephens, 1835, p. 94; Dalla Torre, 1894, p. 197 (=Phyllotoma Fallén).

Type-species: Decatria fuscipennis Stephens. Monotypic and original designation.

Druida E. Newman, 1838, p. 484; Dalla Torre, 1894, p. 197 (=Phyllotoma Fallén).

Type-species: Druida parviceps E. Newman. Monotypic.

Phlebatrophia MacGillivray, 1909b, p. 345; Forsius, 1930, p. 103 (=Phyllotoma Fallén).

Type-species: Phlebatrophia mathewsoni MacGillivray. Monotypic and original designation.

Description.—Antenna 10- to 12-segmented (pl. III, 31). Head flattened, markedly depressed between eyes from dorsal view (pl. III, 30). Malar space twice width of diameter of front ocellus; clypeus truncate; postgenal carina absent. Tarsal claw with two long subequal outer teeth and large acute basal lobe (pl. III, 23). Prepectus absent. Forewing with vein 2A and 3A complete for entire length, connected to 1A by oblique crossvein. Hindwing with radial cell open; cells Rs and M absent; anal cell present (pl. I, 8, 9).

This very distinct genus is separated from other Heterarthrinae genera by the antenna, head, wing venation, and tarsal claw. Benson (1952) placed this genus in a subfamily by itself, the Heterarthrinae, and placed the other taxa treated in this bulletin in the subfamily Blennocampinae. About 10 species are in this genus. Only one species is found in North America, an introduced form that is destructive to Betula spp. in northeastern United States and eastern Canada. The larva is known for this species and is treated under the species description.

# Description of Heterarthrus Species

Heterarthrus nemoratus (Fallén)

Hylotoma nemorata Fallén, 1808, p. 47.
Phyllotoma nemorata: Thomson, 1870, p. 268; Thomson, 1871, p. 176; André, 1880, p. 235; Cameron, 1882, p. 284; Brischke and Zaddach, 1883, p. 254; Konow, 1896, p. 248; Dalla Torre, 1894, p. 198; Konow, 1905, p. 72; Enslin, 1914, p. 258; Rohwer, 1929, p. 63; Taylor, 1929, p. 323; Dovnar-Zapolsky, 1929, p. 39; Pierson, 1929, p. 588; Pierson, Taylor, and Wilkins, 1930, p. 1; Taylor, 1931, p. 452; Glasgow, 1932, p. 693; Conde, 1934, p. 181; Twinn, 1934, p. 76; Pierson and Brower, 1936, p. 37; Goebeil, 1937, p. 1; Hering, 1937, p. 73; Balch, 1939, p. 36; Benson, 1940, p. 213; Buhr, 1941, p. 910; Wahlgren, 1944, p. 139; Reeks and Smith, 1945, p. 10; Berland, 1947, p. 274.
Heterarthrus nemoratus: Ross, 1951, p. 30; Benson, 1952, p. 77; Maxwell. Hylotoma nemorata Fallén, 1808, p. 47.

Heterarthrus nemoratus: Ross, 1951, p. 30; Benson, 1952, p. 77; Maxwell, 1955, p. 56; Raizenne, 1957, p. 25; Lorenz and Kraus, 1957, p. 146; Lindquist, 1959, p. 626; Benson, 1962, p. 389; Burks, 1967, p. 16.

Phyllotoma nemoratis Fallén, 1829, p. 35; Dalla Torre, 1894, p. 198 (= nemo-

rata Fallén).

Phlebatrophia mathewsoni MacGillivray, 1909b, p. 345, 9 ; Yuasa, 1922, p. 60; Prison, 1927, p. 256; Rohwer, 1929, p. 63 (= nemorata Fallen).

Female.--Average length, 4.5 mm. Antenna black; head black with clypeus, mouth parts, inner orbits, malar space, supraclypeal area, and spot above each antenna yellow. Thorax black with posterior half of pronotum and tegula vellow. Legs white with each coxa and basal half of each femur black. Abdomen black with yellow spots on lateral apical margin of each terga. Wings hyaline, infuscate band below stigma, lighter on hindwing; costa of forewing yellow, stigma black.

Antenna with 10 or 11 segments (pl. III, 31). From above, head markedly depressed between eyes (pl. III, 30). Sheath straight above, rounded below, with narrow scopa at apex (pl. IV, 43, 44).

Male,-Unknown, Parthenogenetic.

Larra.—Final feeding stage (pl. XII, 142-147). Length, 8.0 mm. Dorsoventrally flattened. White with head capsule, pronotal plate, prosternal plate, small central areas of mesosternum and metasternum, and ring around anal proleg dark brown.

Head dorsoventrally flattened, prognathous. Antenna conical, four-segmented. Labrum slightly wider than long, truncate; epipharnyx with about 12 long spines on each half; maxillary palpus four-segmented, first and second segments each with one seta; galea long, digitate: lacinia with two long bifurcate spines and numerous smaller spines below these; right mandible with dorsal and ventral cutting edges; left mandible similar to right mandible but dorsal edge divided into three blunt teeth; one seta on basal angle of each mandible; labial palpus three-segmented, mentum with numerous small spines.

Pronotal plate present; prosternal plate present, cut into large circular areas around each prothoracic leg: small dark plates at center of mesosternum and metasternum. Thoracic legs indistinctly three-segmented; tarsal claw minute, appearing absent. Prothoracic spiracle not winged.

Prolegs on abdominal segments 2 to 8 and 10; anal proleg sur-

rounded by sclerotized plate. Segments each two-annulate.

Holotypes.—MacGiffivray's type is at the Illinois Natural History Survey, a female, labeled "New Glasgow, Nova Scotia." Fallén's types are probably in the Zoological Museum, Lund, Sweden.

Distribution.—Northeastern United States and southeastern Canada; Europe. The following records are from specimens examined: Maine: Bar Harbor: Aurora; Bethel; Phillips; Mt. Desert Is. Massachusetts: Amherst. New Brunswick: Fredericton. Newfoundland: Grand Falls. New Hampshire: Mt. Washington; Shelburne; Gorham. New York: Keene; Mt. Marcy summit. Nova Scotia: New Glasgow; Truro. Quebec: Gaspė; Cascapedia R.; Lake St. John. Raizenne (1957) recorded this species from the following localities in southern Ontario: Bruce, Kent, Leeds, Simcoe, Victoria. Welland. and York.

Host.—Leaf miner of Betula spp. Glasgow (1982) stated that the preferred hosts are white and gray birch, although yellow and

black birch are altacked to some extent.

Biology.—Pierson, Taylor, and Wilkins (1930) studied this species in Maine. The adults appear in May or June, later at higher elevations, and the female oviposits in the leaf margin. The feeding period lasts about a month, during which the brivae mine throughout the leaf. The mined areas first appear as small brown patches on the edge of the leaf, and later these patches increase in size and merge together. On maturing, the larvae remain in the leaf to overwinter and pupate. There is one generation a year. Of the four birch leaf-mining sawflies, both nemoratus and Messa nana oviposit on the leaf margins; Fenusa pusilla and Profenusa thomsoni oviposit on the surface of the leaf. However, nana goes into the soil to overwinter and pupate, whereas nemoratus remains in the leaf. Pierson (1929). Glasgow (1932), Pierson and Brower (1936), and Goebeil (1937) have all published notes on the biology of this species.

Discussion.—This species is easily recognized by the antenna, head, and wing venation. The larvae may be separated from other birch leaf-mining sawfly species by the presence of prolegs and the

presence of a sclerotized ring only around the anal proleg.

The first record for nemoratus in North America was from Pictou, Nova Scotia, in 1908. By 1927 this species had reached epidemic proportions in eastern Maine, and by 1928 it was found throughout Maine to New Hampshire and Massachusetts. Glasgow (1932) first reported it from New York. Recently the populations have been at a very low ebb.

# Tribe FENUSINI

The larvae of all species of the tribe Fenusini are leaf miners. The adults may be distinguished by their small size and petiolate

anal cell of the forewing. The larvae are small and dorsoventrally flattened, with a varying number of sclerotized plates on the body.

#### Genus METALLUS Forbes

Metallus Forbes, 1885. p. 87; Dalla Torre, 1894, p. 156 (= Fenusa Leach);
MacGillivray, 1909a, p. 265; MacGillivray, 1916, p. 159; Benson, 1936,
p. 622; Ross, 1937, p. 71; Conde, 1937, p. 108; Benson, 1940, p. 212; Benson, 1941, p. 86; Berland, 1947, p. 261; Ross, 1951, p. 30; Benson, 1952,
p. 106; Takeuchi, 1952, p. 59; Lorenz and Kraus, 1957, p. 133; Smith, 1967, p. 280.

Type-species: Metallus rubi Forbes. Monotypic.

Entodecta Konow, 1886b, p. 184: Dalla Torre, 1894, p. 166; Ashmead, 1898, p. 253; Konow, 1904, p. 4; Konow, 1905, p. 85; Helién, 1935, p. 22; Benson, 1930, p. 622 (= Metallus Forbes); Ross, 1937, p. 71; Ross, 1951, p. 30 (= Metallus Forbes).

Type-species: Tenthredo (Allantus) pumila Klug. Designated by MacGillivray, 1909a.

Polybates MacGillivray, 1909a, p. 264; MacGillivray, 1916, p. 158; Ross, 1937, p. 71 (= Entodecta Konow); Ross, 1951, p. 30 (= Metallus Forbes). Type-species: Polybates slossonae MacGillivray. Monotypic.

Description.—Antenna with second segment broader than long; third segment slightly longer than or subequal in length to fourth segment; in males third and fourth antennal segments widened and laterally flattened, segments beyond fourth less distinctly flattened (pl. III, 38, 39). Postgenal carina absent; malar space linear; clypeus truncate. Prepectus absent. Tarsal claw with long outer tooth and large acute basal lobe (pl. III, 22). Forewing with vein 2A and 3A straight at apex. Hindwing with cell  $R_1$  closed; anal cell present: cells Rs and M absent (pl. II, 10, 11).

The broad second antennal segment and the flattened antenna of the male will separate this genus from Messa, its closest relative. Also, the serrulae of the female lancet are long and lobelike in species of Metallus and flattened in species of Messa, and the penis valve of the male genitalia of Metallus lacks a long apical filament, which is found in males of Messa species.

About eight described species are in this genus, three of which are found in North America. Two of the North American species are associated with *Rubus*.

Larvae are known for two species of *Metallus*. They may be separated from those of all other genera of Fenusini by the presence of prolegs on abdominal segments 2 to 7 and 10. Other genera have no prolegs or they are present on segments 2 to 8 and sometimes 10.

# Keys to Metallus Species

#### ADULTS

1. All tibiae black; mesonotum sometimes partly rufous; antenna of male laterally flattened rohweri MacGillivray Legs entirely yellowish; thorax always black; male unknown 2

Serrulae of lancet long, pointed at apex, with single anterior subbasal tooth near ventral margin of lancet (pl. VIII, 87) bensoni, n. sp. Serrulae short r, rounded at apex, with one anterior and one posterior subbasal tooth near ventral margin of lancet (pl. VIII, 88) capitalis (Norton)

#### LARVAE

Mesosternum and metasternum with dark plates; lacinia with three
or four spines (pl. XIII, 154, 159) rohweri MacGillivray
Mesosternum with small dark plate on anterior margin; metasternum
without plate; lacinia with three spines capitalis (Norton)

### Descriptions of Metallus Species

Metallus bensoni, new species

Female.—Length, 4.5 mm. Black with all legs entirely yellowish.

Wings uniformly, moderately infuscated.

Second segment of antenna broader than long; third segment slightly longer than fourth segment (pl. III, 38). Clypeus truncate; malar space less than diameter of front occilius. Tarsal claw with one outer tooth and large acute basal lobe (pl. III, 22). Forewing with vein 2A and 3A straight at apex. Hindwing with radial cell closed; cells Rs and M absent; anal cell present. Sheath straight above, rounded below, scopa absent (pl. IV, 46). Serrulae of lancet long, each pointed at apex with one anterior subbasal tooth near ventral margin of lancet and no posterior subbasal teeth (pl. VIII, 87).

Male.—Unknown. Larva.—Unknown.

Holotype.—Female, New York, Albany Co., nr. Rensselaerville, Huyck Preserve, 23 June 1967, Malaise trap 1, R. and J. Matthews. Deposited in the Museum of Comparative Zoology, Harvard University.

Paratypes.—British Columbia: Grouse Mt., July 20, 1936, H. H. Ross (1 ?). New York: Same data as for holotype, 9 June 1967 (1 ?), 21 June 1967 (1 ?), 1 July 1967 (1 ?), 9 July 1967, Malaise trap 4 (1 ?), 13 July 1967 (1 ?), 17 July 1967 (1 ?), 23 July 1967 (1 ?), 8-12 August 1967 (1 ?), 9 August 1967 (1 ?), 14 August 1967 (2 ??), 14 August 1967, Malaise trap 4 (1 ?), 17 August 1967 (1 ?), 17 August 1967, Malaise trap 2 (1 ?), 18 August 1967 (2 ??), 19 August 1967 (1 ?), 20-23 August 1967 (2 ??), 5-7 September 1967, Malaise trap 2 (1 ?). Deposited in the Museum of Comparative Zoology, Harvard University, U.S. National Museum, and at the Illinois Natural History Survey.

Distribution.—Known only from British Columbia and New York.

Host.--Unknown.

*Biology.*—The dates of capture extend throughout the summer. There may be several generations a year.

**Discussion.**—This species is similar to capitalis in coloration, but bensoni is slightly larger, the wings are slightly darker, and the serrulae of the lancet are longer, pointed at their apices, and

each has only one anterior subbasal tooth. The entirely yellowish legs will separate bensoni from rohweri. The British Columbia specimen is identical to those from New York.

#### Metallus capitalis (Norton)

Selandria capitalis Norton, 1867, p. 247, 9; Cresson, 1928, p. 4. Blennocampa capitalis: Cresson, 1880, p. 59; Dalla Torre, 1894, p. 170; Kanow, 1905, p. 84.

Scolioneura capitalis: Marlatt, 1895, p. 234.

Metallus capitalis: MacGillivray, 1909a, p. 266; MacGillivray, 1916, p. 159;

Ross, 1951, p. 30; Smith, 1967, p. 280; MacGillivray, 1910, p. 199; Ross, 1951, p. 30; Smith, 1967, p. 280.

Entodecta capitalis: Ross, 1937, p. 71.

Entodecta humilis Konow, 1908, p. 84, \(\chi\); Ross, 1937, p. 71 (= capitalis Norton). Synonymy not certain.

Polybates slossomae MacGillivray, 1909a, p. 265, \(\chi\); MacGillivray, 1916, p. 158; Frison, 1927, p. 256; Ross, 1937, p. 71 (= capitalis Norton).

Polybates secundus Rohwer, 1910, p. 202, \(\chi\); MacGillivray, 1916, p. 159; Ross 1927, p. 71 (= capitalis Norton). Ross, 1937, p. 71 (= capitalis Norton).

Female.—Average length, 4.0 mm. Black with all legs entirely

yellowish. Wings uniformly, lightly infuscate.

Antenna slender, third segment slightly longer than fourth segment (pl. III, 38). Sheath straight above, rounded below, scopa absent (pl. IV, 46). Serrulae of lancet each lobelike, rounded at apex, with one anterior and one posterior subbasal tooth, each located near ventral margin of lancet (pl. VIII, 88).

Male.—Unknown.

Larva.—Very similar to that of rohweri with the following differences: Small plate on anterior margin of mesosternum; metasternum without plate; lacinia with three spines; palpifer with two setae; mandibles slightly different, as in plate XIII, 162 and 163. Described from larvae taken from raspberry leaves, Friday Harbor, Washington, Sept. 17, 1956.

Holotypes.—S. capitalis Norton, a female labeled "N.Y." is type No. 10338 at the Academy of Natural Sciences of Philadelphia. P. slossonae, a female labeled "Franconia, N.H.," is at the Illinois Natural History Survey. P. secundus Rohwer, a female labeled "Red Head, St. John, N.B., Sept. 1, '07, A. G. Leavitt, collector," is U.S.N.M. type No. 12926. Konow's type is probably at the

Deutsches Entomologisches Institut, Eberswalde, Germany.

Distribution.—Probably transcontinental across northern United States and southern Canada. British Columbia: Vancouver, 19-VIII-32. Illinois: Mineral Springs, VIII-28-16. Maine: Mt. Katahdin, Camp Kennedy. 3000', Aug. 1902; Mt. Katahdin, 8-31-23; Bar Harbor, emgd. V-43, miner ex raspberry. New Brunswick: Red Head, St. John. Sept. 1, 1907. Newfoundland: 20 mi. N.E. Deer Lake, June 28, 1966. New Hampshire: Franconia; Hampton. New York: Brooklyn; Niagara Falls, 9-4-11. Oregon: Astoria, 25-5; Astoria, May 4, 1962, raspberry; Spring Cr., Baker, 11-VIII-1952, Quebec: St. Hillaire; He Jesus, 4 mi. N. Montreal, Aug. 19, 1956.

Host.—Adults have been reared from larvae mining in the leaves of raspberry (Rubus sp.).

Biology.—Collection dates of adults range from May to Septem-

ber, most of them in August, indicating that there may be more than one generation a year. There is no literature on the life history of this species.

**Discussion.**—This is a small species that may be separated from bensoni by the lower and more rounded serrulae of the lancet and the presence of a posterior subbasal tooth on each serrula. The yellowish tibiae will separate capitalis from rohweri.

#### Metallus rohweri MacGillivray

Metallus rubi Forbes, 1885, p. 87, &; MacGillivray, 1909a, p. 268; MacGillivray, 1916, p. 160; Yuasa, 1922, p. 99; Frison, 1927, p. 210; Rohwer, 1927, p. 67; Daniel, 1928, p. 5; Ross, 1937, p. 72. Preoccupied.

Fenusa rubi: Cresson, 1887, p. 160; Dalla Torre, 1894, p. 157; Dyar, 1898, p. 137; Konow, 1905, p. 90.

Metallus rohweri MacGillivray, 1909a, p. 267, 9; MacGillivray, 1916, p. 159; Frison, 1927, p. 252; Ross, 1937, p. 72 (= rubi Forbes); Ross, 1951, p. 30; Smith, 1967, p. 280.

Metallus bethunei MacGillivray, 1914, p. 366. \$\(\delta\), \(\text{ ? }\); MacGillivray, 1916, p. 160; Caesar, 1921, p. 38; Yuasa, 1922, p. 99 (?); Frison, 1927, p. 251; Rohwer, 1927, p. 67 (= rubi Forbes); Daniel, 1928, p. 5 (= rubi Forbes).

Scolioneura capitalis of auctt., nec Norton, Houghton, 1908, p. 212; Houghton, 1910, p. 10.

Female.—Average length, 5.2 mm. Black with labrum, each trochanter, tibia, and tarsus white; thorax sometimes rufous with metapleuron and pectus black, or with intermediates between this and those with thorax all black. Wings darkly infuscated, usually darker on basal one-half.

Antenna stout, third segment subequal in length to fourth segment (pl. III, 38). Sheath short, straight above, rounded below, subtruncate at apex; scopa absent (pl. IV, 45). Serrulae of lancet shallow, broad, rounded at apex, each with three or four small anterior subbasal teeth and no posterior subbasal teeth (pl. VIII, 89).

Male.—Average length, 5.0 mm. Color similar to black specimens of females, never with rufous thorax. Antenna thickened, laterally flattened, especially noticeable on segments 3 to 5 (pl. III, 39). Genitalia as in plate X, 108 and 109; penis valve long and slender, with long apical spine.

Larva.—Final feeding stage (pl. XIII, 154-161). Length, 9.0 mm. Body cylindrical, slightly dorsoventrally flattened. White; head capsule, plates on venter and dorsum of thorax, and crescent-

shaped marks about each proleg brownish.

Head dersoventrally flattened, prognathous, diamond shaped from above. Antenna one-segmented. Labrum slightly emarginated; epipharynx with row of about eight to 10 long spines on each half; right mandible with one ventral tooth and three rounded dorsal teeth; left mandible with one rounded ventral tooth, one large rounded dorsal tooth, and one blunt inner tooth; maxillary palpus four-segmented, one seta on second segment; palpifer with one seta; galea long, digitate; lacinia short, inconspicuous, with three or four stout spines; labial palpus small, two-segmented.

Pronotum, mesonotum, and metanotum each with lightly sclero-

tized plate, pronotal plate largest. Prosternum with dark plate covering nearly entire sternum; mesosternum and metasternum with smaller central plates. Prothoracic spiracle winged. Thoracic legs five-segmented; tarsal claw present.

Abdominal segments 2 to 9 each with two annulets; pleural lobes present. Prolegs on abdominal segment 2 to 7 and 10, dark crescent-shaped plate present about anterior margin of each proleg, those about anal prolegs largest. Abdominal spiracles not winged.

Holotypes.—At the Illinois Natural History Survey, a specimen is labeled as holotype of M, rubi Forbes. It is a male and is without data. Frison (1927) stated that there is a lectotype male with the data "Normal, Illinois, reared from mines in leaves of raspberries, August 12, 1884." MacGillivray's types are at the Illinois Natural History Survey: M, rohweri, a female, is labeled "Block Island, R.I., Aug. 28, 1891," and M, bethunci, a female, is labeled "Jordan Harbor, July 5, 1919."

Distribution.—Eastern North America. Connecticut: Lyme, Aug. 27, 1918, 5-4-18, 7-8-18, 7-11-18; Branford, 2-IX-51; Windsor, *Delaware*: Dover, 5-30-1906, *Florida*: Archibold Biological Sta., Highlands Co., 2-6-66, blotch miner, blackberry. Illinois: "N. Ill." Maine: Mt. Desert, emgd. 5-VI-1953, ex black-berry; Augusta, VIII-21-43, about dewberry. Marylard: Cabin John, 7-IX-12, 14-9-14, 28-VIII-1916; Plummers Islan Aug. 4, 1911; Marshall Hall; Bowie, V-30-44; Glen Echo. Massachusetts: Boston, Arnold Arboretum, July 25, 1921; Sherborn, Nantucket Island, Michigan: Stevensville, Berrien Co., V-29-38, Missouri: "C. Mo." New Jersey: Somerville, 1921. New York: Cold Spring Harbor, VIII-9-20; Gardiner's Island; Fredonia; Greenport, L.I., Sept. 6, 1951; Ithaca, Aug. 5, 1898, North Carolina: Chadbourn, Apr. 16, 1910, collected on dewberry. Ohio: Put-in-Bay. Ontario: St. Kits, VIII-12-1911; Jordan Harbor; Vineland, Aug. 14, 1952; Toronto, 7-10-09. Pennsylvania: Harrisburg, Hetzels Swamp, July 9, 1910; Encola, VI-7-08; Colmanville, iss. 14 Aug. '05, leaf miner in blackberry; Presque Is., 7-17-1947, Rhode Island: Block Island, Virginia: Falls Church, VI-17-17, June 9, 1921, June 25, 1921; Whiele, Aug. 28, 1943; Eastern Field Station, Aug. 29, 1918. Wisconsin: Trout Lake, Aug. 1911; Baraboo, 8-10-10.

Host.—Leaf miner of blackberry (Rubus spp.). Daniel (1928) stated that they may attack dewberry close to blackberry patches.

Biology.—Daniel (1928) gave the life history of this species from his studies in New York. Two generations a year occur, adults of the first generation appearing in June and those of the second generation in August. The eggs are inserted into the leaf from the upper side, usually on the surface near the midrib or a prominent vein. The egg and larval stages combined last about 6 weeks, after which the mature larva drops to the ground to pupate or overwinter in the soil. Houghton (1908, 1910) also published notes on the life history of this species in Delaware under the name "Scolioneura capitalis."

*Discussion.*—This species may be separated from the other two species of *Mctallus* by the black femora, lower and broader serrulae

of the lancet, and, when present, the red thorax. Also, rohweri is a larger species and the wings are more darkly infuscated than in bensoni and capitalis.

#### Genus MESSA Leach

Messa Leach, 1817, p. 126; Westwood, 1840, p. 54; Dalla Torre, 1894, p. 156 (= Fonusa Leach); Enslin, 1914, p. 307; MacGillivray, 1914, p. 365; Benson, 1936, p. 625; Benson, 1941, p. 86; Ross, 1951, p. 30; Benson, 1952, p. 107; Takeuchi, 1952, p. 59; Lorenz and Kraus, 1957, p. 136; Malaise, 1964, p. 33; Smith, 1967, p. 280.

Type-species: Tenthredo hortulana Klug. Monotypic.

Fenusella Enslin, 1914, p. 270; Benson, 1936, p. 625; Stritt, 1936, p. 56; Benson, 1941, p. 86 (= Messa Leach); Berland, 1947, p. 265; Malaise, 1964.

Type-species: Fenusa wuestneii Konow. Monotypic and original designation.

Melanobates MacGillivrny, 1916, p. 158; Benson, 1936, p. 622 (= Fenusella Enslin); Ross, 1937, p. 71 (= Scotioneura Konow); Ross, 1951, p. 30 (= Messa Leach).

Type-species: Parabates leucostomus Rohwer. Monotypic.

Description .-- Antenna nine- or 10-segmented; second segment longer than broad; third segment longer than fourth segment; antenna of males not widened or laterally flattened (pl. III, 35, 36). Postgenal carina absent; clypeus truncate; malar space linear. Tarsal claw with long outer tooth and large acute basal lobe (pl. III, 22). Forewing with vein 2A and 3A curved up at apex. Hindwing with cell  $R_1$  closed; anal cell present; cells  $R_2$  and M absent (p. II, 12, 13).

This genus is separated from Metallus by the long second antennal segment and by the third antennal segment, which is always longer than the fourth segment. The antennae of males of Messa species are not enlarged as are those of Metallus species. Also, the genitalia of Messa species are relatively uniform, having serrulae of the female lancet flat and a long apical filament on the penis valve of the male genitalia. The color pattern of the species of this genus appears constant within the species.

About seven or eight species are in this genus, five of which are found in North America. All are leaf miners in the larval stage and are associated with Betula, Populus, and Salix.

The larvae are not well enough known to permit a key for their separation. I have seen some specimens from Populus, but they have not been associated. The only larva I have described and specimens of which I have examined is that of nana. The larvae of Messa may be separated from those of other genera of Fenusini by the following combination of characters: Abdominal prolegs present on segments 2 to 8 and 10; a dark crescent-shaped plate anterior to each proleg; mesosternum, metasternum, and sternum of first abdominal segment each with a dark plate. The larvae are typical of most leaf miners and are dorsoventrally flattened with the head prognathous.

unestneii (Konow)

#### Key to Messa Species

1. From with distinct circular ridge; mesonotum without fine surface sculpture; segment 8 of antenna more than twice as long as broad; forewing with infuscate band below stigma; male unknown; leaf miner of Betula Frons without ridge; mesonotum with fine surface sculpture on front and lateral lobes; segment 8 of antenna less than two times longer than broad; forewing uniformly subhyaline 2 2. Mesopleuron pale yellowish or orange 3 Mesopleuron black 4 3. Head, pectus, and venter of abdomen mostly yellowish orange; male with coloration similar to that of female; leaf miner of Populus populifoliella (Townsend) Head, pectus, and abdomen black; clypeus yellowish; male unknown; hortulana (Klug) leaf miner of Populus 4. Clypeus and pronotum white; male and female similar; leaf miner of Populus. lencostoma (Rohwer) Clypeus and pronotum black; male and female similar; leaf miner

#### Descriptions of Messa Species

#### Messa hortulana (Klug)

Tenthredo (Emphytus) hortulana Klug, 1814, p. 276, 9; Kriechbaumer, 1884, p. 208.

Messa hortulana: Leach, 1817, p. 126; Stephens, 1835, p. 40; Westwood, 1840, p. 54; Enslin, 1914, p. 308; Koornneff, 1925, p. 360; Dovnar-Zapolsky, 1929, p. 38; Hering, 1934, p. 67; Stritt, 1935, p. 187; Benson, 1941, p. 89; Benson, 1943, p. 10; Jones, 1945, p. 218; Benson, 1952, p. 108; Lorenz and Kraus, 1957, p. 137; Malaise, 1964, p. 33; Benson, 1968, p. 150.

Emphytus hortulana: Hartig, 1837, p. 258. Phaenusa hortulana: André, 1880, p. 231.

of Salix

Fenusa hortulana: Fletcher, 1881, p. 127; Cameron, 1882, p. 296; Konow, 1885, p. 297; Konow 1886b, p. 270; Konow, 1890, p. 249; Dalla Torre, 1894, p. 157; Konow, 1905, p. 90.

Feunsella hortulana: Benson, 1936, p. 625; Buhr, 1941, p. 915; Stritt, 1944, p. 25; Berland, 1947, p. 267.

Female.—Average length, 4.0 mm. Antenna black above, lighter on ventral surface; head black with clypeus, basal half of each mandible, and rest of mouth parts whitish. Thorax black with tegula and pronotum white, spot on lateral margins of prescutum and all of mesopleuron yellowish orange. Legs entirely yellowish orange with each trochanter, base of each coxa, and each tarsus infuscated. Abdomen black with white band on posterior margin of each segment. Wings hyaline.

Antenna 10-segmented, if nine-segmented then ninth segment at least 1½ times length of eighth segment (pl. III, 35). Frons without ridge. Front and lateral lobes of mesonotum with fine surface sculpture. Sheath short, straight above, rounded below, with narrow scopa at apex (pl. IV, 51, 52). Lancet with each serrula low and flattened, with one are not four or five posterior subbased teach again plate MH 30

terior subbasal teeth as in plate VII, 80.

Male.—Unknown in North America. The males are very rare in Europe (Benson, 1952).

Larva.—Unknown.

Holotype.--Probably in the Zoological Museum of Berlin.

Distribution.—In North America known only from Massachusetts; Europe. Massachusetts: Malden, lombardy poplar, Gip. Moth Lab. 12164J18, bred specimens, 5-2-24 (6 9 9), 5-3-24 (1 9), 5-10-24 (2 9 9); Wellesley (1 9).

Host.—Adults were bred from larvae on lombardy poplar (Populus nigra var. italica Muenchh.) in Massachusetts. Benson (1952) recorded the best as Populus migra I. in Findend.

(1952) recorded the host as Populus nigra L. in England.

Biology.—There is little information on the life history of this

species except notes by Buhr (1941) and Hering (1934).

Discussion.—This species is separated by the yellowish-orange mesopleuron and black pectus. M. populifoliella has considerably more yellow on the head and abdomen, and leucostoma and wwestness are nearly entirely black.

This is the first North American record for hortulana. The specimens from Massachusetts are identical to specimens from Europe and were also bred from an introduced shade tree, lombardy poplar, which is native to Europe. This is probably an introduced species.

# Messa leucostoma (Rohwer)

Parabates leucostoma Rohwer, 1910, p. 202, Q. Scolioneura leucostoma: Ross, 1937, p. 71.

Messa lencostoma: Ross, 1951, p. 30.

Riennocampa amara MacGillivray, 1923a, p. 161, 9; Frison, 1927, p. 238; Ross, 1951, p. 30 (= leucostoma Rohwer).

Female.—Average length, 4.0 mm. Antenna black, first segment and ventral surface of rest of antenna sometimes brownish. Head black with clypeus, labrum, basal half of each mandible, and rest of mouth parts whitish; supraclypeal region, paraantennal fields, and spot above each antenna sometimes brownish. Thorax black with tegula white and posterior margin of pronotum white; rest of pronotum sometimes brownish. Legs mostly black with white to infuscated areas on extreme apex of each femur and each front and middle tibia. Abdomen black with narrow to wide white margin on posterior edge of each segment. Wings hyaline.

Antenna nine-segmented; ninth segment shorter or subequal in length to eighth segment. Head without circular ridge on frons. Front and lateral lobes of mesonotum with fine surface sculpture. Sheath straight above, rounded below, with narrow scopa at apex (pl. IV, 51, 52). Lancet with each serrula low and flattened, with one anterior and three or four posterior subbasal teeth as in plate

VII. 80.

Male.—Average length, 4.0 mm. In color and structure similar to female. Genitalia as in plate X, 112 and 113; harpe rounded at apex, subtruncate at base and at inner lower margin; penis valve broad, with long apical filament.

Larva.-Unknown.

Holotypes.—P. leucostoma Rohwer is U.S.N.M. type No. 12925, a female labeled "St. John, N.B., 18 Jul., A. G. Leavitt, collector."

B. amara MacGillivray is at the Illinois Natural History Survey, a female, labeled "Edmonton, Alta., 21-V-1917, F. S. Carr."

Distribution.—Transcontinental across southern Canada and northern United States with southern extensions into California and Colorado. Alberta: Edmonton, May 21, 1917. California: Elkhorn Ferry, Yolo Co., March 30, 1963; Winnemucca Lake, Carson Pass, July 14, 1964; Pachaco, Contra Costa Co., on cottonwood, 4-3-1958; Mint Canyon, 3-8-62; Forestville, April 21, 1938. Colorado: "Colo." Iowa: Ames, May 7, 1950. Minnesota: Itasca Park, May 23, 1937. New Brunswick: St. John, July 18. Oregon: Willamette River, Benton Co., May 4, 1961. Washington: Yakima, March 30, 1932.

**Host.**—Adults have been collected from cottonwood.

Biology.—Unknown.

Discussion.—This widespread species is variable in coloration except for the black mesopleuron, white margin of the pronotum, and white clypeus, which appear constant in all the specimens examined. M. wuestneil has the pronotum and clypeus black, and populifolicila and leucostoma have a vellowish-orange mesopleuron. The color variations, such as the brownish areas on the head, are most common in the California specimens, but they appear to be the same species.

#### Messa nana (Klug)

Tenthredo (Allantus) nana Klug, 1814, p. 72; Hartig, 1837, p. 266; Kriechbaumer, 1884, p. 117.

Fenusa nana: Stephens, 1885, p. 41.

Blennocampa nana: Thomson, 1870, p. 285; Thomson, 1871, p. 212; André, 1881, p. 302; Cameron, 1882, p. 254; Brischke and Zaddach, 1883, p. 278; Konow, 1886b, p. 215.

Scolioneura nama: Konow, 1890, p. 249; Dalla Torre, 1894, p. 168; Konow, 1905, p. 85; Enslin, 1914, p. 298; Hering, 1931, p. 109; Conde, 1934, p. 185; Benson, 1936, p. 625; Hering, 1937, p. 74; Benson, 1940, p. 212; Buhr, 1941, p. 911; Wahlgren, 1944, p. 145; Berland, 1947, p. 260; Wahlgren, 1951, p. 74.

Messa nana: Benson, 1941, p. 89; Benson, 1952, p. 107; Lorenz and Kraus, 1957, p. 136; Smith, 1967, p. 280.

Female.—Average length, 4.5 mm. Antenna and head black; labrum and apical segments of maxillary and labial palpi whitish. Thorax black with tegula and posterior margin of pronotum white. Legs with each coxa, trochanter, and femur black; extreme apex of each femur, tibia, and tarsus white. Abdomen black. Wings subhyaline with infuscate band on forewing below stigma; costa and basal parts of veins whitish, stigma and remainder of veins black.

Antenna nine-segmented, slender; eighth segment more than twice as long as broad (pl. III, 36). Head with circular ridge on frons, enclosing anterior ocellus at posterior edge of ridge. Mesonotum without surface sculpture. Sheath straight above, rounded below, with narrow scopa at apex (pl. IV, 49, 50). Serrulae of lancet each low and rounded, with about 10 subbasal teeth and one prominent anterior subbasal tooth (pl. IX, 95).

Male.—Unknown in North America. Benson (1952) has seen

only two males from Britain. His description of the male is as follows: "5 mainly yellow; white are: the tegulae and the outer half of each side of the pronotum; black or piceous are: the vertical and postoccipital parts of the head, middle of pronotum, mesonotum except for sides of front lobes, mesosternum, mesepimeron, most of metathorax, and also the medial basal portions of the basal tergites, though fading out between the 4th and 6th; costa and stigma pale yellow, rest of venation brownish white. 4-4.5 mm."

Larva.—Final feeding stage (pl. XII, 148-153). Length, 9.0 mm. Slightly dorsoventrally flattened. Whitish with head capsule

and thoracic and abdominal plates brownish.

Head dorsoventrally flattened, prognathous. Antenna conical, two-segmented. Labrum as wide as long, truncate; epipharynx with row of 10 to 13 spines on each half; right and left mandibles with dorsal and ventral cutting edges, without distinct teeth, left mandible with one inner tooth; maxillary palpus four-segmented, one seta on second segment; palpifer with one seta; galea long, digitate; lacinia with eight to 10 spines; labial palpus three-segmented.

Single dark plate on pronotum; mesonotum with anterior and posterior transverse narrow plates; metanotum with anterior transverse narrow plate. Prosternum, mesosternum, and metasternum with dark plates, prosternal plate largest. Thoracic legs each five-segmented; tarsal claw present; thoracic legs dark brownish.

Prolegs present on abdominal segments 2 to 8 and 10; dark ring surrounding anterior margin of each proleg; anal proleg with complete dark ring. Sternum of first abdominal segment with small central dark plate; sternum of ninth abdominal segment with two small dark spots and sometimes with another dark spot lateral to each of these. Spiracles winged. Abdominal segments each with two annulets.

The larvae described here were taken from birch in Maine. Lorenz and Kraus (1957) also described the larva.

Holotype.—Probably in the Zoological Museum of Berlin.

Distribution.—Northeastern United States; Europe. Maine: Scarboro, June 4. 1966, ex red oak, L. J. Lipovsky (1 ?); Scarboro, Cumberland Co., ex birches in nursery, June 2, 1967, D. R. Smith (10 ??). New York: Oswego Co., Oswego Tws., from sticky board trap in pear orchard, 6-3-66, R. W. Stelle (2 ??); Albany Co., nr. Rensselaerville, Huyck Preserve, 8 June 1967, R. and J. Matthews (Malaise trap 3) (1 ?).

Host.—A leaf miner of Betula spp.

Biology.—In Maine there is apparently one generation a year. The adults emerge early in the spring, usually the end of May or the first of June, and are found flying about the host at about the same time Fenusa pusilla is active. The female oviposits in the margin of the leaf, similar to Heterarthrus nemoratus, and the larvae mine toward the center. F. pusilla and Profenusa thomsoni both oviposit on the surface near the center of the leaf. On maturing, the nana larvae drop to the soil where they pupate and over-

winter. In this respect nana differs from nemoratus, which remains in the leaf.

Hering (1937), Buhr (1941), and Wahlgren (1944, 1951) pub-

lished notes on the life history of this species in Europe.

Discussion.—This is the most distinct species in this genus and is recognized by its larger size, presence of a ridge on the frons, lack of surface sculpture on the mesonotum, the infuscate band on the forewing, and the more rounded serrulae of the lancet. It was first found in Maine in 1966 and reported by Smith (1967) from Maine and New York. It is probably an introduced species and adds another sawfly leaf-mining pest for Betula, which now has four in North America.

### Messa populifoliella (Townsend)

cottonwood leaf miner, Townsend, 1892a, p. 26. Tineidae? larva, Townsend, 1892b, p. 234.

Blennocampa populifaliella Tovensend, 1893, p. 304.

Messa populifoliella: Burks, 1958, p. 13; Titus and Underwood, 1966, p. 1; Underwood and Titus, 1968, p. 407.

Scolioneura populi Marlatt, 1895, p. 235, 3, 9; Burks, 1958, p. 13 (= populifoliella Townsend)

Messa populi: Ross, 1951, p. 30.

Scotioneura luteopicta Rohwer, 1911d, p. 398, 8; Ross, 1951, p. 30 (= populi Marlatt); Burks, 1958, p. 13 (= populifoliella Townsend).

Female.—Average length, 4.0 mm. Antenna yellow with dorsal surface beyond first segment black. Head yellow with middle fovea, lateral furrows, areas around each ocellus, and sutures of postocellar area black; clypeus white. Thorax yellow with katepimeron, mesonotum except lateral margins of anterior lobe and posterior lateral and anterior mesal margins of lateral lobes, and metanotum black; pectus yellow or black. Legs entirely yellow. Abdomen yellow with most of dorsum black, black decreasing toward posterior; sheath black. Wings subhvaline; veins black with basal parts yellow; stigma and costa light to dark brown.

Antenna nine-segmented; ninth segment of antenna shorter than or subequal in length to eighth segment. Anterior and lateral lobes of mesonotum with fine surface sculpture, sometimes difficult to see. Frons without circular ridge. Sheath straight above. rounded below, with narrow scopa at apex (pl. IV, 51, 52). Each serrula of lancet low and flattened, with one anterior and three or four posterior subbasal teeth (pl. VII, 80).

Male.—Average length, 4.0 mm. In color and structure similar to female. Hypandrium narrowed posteriorly, tapering to blunt point. Genitalia as in plate X, 110 and 111; harpe rounded, oblong; penis valve with long apical filament; parapenis wider than those of wuestneii and leucostoma.

Larva.—Underwood and Titus (1968) described the larva. The later feeding stages have a large pronotal plate, a small transverse mesonotal plate, a large prosternal plate, smaller mesosternal and metasternal plates, a small plate on the sternum of the first abdominal segment, crescent-shaped dark plates at anterior of each proleg, and a large dark plate about the anal proleg. From the

description I am unable to clearly differentiate the larva from other

known Messa larvae, and I have not seen specimens.

Holotypes.—S. populi Marlatt is U.S.N.M. type No. 4032, a female labeled "51" and "Townsend, 1891, New Mexico." S. luteopicta Rohwer is U.S.N.M. type No. 14011, a female labeled "Brookings. S.D." Townsend did not select a type. He (1892b) described the larva, but believed it to be a "Tineid?." Later Townsend obtained adults and described them as a manuscript species of Dr. Riley, Blennocampa populifoliella. Marlatt (1895) described populi from the same locality where Townsend obtained specimens and probably from the same series Townsend examined. Therefore, I have designated a lectotype for Townsend's species from a series collected by Townsend: U.S.N.M. type No. 70217, a female labeled "Las Cruces, N.M."

Distribution.—Central and eastern North America, west to Arizona. Arizona: "Ariz." Colorado: Deric, June 1, 1901, leaf miner on cottonwood, H. G. Dyar. Connecticut: Windsor, 4-1951, James B. Kring. Michigan: Ottawa Co., May 23, 1937, Dreisbach. New Mexico: "N. Mex."; Las Cruces, Apr. 21, 1897, C. J. Ckll. South

Dakota: Brookings, June 20, 1909, H. C. Severin.

Also in Canada from New Brunswick and from eastern Ontario

to the Manitoba border (Underwood and Titus, 1968).

Host.—Leaf miner of Populus spp. Populus fremonti Wats. (Townsend, 1892a), P. cuacuei Simon-Louis, P. nigra var. italica Muenchh., P. tremuloides Michx., and P. balsamifera L. (Underwood and Titus, 1968),

Biology.--Underwood and Titus (1968) gave the life history of this species in New Brunswick. The adults appear during the last 2 weeks of May, and the females oviposit on the margin of the leaf, usually near the tip. The larvae feed for 14 to 20 days, making blotch mines throughout the leaf. On maturing, they drop to the ground and make cells in the soil where they overwinter and pupate. There is one generation a year.

Discussion.—The color pattern makes this species distinctive. The yellow markings of the head, thorax, and abdomen will separate it from other Messa species. I have seen only a few specimens

from widely scattered localities.

# Messa wuestneii (Konow)

Fenusa wuestneii Konow, 1894, p. 91, \$, 9; Konow, 1905, p. 90.

Fenusella wuestneii: Enslin, 1914, p. 305; Malaise, 1920, p. 104; Malaise, 1931, p. 59; Conde, 1937, p. 108; Buhr, 1941, p. 923; Malaise, 1964, p. 33.

Messa wuestneii: Benson, 1941, p. 89; Benson, 1952, p. 108; Lorenz and Kraus, 1957, p. 137; Benson, 1959, p. 90; Benson, 1962, p. 391; Burks, 1967, p. 16.

Fenusa alaskana Kincaid, 1900, p. 345, 3; Konow, 1905, p. 90 (alascana); Benson, 1959, p. 90 (= weestneii Konow).

Messa alaskana: Ross, 1951, p. 30.

Female.—Average length, 3.7 mm. Antenna and head black; ventral surface of antenna and labrum sometimes brownish. Thorax black with tegula white; posterior angles of pronotum sometimes with touch of white. Legs black with extreme base of

each femur, front tibia, and apices of middle and hindtibiae whitish; each tarsus infuscated. Abdomen black, sometimes with narrow white margin on posterior edge of each segment. Wings lightly, uniformly infuscated.

Antenna nine-segmented; ninth segment shorter than or subequal in length to eighth segment. Circular ridge on frons absent. Front and lateral lobes of mesonotum with fine surface sculpture. Sheath short, straight above, rounded below, with narrow scopa at apex (pl. IV, 51, 52). Each serrula of lancet low and flattened, with one anterior and three or four posterior subbasal teeth (pl. VII, 81).

Male.—Average length, 3.7 mm. In color and structure similar to female. Genitalia as in plate X, 114 and 115; harpe oblong, rounded at apex; parapenis narrow; penis valve with long apical filament.

Larva.—Malaise (1920), Buhr (1941), and Lorenz and Kraus (1957) described the larva of this species. I have not seen specimens.

Holotypes.—F. alaskana Kincaid is U.S.N.M. type No. 5281, a male labeled "Kukak Bay, Alaska, July 4, '99, Harriman Expedition '99, T. Kincaid, collector." Konow's type is probably at the Deutsches Entomologisches Institut, Eberswalde, Germany.

Distribution.—Western North America from Alaska to California and Colorado; Europe. Alaska: Kukak Bay, July 4, '99; Anaktuvuk Pass, on willow, 7-14-68. British Columbia: Vancouver, April 21, 1931. California: San Francisco, March 3, 1927, March 12, 1932, on Salix lasiolepis: Carnelian Bay, Placer Co., June 27, 1965, Salix lemmonii; 1 mi. W. Hobart Mills, Nevada Co., May 13, 1966, Salix lemmonii, S. geveriana; Lassen Co., McCoy Flat Res., 31 mi. N.W. Susanville, June 15, 1966, Salix lemmonii bebb, complex; Willows. Colorado: "Colo."; Green Mt. Falls, June 14, 1938. Nevada: Mt. Rose Pass, 11 mi. S.W. Reno, Washoe Co., July 10, 1965, Salix lemmonii; same data, June 27, 1965; same data, July 18, 1965; same data, July 7, 1965.

Host.-Leaf miner of Salix spp.

Biology.—Malaise (1931) and Buhr (1941) described the life history of this species in Europe; there is no North American literature on its life history.

Discussion.—This small species may be separated from other Messa species by the almost entirely black coloration. It is a Holarctic species and was recorded from North America by Benson (1959).

# Genus SETABARA Ross

Parabates MacGillivray, 1909a, p. 262; Benson, 1936, p. 625 (= Fenusella Enslin); Ross, 1937, p. 70 (= Scolioneura Konow). Preoccupied.
 Type-species: Parabates histrionicus MacGillivray. Original designation.
 Setabara Poss, 1951, p. 31 (new name for Parabates MacGillivray); Smith, 1967, p. 281.

**Description.**—Antenna stocky; second segment longer than broad; third segment longer than fourth segment; segments beyond third subequal in length. Clypeus truncate; malar space linear; postgenal carina absent. Prepectus absent. Tarsal claw simple with blunt inconspicuous basal lobe (pl. III, 24). Forewing with stub of vein 2A and 3A curved up at apex. Hindwing with cells Rs and Rs and Rs and Rs and Rs and Rs consequence (similar to pl. II, 13).

The closed radial cell of the hindwing and inconspicuous basal lobe of the tarsal claw will distinguish this genus from other North American genera. The single species of this genus is known only from western United States and is probably associated with

Prumus.

### Description of Setabara Species

Setabara histrionica (MacGillivray)

Parabates histrionicus MacGillivray, 1909a, p. 262, 9; Frison, 1927, p. 255. Scolioneura histrionicus: Ross, 1937, p. 71. Setabara histrionicus: Ross, 1951, p. 31; Smith, 1967, p. 281.

Female.—Average length, 4.0 mm. Black with labrum brownish and extreme apex of each femur, extreme base of middle and back tibiae, and front tibia whitish. Wings lightly infuscated.

Front and lateral lobes of mesonotum with fine surface sculpture. Sheath short, straight above, rounded below, with small scopa at apex (pl. IV, 60, 61). Lancet with each serrula low, broad, truncate at apex, without distinct anterior or posterior subbasal teeth (pl. VII. 83).

Male.—Average length, 4.0 mm. In color and structure similar to female. Genitalia as in plate X, 116 and 117; harpe long, inner apical margin flat; penis valve rectangular, with short apicolateral spine.

Larva.—Unknown.

Holotype.—U.S.N.M. type No. 12177, a female labeled "Colo." Distribution.—Western North America. California: Hambone, Shasta N.F., 5-8-34. Prunus sp. Colorado: "Colo." Idaho: Lenore, 1000', May 19, 1937. Nevada: "Nev." Oregon: Talent, Apr. 19, 1960, on Prunus; Talent, April 11, 1935; Klamath Falls, 5-11-64, Prunus subcordata: Boyer, May 15, 1937. Washington: Almota, 4-17.

Host.—Adults have been collected from Prunus sp.

Biology.—Unknown.

**Discussion.**—This is not a well-known species, apparently closely related to some *Messa* species but differing by the small basal lobe of the tarsal claw and lack of a long filament on the penis valve of the male genitalia.

# Genus PROFENUSA MacGillivray

Profesusa MacGillivray, 1914, p. 364; MacGillivray, 1916, p. 156; Ross, 1937,
 p. 72; Benson, 1941, p. 86; Ross, 1951, p. 30; Benson, 1952, p. 108;

Lorenz and Kraus, 1957, p. 137; Malaise, 1964, p. 33; Smith, 1966, p. 719; Smith, 1967, p. 280.

Type-species: Profemusa collaris MacGillivray. Monotypic. Gunnea Malaise, 1964, p. 33. New synonymy.

Type-species: Tenthredo pygmaca Klug. Original designation.

**Description.**—Second antennal segment longer than broad; third segment longer than fourth segment. Clypeus truncate; postgenal carina absent; malar space less than diameter of front ocellus. Prepectus absent. Tarsal claw with one long outer tooth and large acute basal lobe (pl. III, 22). Forewing with stub of vein 2A and 3A straight at apex. Hindwing with cell  $R_1$  open; anal cell present or absent, if present then cell is longer than its petiole (pl. II, 14–16).

The open radial cell of the hindwing, the straight 2A and 3A vein of the forewing, the single outer tooth and basal lobe of the tarsal claw, and the absence of a prepectus will distinguish this genus.

About eight species are in this genus, five of which are found in North America. The North American species are associated with Betula, Crataegus, and Quercus.

Malaise (1964) established the genus *Gunnea* and separated it from *Profenusa* only on the basis of the presence of an anal cell in the hindwing. This is a variable character in *Profenusa* and cannot be valid for separation of species that are so closely related in habits and larval and other adult morphological characters.

Larvae are known for all five species of *Profemusa*. All species, except canadensis, are very distinct and conform to a similar pattern. The larvae of lucifex, inspirata, thomsoni, and alumna are distinctly dorsoventrally flattened, lack prolegs, have very reduced thoracic legs, lack tarsal claws (except very minute tarsal claws in lucifex), have three small spines or tubercles on the coxa of each thoracic leg, and have a minutely spiniform integument. The larva of canadensis has prolegs, well-developed thoracic legs with tarsal claws, lacks the spines on the coxae, and has a smooth integument. The larva of canadensis may be confused with those of Messa or Metallus, but canadensis may be separated by the presence of prolegs on abdominal segments 2 to 8 and 10 and the lack of dark plates on the mesosternum, metasternum, and sternum of the first thoracic segment. The larva of canadensis is as in plate XVI, 201. The larvae of the other four species of Profesusa are similar in general habitus, as in plate XV, 181.

# Keys to Profenusa Species

#### ADULTS

1.	Female	2
2.	Thorax, either part of mesonotum, upper part of mesepisternum, o prothorax, rufous; clyneus usually white	ייי
	Thorax black; clypeus black	4

3. Mesonotum, except scutellum, rufous; serrulae of lancet lobelike (pl. VIII, 90); leaf miner of Cratacyus canadensis (Mar Mesonotum black; serrulae of lancet pointed at apex (pl. VIII, 94); canadensis (Marlatt) lenf miner of Quercus lucifex (Ross)

4. Hindtibia black; California and Oregon; leaf miner of Quercus inspirata (MacGillivray)

Hindtibia white; eastern species Hindwing without anal cell; serrulae of lancet with eight or nine small posterior and equal number of anterior subbasal teeth (pl. VIII, 91); sheath rounded below (pl. IV, 57); leaf miner of Retula: thomsoni (Konow) Hindwing with anal cell; serrulae of lancet with two or three anterior

subbasal teeth and five or six posterior subbasal teeth (pl. VIII, 92); sheath subtruncate at apex (pl. IV, 56); leaf miner of Quercus alumna (MacGillivray)

Abdomen, mesopleuron, and pectus orange yellow canadensis (Marlatt) Entirely black

Western species, California and Oregon Eastern species; rare

inspirata (MacGillivray) alumna (MacGillivray)

#### LARVAE

1. Prolegs present; tarsal claw distinct; sternal plates, except for that on prosternum, absent; labrum wider than long, emarginated; coxae without spines; leaf miner of Crataegus (pl. XVI, 201-206)

canadensis (Marlatt) Prolegs absent; tarsal claw absent or very minute; mesosternal and metasternal plates sometimes present; labrum longer than wide, truncate; each coxa with three small spines or tubercles 2

2. Mesosternum and metasternum each with central dark plate; thoracic legs two-segmented; leaf miner of Betula (pl. XV, 181-814)

thomsoni (Konow) Metasternum without plate; mesosternum with or without plate; thoracic legs with more than two segments; leaf miners of Quer-

Thoracic legs each with minute tarsal claw (pl. XVI, 200) lucifex (Ross) Tarsal claw absent

4. Mesosternal plate present; thoracic legs four-segmented; western species, California and Oregon (pl. XIV, 178, 179)

inspirata (MacGillivray) Mesosternal plate absent; thoracic legs three-segmented; castern species (pl. XV, 189, 190)

alumna (MacGilliv) \_alumna (MacGillivray)

# Descriptions of Profenusa Species

Profenusa alumna (MacGillivray)

Messa alumna MacGillivray, 1923b, p. 23, 9; Frison, 1927, p. 251; Benson, 1959, p. 91 (= thomsoni Konow [1]).

Fenusa alumna: Ross, 1936, p. 265.

Profenusa alumna: Ross, 1951, p. 30. Fenusa curta of auett., nec Norton, Dyar, 1895b, p. 343; Dyar, 1898, p. 137 (?).

Messa amica MacGillivray, 1923b, p. 23, 4, Frison, 1927, p. 251; Ross, 1936, p. 265 (= alumna MacGillivray).

Messa appota MacGillivray, 1923b, p. 24, &; Frison, 1927, p. 251; Ross, 1936, p. 265 (= alumna MacGillivray). Profenusa mainensis Smith, 1966, p. 720, Q, larva. New synonymy.

Female.—Average length, 4.0 mm. Black with each tibia and tarsus whitish. Wings infuscated, slightly darker toward base. Hindwing with anal cell present. Sheath straight above and below, subtruncate at apex (pl. IV, 56), not as rounded as that of thomsoni (pl. IV, 57). Serrulae of lancet low, far apart, each serrula with two or three anterior and five or six posterior subbasal teeth; clumps of lateral spines located centrally on segments 3 to 11 (pl. VIII, 92).

Male.—Average length, 4.0 mm. In color and structure similar to female. Genitalia as in plate X, 120 and 121; penis valve oblong, without lobes, with two small spines near apex, one directed dorsally, the other ventrally.

Larva.—Final feeding stage (pl. XV, 189–196). Length, 7.5 mm. Body dorsoventrally flattened; integument covered with minute rounded spines. White to light brown with pronotal plate, prosternal plate, head capsule, thoracic legs, and spiracles dark brown; usually black spot on each side of head. In lateral view similar to thomsoni (pl. XV, 181).

Head dorsoventrally flattened; prognathous. Antenna with one segment; ocularium small; eyespot conspicuous, located posterior to antenna. Labrum longer than wide, truncate; epipharynx with row of eight or nine spines on each half and row of shorter spines anterior to these; maxillary palpus four-segmented, one seta on second segment; galea long, digitate, subequal in length to maxillary palpus; lacinia with five stout spines; each mandible similar, right mandible with several irregular ventral and dorsal teeth, left mandible with several irregular ventral teeth, rounded dorsal cutting edge, and one inner tooth; each mandible with one seta on outer basal angle; labial palpus two-segmented.

Thoracic legs small, directed laterally, three-segmented with apical segment minute; tarsal claw absent; each coxa with three small tubercles. Prothoracic spiracle winged. Pronotal and prosternal plates present.

Prolegs absent. Abdominal spiracles winged, located dorsal to pleural lobes.

Smith (1966) described the larva of this species, and those references to *curta* by Dyar (1895b, 1898) probably pertain to the larva of this species.

Holotypes.—MacGillivray's types are at the Illinois Natural History Survey: M. alumna, a female, is labeled "N. Ill."; M. amica, a female, is labeled "N. Evans, N.Y., 8-2-08, M. C. V. Goll."; M. appota, a male, is labeled "Buffalo, N.Y., 6-27-09, M.C.V. Coll." P. mainensis Smith is U.S.N.M. type No. 67982, a female labeled "Canton, Maine, May 30 1964, ovipositing on red oak leaves, G. LaBonte."

Distribution.—Northeastern United States. Illinois: "N. Ill."; Algonquin. Maine: Many adults from the following localities collected during the last of May and in June: Augusta. Gilead. Canton, South Paris, Peru, Strong. Maryland: Plummers Island, 23-7-16. New York: N. Evans, 8-2-08; Buffalo, 6-27-09. Pennsylvania: Dupont, Aug. 14-16, 1944. Virginia: Falls Church, Aug. 12, 1912, Quercus, July 15, 1912.

Host.—Most of the damage in Maine is on red oak (Quercus rubra L.). It may also attack Q. macrocarpa Michx., Q. alba L., Q.

velutina Lam., and Q. ilicifolia Wangh, Dyar (1895b) found the

larva on Q. macrocarpa.

Biology.—This species has done considerable damage to oaks in Maine where the leaf-mining larvae cause severe browning and blotching of the leaves. The adults fly in spring and early summer, from the end of May to the first of July, and oviposit in the upper surface of the leaf. When the larvae mature, they drop to the ground to overwinter and pupate in the soil. Larvae have been observed on the foliage until mid-August. This species is apparently parthenogenetic with very rare males. Among the long series from Maine, males have never been seen; however, males have been taken in other parts of its range—in Illinois, New York, and Virginia.

Discussion.—The general habitus of this species closely resembles that of thomsoni, but the presence of the anal cell in the hindwing, the more truncate sheath, and small serrulae of the lancet will separate alumna. The larvae are separated from those of other Profenusa species by the absence of mesosternal and metasternal plates, absence of prolegs, and absence of tarsal claws.

The close similarity of alumna to thomsoni led to the synonymy of these two species by Benson (1959) and subsequent description of a new species, mainensis, by Smith (1966). The examination of the type of alumna proved that this synonymy was a mistake and that alumna is actually the oak leaf miner, probably native to North America, whereas thomsoni is undoubtedly an introduced species. The references to alumna by Burks (1958), Lindquist (1959), Watson (1959), or any other literature treating alumna as a birch leaf miner do not pertain to alumna (MacGillivray) but actually to thomsoni (Konow).

# Profenusa canadensis (Marlatt)

Scolioneura canadensis Marlatt, 1895, p. 235, Q. Metallus canadensis: MacGillivray, 1916. p. 160.

Professis anadensis: MacGillivray, 1916, p. 160.
Professis canadensis: Ross, 1937, p. 72; Hamilton, 1943, p. 59; Hamilton, 1950, p. 694; Ross, 1951, p. 30; Peterson, 1956, p. 270; Malaise, 1964, p. 28; Martineau, 1965, p. 46; Smith, 1966, p. 719.
Professis acollaris MacGillivray, 1914, p. 364, 3, 2; Parrott and Fulton, 1915a, p. 551; Parrott and Fulton, 1915b, p. 519; MacGillivray, 1916, p. 157; Frison, 1927, p. 258; Ross, 1937, p. 72 (= canadensis Marlatt).

Female.—Average length, 4.0 mm. Antenna black, sometimes with ventral surface whitish; head black, clypeus and mouth parts white. Thorax black with mesonotum, except scutellum, rufous; mesopleuron and pectus rufous or black; prothorax usually rufous; tegula and posterior margin of pronotum white. Legs white to yellowish orange, usually with hindcoxa and hindfemur darker or black. Abdomen black; each segment with narrow white band on posterior edge. Wings hyaline.

Hindwing with anal cell present. Sheath short, rounded at apex (pl. IV, 53). Each serrula of lancet deep, lobelike, rounded at apex,

without subbasal teeth (pl. VIII, 90).

Male.—Average length, 4.0 mm. Antenna black, sometimes with ventral surface whitish; head black, clypeus and mouth parts white.

Thorax yellowish orange with mesonotum and metanotum black. Legs entirely yellowish orange. Abdomen entirely yellowish orange, sometimes with black or infuscate areas on basal terga or black longitudinal line on dorsum. Structure as for female. Genitalia as in plate X, 118 and 119; penis valve with long ventral lobe.

Larva.—Final feeding stage (pl. XVI, 201–206). Length, 7.5 mm. Cylindrical, slightly dorsoventrally flattened. White; head capsule, spiracles, thoracic legs, prosternal plate, and crescent-shaped marks about each proleg light brown.

Head dorsoventrally flattened, nearly hypognathous. Antenna conical, indistinctly three-segmented. Eyespot posterior to antenna. Labrum wider than long, emarginated; epipharynx with row of seven to nine long spines on each half; right mandible with two ventral teeth and two dorsal teeth; left mandible with three ventral teeth and two dorsal teeth; each mandible with one seta on outer basal angle; maxillary palpus four-segmented, one seta on second segment; galea digitate, subequal in length to palpus; lacinia with seven to nine long spines; labial palpus three-segmented, relatively long and distinct compared to that of other *Profenusa* species.

Thoracic legs five-segmented, directed laterally; tarsal claw present. Pronotal and mesonotal plates light colored; prosternal plate light colored; mesosternal and metasternal plates absent. Prothoracic spiracle not winged. Coxa of each thoracic leg without small spines or tubercles.

Prolegs present on abdominal segments 2 to 8 and 10, rudimentary on segments 1 and 9. Spiracles of abdomen not winged. Pleural lobes present. Abdominal segments 1 to 8 each with three annulets. Light crescent-shaped plates present at anterior of each proleg.

The larva has been described by Parrott and Fulton (1915b), Hamilton (1950), and Peterson (1956).

Holotypes.—S. canadensis, a female labeled "Can.," is type No. 10222 at the Academy of Natural Sciences of Philadelphia. It also bears a label "Type No. 4031 USNM." There is a specimen with the same data in the U.S. National Museum collection; however, it cannot be the type, since Marlatt (1895) stated that the type was in the collection of the American Entomological Society. P. collaris MacGillivray, a female labeled "cherry, 5/4/11, Geneva,

N.Y.," is at the Illinois Natural History Survey.

Distribution.—Widespread in eastern North America. Arkansas: Bentonville, 4-14-19, Crataegus. District of Columbia: Apr. 3, 1945, on Crataegus. Illinois: Muncie, V-1-27, May 4, 1936; White Heath, May 1, 1915, May 5, 1940, April 30, 1916: Monticello, May 4, 1947; St. Joseph. May 3, 1914; Oakwood, May 4, 1936. Iowa: Shenandoah, 4-21-46. Kentucku: Lexington, Apr. 17, 1919, hawthorn. Maine: Augusta, emgd. 23-VI-56, ex Crataegus; Brownville Junction. June 9, 1967. Massachusetts: Boston, May 10, 1909; Forest Hills, V-3-12. Michigan: East Lansing. May 17, 1963. Missouri: Columbia. New York: Geneva. 5-20-15; Greendale, V-11-39. soil under cherry tree; Oswego, 6-3-66; Ithaca; Nassau, May 27, 1917. Ontario: Manotick, 20-V-66, Crataegus sp. Penn-

sylvania: Harrisburg, V-11-20, on foliage of Crataegus sp.; Chestnut Hill, 5-4-25. Quebec: Ste. Foy. Texas: Longview, III-26-08.

Hosts.—Leaf miner of Crataegus spp. and Prunus sp. This species is apparently most destructive to hawthorn. Parrott and Fulton (1915a, 1915b) recorded a population on cultivated cherry, the English Morello variety, but other varieties of cherry were not subject to attack.

Biology.—Parrott and Fulton (1915b) and Hamilton (1950) published the life history of this species. The adults emerge early in the spring, usually during April and May. The females oviposit near the margin of the leaf, but not on the margin nor near the center. The larva mines toward the center of the leaf leaving blotch mines. On maturing, the larvae drop to the ground and form earthern cells in which they overwinter and pupate the following spring. There is one generation a year.

Discussion.—The rufous mesonotum of the female and yelloworange coloration of the male will separate this species from other Profesusa species. The genitalia of both sexes are also distinct. This species is of occasional economic importance, especially in

ornamental plantings of Cratacgus.

# Profenusa inspirata (MacGillivray)

Parabates inspiratus MacGillivray, 1909a, p. 264, Q. Fenusa inspirata: Ross, 1936, p. 265. Profenusa inspirata: Ross, 1951, p. 30; Smith, 1966, p. 719.

Female.—Average length, 4.0 mm. Black with each front and middle tibia and tarsus and hindtarsus whitish. Wings uniformly, lightly infuscated.

Hindwing with anal cell present. Sheath straight above, rounded below (pl. IV, 57). Each serrula of lancet pointed, finely serrate, with greater number of posterior than anterior subbasal teeth; lateral spines on segments lacking (pl. VIII, 93).

Male.—Average length, 4.0 mm. In color and structure similar to female. Genitalia as in plate X, 122 and 123; penis valve oblong, without lobes; two short spines present near apex, one directed

dorsally, the other ventrally.

Larva.—Final feeding stage (pl. XIV, 174-180). Length, 7.5 mm. Dorsoventrally flattened; integument covered with minute rounded spines. White to light brown with head capsule, pronotal plate, prosternal plate, mesosternal plate, thoracic legs, spiracles, and spots surrounding abdominal spiracles dark brown. In lateral view similar to larva of thomsoni (pl. XV, 181).

Head dorsoventrally flattened, from above similar to that of alumna (pl. XV, 192). Antenna apparently with small second segment; eyespot located posterior to antenna; antennaria small. Labrum longer than wide; epipharynx with row of eight or nine spines on each half with row of smaller spines anterior to these; right and left mandible similar, each with three ventral teeth and rounded dorsal cutting edge; maxillary palpus four-segmented,

one seta on second segment; galea long, digitate, subequal in length to maxillary palpus; lacinia with four stout spines; labial palpus

two-segmented.

Thoracic legs reduced, directed laterally, apparently four-segmented with apical segment minute; tarsal claw absent; each coxa with three rounded tubercles. Large pronotal and prosternal and small mesosternal plates present. Prothoracic spiracle winged.

Abdominal prolegs absent. Annulation indistinct. Abdominal spiracles winged. Small darkened spots surrounding each spiracle. The larvae are described from specimens taken from valley oak,

Merced, Calif., on July 27, 1959.

Holotype.—U.S.N.M. type No. 12176, a female labeled "Nev."

Distribution.—Pacific coastal region of North America. California: 4 mi. W. Quincy, Plumas Co., VI-19-49; Quincy, July 1, 1958; Yosemite, 3880'-4000', 6-12-28; Hatchet Mtn., Shasta Co., 7-14-33, adults on Alnus sp. Nevada: "Nev." Oregon: Corvallis, May 10, 1931; Touvelle St. Pk., 8 mi. N. Medford, May 21, 1960; Jackson Co., Buckhorn Mineral Springs, 11 mi. E.S.E. Ashland, 2800', May 19, 1960; Jackson Co., 1 mi. E. Brownsboro, May 18, 1962; Wasco Co., 3 mi. E. Mosier, May 23, 1959; Talent, May 20, 1959, on Quercus garryana; Medford, V-5-53; Klamath Co., 10 mi. S.W. Keno, May 23, 1962, on Garry oak; Jackson Co., 1 mi. N. Sams Valley, May 24, 1964, on oak leaves.

Host.—Leaf miner of Quercus spp. Adults have been taken from Q. garryana Dougl. and Alnus sp., and larvae were taken from

valley oak (Q. lobata Née).

**Biology.**—No information is available.

Discussion .- This is the only known western species of Profenusa. It is similar to thomsoni and alumna, but it may be distinguished by the dark hindtibia and characters of the male and female genitalia.

# Profenusa lucifex (Ross)

Fenusa lucifex Ross, 1936, p. 266, 9.
Profenusa lucifex: Ross, 1951, p. 31; Lindquist and Jackson, 1965, p. 1;

Smith, 1966, p. 719.

Female.—Average length, 4.0 mm. Head and antenna black; clypeus and mouth parts white to yellow orange. Thorax black with pronotum, tegula, and upper half or less of mesepisternum rufous to yellow orange. Legs entirely yellow orange. Abdomen black. Wings lightly, uniformly infuscated.

Hindwing with anal cell open, vein 2A and 3A partially atrophied. Sheath straight above, rounded below (pl. IV, 57). Lancet with each serrula pointed, central serrulae each with about 10 fine anterior and 10 fine posterior subbasal teeth (pl. VIII, 94).

Male.—Unknown.

Larva.—Final feeding stage (pl. XVI, 197-200). Very similar to larva of alumna except for following differences: Antenna slightly longer with small second segment; spots on each side of head larger and darker; tarsal claw present, very small, and located on anterior side of tarsus; thoracic legs four-segmented; mesosternum sometimes with narrow dark plate on posterior edge.

The prepupa may be distinguished from that of alumna by the

presence of a tarsal claw.

Holotype.—At the Illinois Natural History Survey, a female

labeled "Fox Lake, Ill., June 30, 1935, DeLong and Ross."

Distribution .- Northeastern United States and southeastern Canada. Illinois: Fox Lake, June 30, 1935. Maine: Augusta. emerged April 18, 1966, from larva on white oak; Augusta, July 9, 1965, emgd. April 8, 11, 1966, bred ex white oak; Berwick, June 1, 1966, sweeping under red oak; Portland, Baxter Park, June 11, 1967. New York: Orient L.I., June 16, 1947; Riverhead, L.I., August 24, 1934. Ontario: Porest districts of Kenora and Fort Frances (Lindquist and Jackson 1965).

Host.-Leaf miner of Quercus spp. Most common on white oak (Quercus alba L.) and also on red oak (Q. rubra L.) in Maine. Lindquist and Jackson (1965) reaced this species from bur oak

(Q. macrocarpa Michx.) in western Ontario.

Biology.—The note by Lindquist and Jackson (1965) is the only information available on the life history of this species. In western Ontario the larvae complete their feeding in late July or early August and drop to the ground to overwinter in the soil. The larvae make irregular blotch mines in the leaves. In Maine the adults appear in June.

Discussion.—This species received little attention from the time of the description in 1936 until 1965, when it was discovered damaging oaks in Ontario and Maine. It may be distinguished from other Profesusa species by the rufous pronotum and partly rufous mesepisternum. Some specimens have the upper half of the mesepisternum rufous, other specimens only the dorsal margir.

# Profenusa thomsoni (Konow)

Fenusa thomsoni Konow, 1886b, p. 270, 9; Konow, 1890, p. 249; Konow, 1905,

Fennsella thomsoni: Enslin, 1914, p. 306; Dovnar-Zapolsky, 1931, p. 56; Conde, 1934, p. 186; Hering, 1937, p. 75; Buhr, 1941, p. 912; Wahlgren,

1951, p. 75.

 Profemusa thomsoni: Benson, 1941, p. 88; Benson, 1943, p. 11; Benson, 1952, p. 108; Lorenz and Kraus, 1957, p. 137; Benson, 1959, p. 91; Forbes, Underwood, Cuming, and Eidt, 1960, p. 29; Martin, 1960, p. 376; Pierenek, 1960, p. 376; 1962, p. 285; Benson, 1962, p. 392; Smith, 1966, p. 719; Burks, 1967, p. 16; Benson, 1968, p. 149.

Profenusa alumna of auett., nec MacGillivray, Burks, 1958, p. 13; Lindquist,

1959, p. 626; Watson, 1959, p. 618.

Female.-Average length, 4.0 mm. Black with each tibia and

tarsus whitish. Wings infuscated, darker on basal half.

Hindwing with anal cell absent. Sheath straight above, rounded below (pl. IV, 57). Serrulae of lancet wide, close together, central serrulae each with six or seven anterior and six or seven posterior subbasal teeth; apical serrulae each with four or five anterior and six or seven posterior subbasal teeth; group of lateral spines present on segments 4 to 9 (pl. VIII, 91).

Male.—Unknown. Parthenogenetic.

Larva.—Final feeding stage (pl. XV, 181–188). Length, 7.0 mm. Body dorsoventrally flattened; integument covered with minute spines, venter of 10th abdominal segment smooth. White to light brown with head capsule, pronotal, prosternal, mesosternal, and metasternal plates, thoracic legs, and spiracles dark brown.

Head dorsoventrally flattened, prognathous. Antenna one-segmented; antennaria small; eyespot posterior to antenna. Labrum longer than wide, truncate; epipharynx with row of seven to nine spines on each half; left and right mandibles similar except for inner tooth of left mandible, ventral margin of each mandible with four or five fine teeth, dorsal margin without teeth; one seta on outer basal angle of each mandible; maxillary palpus four-segmented, second segment with one seta; galea long, digitate, subequal in length to maxillary palpus: lacinia with five or six stout spines; labial palpus two-segmented.

Thorax with large dark pronotal and prosternal plates and small mesosternal and metasternal plates. Prothoracic spiracle winged. Thoracic legs two-segmented, tarsal claw absent; each coxa with

three short spines.

Abdominal prolegs absent. Abdominal spiracles winged, located dorsal to pleural lobes. Segments indistinctly three-annulate.

Watson (1959) described the larva of this species in detail, and Lindquist (1959) gave characters to separate it from other birch leaf-mining sawfly larvae.

Holotype.—Probably at the Deutsches Entomologisches Institut,

Eberswalde, Germany.

Distribution.—Northeastern United States and southeastern Canada; Europe. Connecticut: Hartland, July 25, 1930; Hamden, July 5, 1926. Maine: Canton, 21-VII-64, white birch; Gilead, July 7, 1965, white birch; Brunswick, many dates from June 23 to July 16, 1965. Ontario: Sault Ste. Marie, 4-VII-55, white birch. Quebec: Routhierville, July 2, 1966. Vermont: Stratford, IV-19-1962.

**Host.**—Leaf miner of *Betula* spp. Martin (1960) recorded *B.* papyrifera Marsh., *B. alleghaniensis* Britt. (= B. lutea L.), and *B.* populifolia Marsh.

Biology.—Martin (1960) presented the life history of this species. In Ontario the adults emerge from the middle of July to the first of August. The females oviposit near the center of the leaf, usually near a vein, with very few eggs placed near the margin of the leaf. The larvae form typical blotch mines while feeding. On maturing, the larvae drop to the ground and overwinter in the soil. In Maine the adults begin emerging during the last part of June.

**Discussion.**—This species is separated from alumna by the absence of an anal cell in the hindwing, more rounded sheath, and larger serrulae of the lancet. The larvae have mesosternal and metasternal plates, both absent in alumna larvae. This is one of the four sawfly leaf miners of Betula known in North America.

As explained under alumna, the synonymy of alumna with

thomsoni by Benson (1959) was a mistake, and caution should be used when referring to distributional and taxonomic data in those publications that report "Profenusa alumna (MacGillivray)" as a leaf miner of Betula. The distributional data by Watson (1959), for example, are not entirely correct. P. thomsoni is probably an introduced species.

# Genus BIDIGITUS Smith

Bidigitus Smith, 1967, p. 280.

Type-species: Profenusa platani Burks. Original designation.

Description.—Antenna with second segment longer than broad; third segment longer than fourth segment; segments beyond third gradually decreasing in length. Clypeus truncate; malar space less than half diameter of front ocellus; postgenal carina absent. Prepectus absent. Tarsal claw with two long subequal outer teeth and large acute basal lobe (pl. III, 23). Forewing with stub of vein 2A and 3A faint, curved up at apex; vein 1m-cu strongly directed downward meeting Cu, closer to vein Cu,a than to crossvein cu-a. Hindwing with cell  $R_i$  open at apex; anal cell present, petiole much longer than cell; cells Rs and M absent (pl. II, 17, 18).

This genus is close to Profenusa, but it is separated by the bifid tarsal claw and wing venation. The single species in this genus is known only from California and is associated with Platanus.

# Description of Bidigitus Species

Bidigitus platani (Burks)

Profenusa platanae Burks, 1957, p. 207, S, Q, larva; Brown and Eads, 1965, pp. 14, 22-32.

Profesusa platoni: Smith, 1966, p. 719; Burks, 1967, p. 16. Bidigitus platani: Smith, 1967, p. 280.

Female.—Average length, 4.0 mm. Black with tegula, extreme apex of each femur, each front and middle tibia, basal two-thirds of hindtibia, and each tarsus whitish. Wings uniformly, lightly infuscated.

Sheath long and narrow, pointed at apex (pl. IV, 47). Serrulae of lancet far apart, each low, with two anterior and three or four

posterior subbasal teeth (pl. VII, 85).

Male.—Average length, 4.0 mm. In color and structure similar to female. Hypandrium truncate. Genitalia as in plate X, 128 and 129; harpe long and narrow; parapenis rounded, about as wide as long; penis valve subrectangular with apical and dorsoapical lobes.

Larra.—Final feeding stage (pl. XVII, 207-212). Length, 9.0 mm. Slightly dorsoventrally flattened. White; head capsule and legs light brownish. In dorsal view slightly widened at thorax and narrowing toward apex of abdomen.

Head dorsoventrally flattened, prognathous. Antenna two-segmented, second segment small; antennaria large; eyespot posterior to antenna. Labrum wider than long, emarginated; epipharynx with row of nine to 10 spines on each half; left mandible with large apical tooth, two small ventral teeth, and three larger dorsal teeth; right mandible similar to left mandible but with two dorsal teeth; each mandible with one seta on outer basal angle; maxillary palpus four-segmented, one seta on second segment; galea long, digitate, subequal in length to palpus; lacinia long, with about seven spines of unequal length all directed mesally; labial palpus threesegmented.

Thoracic legs five-segmented, directed laterally, relatively long and segments distinct: tarsal claw present. Prosternum with large smooth plate of similar color as rest of body, smaller plates located

centrally on mesosternum, metasternum, and pronotum.

Prolegs present on abdominal segment 2 to 8 and 10; rudimentary on ninth segment. Abdominal segments 1 to 7 each twoannulate, first annulet one-third size of second annulet. Conspicuous pleural lobes present.

Burks (1957) and Brown and Eads (1965) also described the

larva.

Holotype.—U.S.N.M. type No. 63460, a female labeled "Santa Barbara, Calif., May 20, 1956, reared from Platanus racemosa, Clark O. Eads."

Distribution.—Known only from California. California: The only adult record is that of the type locality. Larvae have been taken from the following: Anaheim. Apr. 20, 1962, on *Platanus*; Lakeport. V-16-1952, ex plane tree (leaf blisters); Claremont, April 24, 1961, Platanus racemosa. Brown and Eads (1965) stated that this species is "very common throughout southern California, as well as the rest of California."

Host .- Leaf miner of the California plane tree (Platanus race-

mosa Nutt.).

Biology.—Brown and Eads (1965) described the life history of this species. The adults appear in February and oviposit in the upper surface of the leaf. The larvae feed for about 30 days making blotch mines in the leaves. Up to 95 larvae have been found in a single leaf. On maturing, the larvae drop to the soil and form small cells lined with a tough papery material in which they pupate or overwinter. There are from three to five generations a year, and 7 to 11 weeks are required to complete a life cycle.

Discussion.—The damage caused by this species has been known for a long time in California, but it was believed to be caused by a lepidopterous blotch miner. In 1956 the damage was first recognized as being caused by a sawfly, and the sawfly was described as new in 1957 by Burks. The tarsal claw and wing venation will

easily distinguish this species from those of related genera.

#### Genus NEFUSA Ross

Nefusa Ross, 1951, p. 31; Benson, 1959, p. 92; Smith, 1967, p. 281. Type-species: Messa unita MacGillivray. Original designation.

**Description.**—Antenna with second segment longer than broad; third segment longer than fourth segment; segments beyond third subequal in length. Clyneus truncate; malar space linear; postgenal carina absent. Prepectus present, narrow, set off by suture. Tarsal claw with one outer tooth and large acute basal lobe (pl. III, 22). Forewing with stub of vein 2A and 3A straight at apex. Hindwing with cells Rs and M absent; anal cell present; cell  $R_1$  open (similar to pl. II, 14, 15).

The presence of a prepectus will separate this genus from all other genera of North American Fenusini. The single species known for this genus is a leaf miner of Viola in eastern North

America.

### Description of Nefusa Species

Nefusa ambigua (Norton), new combination

Fenusa ambiguus Norton, 1867, p. 225, &; Dalla Torre, 1894, p. 156; Konow, 1905, p. 90.

Messa ambigua: MacGillivray, 1916, p. 157.

Messa anita MucGillivray, 1923b, p. 23, 9; Frison, 1927, p. 251. New

synonymy.

Fenusa anita: Ross, 1936, p. 265.

Nefusa anita: Ross, 1951, p. 31; Burks, 1967, p. 17; Smith, 1967, p. 281. Fenusa sp. ?, Shaw, 1940, p. 951.

Female.—Average length, 4.0 mm. Antenna and head black with first and second antennal segments, labrum, and mouth parts yellowish. Thorax black with tegula and pronotum white to yellowish. Legs entirely yellowish. Abdomen mostly black on dorsum, yellowish on venter. Wings very lightly infuscated.

Sheath straight above, rounded below, scopa absent (pl. IV, 48). Lancet with each serrula pointed, with one prominent anterior and

four or five fine posterior subbasal teeth (pl. VII, 86).

Male.—Average length, 4.0 mm. In color and structure similar to female. Genitalia as in plate X, 126 and 127; penis valve long and slender, subrectangular; harpe truncate at apex; parapenis large, rectangular.

Larva.—Shaw (1940) briefly described the larva, but it cannot be characterized by this description. I have not seen specimens.

Holotypes.—M. anita MacGillivray, a female labeled "Wis.," is at the Illinois Natural History Survey. Norton's type of ambiguus cannot be located. In his original description Norton (1867) stated "Penn. 1 &. (Smith. Inst.)"; however, I could not locate such a specimen in the U.S. National Museum. Norton's original description fits the interpretation of this species, and three specimens of this species at the Academy of Natural Sciences of Philadelphia bear a determination label "Fenusa ambigua," apparently written by Norton.

Distribution.—Eastern North America. Maine: Augusta, emgd. VII-2,7,12-1943, Viola; Portland, Baxter Park, June 11, 1967. Maryland: Bowie, V-29-1945; Glen Echo; Cabin John, VIII-21-1917. Massachusetts: Salem, reared, June 27, 1924, violet; Amherst, VIII-40. Michigan: Bay Co., June 2, 1940; Livingston Co., June 5-6, 1943. New York: Ithaca, Aug. 5, 1918. Ohio: Hocking Co., 5-25-30. Pennsylvania: State College, 7-3-47, violet leaf miner; Mt. Holly Springs, VII-4-1918. Quebec: Montreal, 3-VI-

06. Tennessec: Gatlinburg, GSMNP, 5500', July 18, 1947. Wisconsin: "Wis."

Host.—Leaf miner of Viola sp. Shaw (1940) reared it from

"Viola vavilionacea."

Biology.—The only information on the life history of this species is a note by Shaw (1940) on a Fenusa? attacking violet in Massachusetts. Adults were observed from the last of July to the middle of August. They deposit their eggs singly in the upper surface of the leaf. The larvae feed for 22 to 28 days, then go into the soil where they overwinter.

Discussion.—This species is very distinct and is recognized by the presence of a prepectus and the light coloration of the first two antennal segments, labrum, pronotum, legs, and venter of the

abdomen.

#### Genus PROLATUS Smith

Prolatus Smith, 1967, p. 282. Type-species: Prolatus artus Smith. Original designation.

Description.—Antenna with second segment about as long as broad; third segment subequal in length to or only slightly longer than fourth segment (pl. III, 37). Clypeus truncate; malar space linear; postgenal carina absent. Tarsal claw simple, long and slender (pl. III. 21). Front tarsus long, 11/2 times or more length of front tibia; middle tarsus 11/2 times length of middle tibia; hindtarsus shorter than hindtibia (pl. III, 25). Prepectus absent. Forewing with vein 2A and 3A curving up and meeting 1A to form small basal anal cell. Hindwing with cell  $R_1$  open; anal cell absent; cells Rs and M absent (similar to pl. III, 20).

This genus is distinguished by the simple tarsal claw, the unusually long front and middle tarsi, and the subequal third and fourth antennal segments. The tarsal claw and wing venation most

closely resemble those of Fenusa.

The single species of this genus is known only from Oregon and the host is not known.

# Description of Prolatus Species

Prolatus artus Smith

Prolatus artus Smith, 1967, p. 283, 9.

Female.-Length, 3.8 mm. Black with clypeus, labrum, each mandible, tegula, upper posterior margins of pronotum, extreme apex of each femur, and extreme base of each tibia light brown to whitish. Wings uniformly, lightly infuscated.

Head and body, except anterior and lateral lobes of mesonotum, densely covered with fine white hairs. Sheath long and slender, rounded below and at apex (pl. IV. 55). Each serrula of lancet broadly and flatly rounded with eight or 10 subbasal teeth and single lateral tooth at anterior of each serrula (pl. VII, 84).

Male.—See Discussion.

Larva.—Unknown.

Holotype.—U.S.N.M. type No. 69159, a female labeled "Oregon, Josephine Co., 10 mi. N.W. Pinehurst, May 5, 1962, David R. Smith."

Distribution.—Oregon: Pinehurst, May 5, 1962; Corvallis, Scott's Hill, moss and ground litter, March 10, 1960 (a, see Discussion).

Host.—Unknown.

Biology.—Unknown.

Discussion.—A character not included in the description is the presence of a radial crossvein in the hindwing. This is sometimes a variable character in sawflies, and extra veins may appear at random in any species. More specimens will be needed to ascertain its value. A male from near Corvallis, Oreg., fits the description given here, but this specimen is of doubtful association because of the lack of a radial crossvein in the hindwing. It does fit the generic description.

#### Genus FENUSA Leach

Femisa Leach, 1817, p. 126; Westwood, 1840, p. 54; Cameron, 1882, p. 290; Dalla Torre, 1894, p. 156; Ashmead, 1898, p. 250; Konow, 1905, p. 89; MacGillivray, 1914, p. 365; Enslin, 1914, p. 301; MacGillivray, 1916, p. 157; Benson, 1936, p. 623; Ross. 1936, p. 263; Ross., 1937, p. 72; Benson, 1941, p. 87; Berland, 1947, p. 263; Ross, 1951, p. 31; Benson, 1952, p. 108; Takeuchi, 1952, p. 59; Zirngiebl, 1955, p. 93; Lorenz and Kraus, 1957, p. 138; Malaise, 1965; p. 37; Smith, 1967, p. 282.

Type-species: Tenthredo (Emphytus) pionila Klug. Monotypic.

Kalisusanhinga Tischbain, 1816, p. 76; Verrey, 1826, p. 260; Pelle Terrey

Kaliosysphinga Tischbein, 1846, p. 79; Konow, 1886b, p. 269; Dalla Torre, 1894, p. 158; Ashmead, 1898, p. 250; Konow, 1905, p. 89 (Caliosysphinga); Enslin, 1914, p. 301 (= Fennsa Leach); Benson, 1936, p. 623; Zirngiebl, 1955, p. 93.

Type-species: Kaliosysphinga dohrnii Tischbein. Monotypic.

Aphadniurus O. Costa, 1859, p. 40; Dalla Torre, 1894, p. 158 (= Kaliosysphinga Tischbein); Benson, 1936, p. 623 (= Fenusa Leach).

Type-species: Aphadniurus tantillus O. Costa, Monotypic.

Kaliotenusa Viereck, 1910, p. 591; MacGillivray, 1916, p. 157; Benson, 1936, p. 623 (= Fenusa Leach).

Type-species: Fenusa ulmi Sundevall. Monotypic.

Description.—Antenna with second segment as long as broad; third segment longer than fourth segment; segments beyond third slightly decreasing in length (pl. III, 32, 33). Clypeus truncate: malar space less than diameter of front ocellus; postgenal carina absent. Prepectus absent. Tarsal claw simple. Front and middle tarsi subequal in length to their respective tibiae. Forewing with vein 2A and 3A curved up, meeting 1A to form small basal anal cell. Hindwing with cell  $R_1$  open; anal cell absent; cells  $R_2$  and Mabsent (pl. III, 19, 20).

The simple tarsal claw, basal anal cell of the forewing, open radial cell of the hindwing, and short tarsi will distinguish species of this genus. Three species are in this genus, all of which are Holarctic. They are leaf miners of Alnus, Betula, and Ulmus.

Larvae are known for all three species of Fenusa. As a group they are difficult to distinguish from those of Bidigitus and Fenella,

but the following combination of characters will help to differentiate them: Reduced prolegs present on abdominal segments 2 to 8 and 10; dark crescent-shaped marks about prolegs absent; mesosternum and metasternum with light or dark plates; sternum of first abdominal segment with dark plate (except in dohrnii). The generic key (p. 7) to larvae and hosts will help to separate the larvae from those of other Fenusini.

#### Keys to Fenusa Species

#### ADULTS

1. Lateral lobes of mesonotum with fine surface sculpture; abdominal tergites dull with alutaceous surface sculpture; vein 2r of forewing joins Rs before 3r-m; male rare; leaf miner of Ulmus ulmi Sunde ulmi Sundevall Lateral lobes of mesonotum and abdominal tergites smooth, without surface sculpture; vein 2r of forewing joins Rs beyond 3r-m (pl. III, 19)

2. Antenna longer than thorax with third segment only 116 times as long as fourth segment and segments 4 and 5 at least more than twice as long as broad (p. III, 32); hindtibia usually black; male unknown; leaf miner of Alnus dohrnii (Tischbein)

Antenna shorter than thorax with third segment twice as long as fourth segment and segments 4 and 5 less than twice as long as broad (pl. III, 33); hindtibia usually white; males common; leaf miner of Betula ... ... .. . pusilla (Leneletier)

#### LARVAE

1. Large central dark-brown plates on prosternum, mesosternum, metasternum, and sternum of first abdominal segment (pl. XVIII, 232, 233); leaf miner of Betula pusilla (Lepeletier) Small sclerotized plates on mesosternum and metasternum, plate on sternum of first abdominal segment present or absent

2. Small plate present on sternum of first abdominal segment; distinct short spines on venter of ninth abdominal segment and about prolegs of eighth abdominal segment (pl. XVIII, 220-222); leaf miner of Ulmus

Plate on sternum of first abdominal segment absent; venter of apical abdominal segments without spines; leaf miner of Alnus dohrnii (Tischbein)

# Descriptions of Fenusa Species

# Fenusa dohrnii (Tischbein)

Kaliosysphinga dohrnii Tischbein, 1846, p. 80; André, 1880, p. 238; Konow, 1886b, p. 269; Konow, 1890, p. 249; Dalla Torre, 1894, p. 158; Konow, 1905, p. 89; Slingerland, 1905, p. 58.
Fenusa dohrnii: Enslin, 1914, p. 302; MacGillivray, 1916, p. 157; Yuasa, 1922, p. 97; Scidel, 1926, p. 239; Obarski, 1933, p. 157; Conde, 1934, p. 186; Ross, 1936, p. 263; Crevecoeur and Maréchal, 1938, p. 496; Benson, 1940, p. 212; Penson, 1941, p. 200; Rubby 1941, p. 200; Webbyrop, 1944, p. 146; Berson, 1950, p. 200; Crevecoeur and Marcenal, 1938, p. 496; Benson, 1940, p. 213; Benson, 1941, p. 90; Buhr, 1941, p. 909; Wahlgren, 1944, p. 146; Bersond, 1947, p. 264; Hering, 1951, pp. 220, 314; Ross, 1951, p. 32; Wahlgren, 1951, p. 75; Benson, 1952, p. 109; Takeuchi, 1952, p. 60; Maxwell, 1955, p. 59; Raizenne, 1957, p. 25; Lorenz and Kraus, 1957, p. 140; Rafes, 1958, p. 40; Benson, 1962, p. 392; Smith, 1967, p. 282; Benson, 1968, p. 150.

Fenusella dohrnii: Dovnar-Zapolsky, 1929, p. 38. Fenusa curtus Norton, 1862, p. 199, 9; Norton, 1867, p. 225; Provancher, 1888, p. 347 (?); Dalla Torre, 1894, p. 156; Konow, 1905, p. 90; Cresson, 1928, p. 5; Taylor, 1931, p. 451; Ross, 1936, p. 263 (= dohrnii Tischbein).

Phaemisa melanopoda Cameron, 1876a, p. 6.

Fenusa melanopoda: Cameron, 1882, p. 292; Fletcher, 1891, p. 252. Fenusa varipes of nuctt., nec Lepeletier, Dyar, 1893, p. 247; Harrington, 1893,

p. 69; Dyar, 1898, p. 137.

The complete European synonymy is not given here. Benson (1952) gave mclanopoda Cameron and westwoodi Cameron as synonyms of this species.

Female.—Average length, 4.0 mm. Black with front and middle tibiae whitish; hindtibia and all tarsi black to whitish, variable.

Wings moderately, uniformly infuscated.

Antenna more slender and longer than that of *pusilla*; longer than thorax with third segment  $1^{1}$ , times as long as fourth segment and segments 4 and 5 at least more than twice as long as broad (pl. III, 32). Surface sculpture on mesonotum and abdomen absent. Vein 2r of forewing joins Rs apical to 3r-m. Sheath short and rounded (pl. IV, 59). Lancet with each serrula low, flat, with one anterior and about three posterior subbasal teeth (pl. VII, 78).

Male.—Unknown. Parthenogenetic.

Larva.—Final feeding stage (pl. XVIII, 228, 229). Length, 7.0 mm. Similar to the larva of *pusilla* with the following differences: Body evenly covered with minute darkened spines; sternum of first abdominal segment without dark plate; pronotal and sternal plates of thorax lightly sclerotized; mandibles with fewer teeth.

The larva differs from that of *nlmi* by the lack of a plate on the sternum of the first abdominal segment, lack of a concentration of small spines on the venter of the ninth abdominal segment and

about the prolegs, and by having two spines on the lacinia.

Dyar (1893) described six larval stages of this species, and Slingerland (1905) described the larva but could not distinguish it from *ulmi*. Lorenz and Kraus (1957) also described the larva.

Holotypes.—F. curtus Norton, a female labeled "Penna.," is type No. 10317 at the Academy of Natural Sciences of Philadelphia. Tischbein's type may be at the Zoological Museum, Hamburg.

Germany.

Distribution .-- Transcontinental across North America; Europe, Japan: South Africa. The following records are merely a list of localities taken from the specimens I have examined. Alberta: Edmonton; Wabanum, British Columbia: Fernie, Colorado; Eldora; Crystola; Granby: Rocky Mountain Nat'l, Park; Green Mtn. Falls; Cameron Pass; Ward: Empire; Boulder Canyon; Poudre River Canyon. Connecticut: Lyme; Wallingford: Stamford. Illinois: Chicago. Maine: North Bridgeton; Orono. Massachusetts: N. Adams, Michigan: Ann Arbor, New Brunswick: Nerepis, Newfoundland: Cook's Brook; Spruce Brook; Brichy Lk. Bog; Baie Verte Cp.: Terra Nova Nat'l, Park: Aspen Brook Cp.; Catamaran Cp.; Crabbs River Camp; Cormack; Codroy Pond; Corner Brook; Deer Lake: Big Falls, New Hompshire: Franconia, New Mexico: Taos. New York: Ithaca: Brooklyn. Nova Scotia: Antigonish. Ontario: Ottawa; Weston; Swansea. Oregon: Hood River; Wallowa Co.; Wallowa Mtns. Pennsylvania: North East, Saskatchewan: Waskesia, Utah: Hana; Unita Mins, Washington: Nehcotte.

Host.—Leaf miner of Alnus spp. Dyar (1893) bred this species

from "Alnus rugosa (serrulata)" in Massachusetts, and Slingerland (1905) recorded this species from European alder (Almus

glutinosa (L.) Gaertn.).

Biology.—Slingerland (1905) gave the most complete account of the biology of this species from his observations in New York. The adults are active about the middle of May and oviposit in the surface of the leaf, usually in the central part between the larger veins. The larvae feed for about 3 weeks, mining through the leaf, then drop to the ground where they pupate or overwinter in a papery cocoon in the soil. All stages have been reported active through September, and there are probably two or three generations a year. The damage is recognized by the "blisterlike" appearance of the leaves. Up to 12 larvae may feed in a single leaf and their mines coalesce to form the "blisters."

Discussion.—This species is most likely to be confused with pusilla but may be separated by the more slender and longer antenna and darker back tibia. The lack of surface sculpture on the mesonotum and flat serrulae of the lancet will separate dohrnii

from ulmi.

Although Norton described this species in 1862 under the name "curtus," it first received attention in North America when it was found damaging European alder at the Experimental Farm in Ottawa by Fletcher (1891). Harrington (1893) also found this species in Ottawa, Dyar (1893) discovered this species in Massachusetts, and Slingerland (1905) first recorded it from Newark and Ithaca, N.Y.

The European alder leaf miner is the approved common name for this species.

# Fenusa pusilla (Lepeletier)

Tenthredo pumila Klug, 1814, p. 277; Kriechbaumer, 1884, p. 209. Preoccu-

Fenusa pumila: Leach, 1817, p. 126; Stephens, 1835, p. 41; Westwood, 1840, p. 54; Thomson, 1870, p. 272; Thomson, 1871, p. 186; Cameron, 1882, p. 293; Enslin, 1914, p. 303; Britton, 1925, p. 340; Britton, 1926, p. 329; 233; Ensila, 1914, p. 303; Britton, 1929, p. 349; Britton, 1920, p. 629; Dovnar-Zapolsky, 1929, p. 39; Friend, 1931, p. 171; Glasgow, 1932, p. 693; Malaise, 1932, p. 27; Friend, 1933, p. 291; Obarski, 1933, p. 157; Twinn, 1934, p. 76; Marriner, 1936, p. 42; Hering, 1937, p. 75; Balch, 1939, p. 37; Brown, 1940, p. 16; Benson, 1940, p. 213; Buhr, 1941, p. 912; Wahigren, 1944, p. 147; Berland, 1947, p. 261; Hering, 1951, p. 314.

Emphytus pumila: Hartig, 1837, p. 259.

Phaenusa pumila: Cameron, 1876a, p. 7; André, 1880, p. 231.

Kaliosysphinga pumila: Konow, 1885, p. 297; Konow, 1886b, p. 269; Konow, 1890, p. 249; Dalla Torre, 1894, p. 158; Konow, 1905, p. 89; Zirngiebl, 1955, p. 93.

Dolerus pusillus Lepeletier, 1823, p. 120; Blanchard, 1840, p. 241.

Phaenusa pusilla: André, 1880, p. 229. Fenusa pusilla: Ross, 1936, p. 265; Benson, 1940, p. 213; Ross, 1951, p. 31; Wahlgren, 1951, p. 75; Benson, 1952, p. 109; Maxwell, 1955, p. 59; Peterson, 1956, p. 270; Ghent, 1956, p. 17; Raizenne, 1957, p. 25; Lorenz and Kraus, 1957, p. 140; Schuder, 1958, p. 150; Lindquist, 1959, p. 626; Benson, 1962, p. 392; Cheng and LeRoux, 1965, p. 175; Smith, 1967, p. 282; Burks, 1967, p. 16.

Female.—Average length, 3.7 mm. Black with extreme apex of

each femur, each tibia, and tarsus whitish. Wings lightly infus-

cated, darker on basal half.

Antenna stocky, shorter than thorax; third segment twice as long as fourth segment; segments 4 and 5 less than twice as long as broad (pl. III, 33). Mesonotum and abdomen without surface sculpture. Vein 2r of forewing joins Rs apical to 3r-m (pl. III, 19). Sheath short and rounded (pl. IV, 59). Lancet with each serrula low and flattened, with one anterior and five or six posterior subbasal teeth (pl. VII, 79).

Male.—Average length, 3.5 mm. In color and structure similar

to female. Genitalia as in plate X, 124 and 125.

Larva.—Final feeding stage (pl. XVIII, 230-235). Length, 5.5 mm. Slightly dorsoventrally flattened. White; head capsule light brown; plates of thoracic sterna and sternum of first abdominal

segment dark brown.

Head dorsoventrally flattened, round from above, prognathous. Antenna one-segmented. Labrum slightly emarginated with two setae on dorsal surface; epipharynx with row of four or five stout spines on each half; maxillary palpus four-segmented, first, second, and third segments each with one small seta; galea conical, subequal in length to maxillary palpus; lacinia with two stout spines; palpifer with four setae; right mandible with four ventral and two dorsal teeth; left mandible with four ventral and three dorsal teeth; each mandible with one seta on outer basal angle; labial palpus three-segmented; mentum with four setae.

Thorax with dark prosternal, mesosternal, and metasternal plates. Thoracic legs five-segmented, short and directed laterally;

tarsal claw small but present.

Sternum of first abdominal segment with dark plate. Prolegs on abdominal segments 2 to 8. Abdominal segments 1 to 8 each with two annulets. Small spines on venter of ninth segment and about prolegs absent.

The larva has been described by Friend (1933), Peterson (1956),

Lorenz and Kraus (1957), and Lindquist (1959).

Holotypes.—Klug's type is probably in the Zoological Museum of Berlin. The location of Lepeletier's type is not known; it may

be in the Museum of Natural History, Paris.

Distribution.—Northeastern and northwestern United States, southeastern Canada; Europe. The following records are from those specimens I have examined; in States where this species is widespread, specific localities are not given. Connecticut: Widespread. Maine: Widespread. Massachusetts: Widespread. New Brunswick: Widespread. Newfoundland: Insular, widespread. New Hampshire: Widespread. New Jersey: Widespread. New York: Widespread. Nova Scotia: Widespread. Ontario: St. George; Islington. Orcgon: Gresham. Quebec: Widespread in southern part of Province. Vermont: Widespread. This species has also been recorded in the U.S. Agr. Res. Serv. Coop. Econ. Insect Rpt. for 1967 (17 (34):800) from the following States, but I have not seen specimens: Delaware, Indiana. Iowa, Maryland, Michigan, Minnesota, Ohio, Pennsylvania, Rhode Island, Washington, West Virginia, and Wisconsin.

**Host.**—Leaf miner of Betula spp.

Biology.—Friend (1938) published the most complete biological treatment of this species in North America. The adults fly early in the spring, usually in May, and are very common everywhere around birches. There are several generations a year, and the adults may be collected from May to August. Oviposition is on the surface of the leaf, not on the margins as is that of Heterarthrus nemoratus and Messa nana. On maturing, the larvae drop to the soil to pupate or overwinter. The birch forests of the Northeastern States may turn entirely brown as a result of the damage by the mining larvae.

Discussion.—This species may be confused with dohrnii but is separated by the shorter antenna and white back tibia. The lack of surface sculpture on the mesonotum and low flat serrulae of the

lancet will separate it from ulmi.

This species was first discovered in North America by Britton (1925) in Connecticut. It has since become widespread in northeastern United States, eastern Canada, and more recently has been introduced into Oregon and Washington. Friend (1933) reported on its history and included a good bibliography.

The approved common name for this species is the birch leaf

miner.

#### Fenusa ulmi Sundevall

Fenusa ulmi Sundevall, 1844, p. 240; Healy, 1869, p. 297; Kaltenbach, 1874, p. 539; Cameron, 1882, p. 295; Enslin, 1914, p. 302; Dovnar-Zapolsky, 1929, p. 38; Conde, 1934, p. 185; Stritt, 1935, p. 187; Grandi, 1936, p. 246; Skala, 1936, p. 56; Ross, 1936, p. 265; Crevecoeur and Maréchal, 1938, p. 497; Benson, 1940, p. 213; Benson, 1941, p. 89; Buhr, 1941, p. 925; Wahlgren, 1944, p. 147; Berland, 1947, p. 264; Wahlgren, 1951, p. 75; Ross, 1951, p. 31; Benson, 1952, p. 109; Maxwell, 1955, p. 58; Zirngiebl, 1955, p. 93; Raizenne, 1957, p. 25; Lorenz and Kraus, 1957, p. 140; Benson, 1962, p. 392; Smith 1967, p. 295

1902, p. 392; Smith, 1967, p. 282,

Phaenusa ulmi: Cameron, 1876a, p. 9; André, 1880, p. 230.

Kaliosysphinga ulmi: Konow, 1886b, p. 269; Konow, 1890, p. 249; Dalla Torre,
1894, p. 159; Slingerland, 1905, p. 49; Konow, 1905, p. 89; Herrick, 1913,

Kaliofenusa ulmi: Viereck, 1910, p. 591; MacGillivray, 1916, p. 157; Yuasa, 1922, p. 97; Twinn, 1934, p. 77.

"an elm leafminer." Felt, 1898a, p. 21; Felt, 1898b, p. 237. Messa alsia MacGillivray, 1923b, p. 22, 9; Frison, 1927, p. 251; Ross, 1936, p. 265 (=ulmi Sundevail).

Female.—Average length, 4.0 mm. Entirely black with front tibia whitish and middle tibia and hindtibia brownish. Wings uniformly, lightly infuscated.

Antenna stocky; third segment 114 times length of fourth segment; segments 4 and 5 less than twice as long as broad (pl. III, 33). Lateral lobes of mesonotum with fine surface sculpture; abdominal tergites dull with alutaceous surface sculpture. Vein 2r of forewing meets Rs basal to 3r-m. Sheath short, rounded (pl. IV, 58). Lancet with each serrula long, lobelike, rounded at apex, without subbasal teeth (pl. VII, 77).

Male.—Unknown in North America. Rare in Europe (Benson,

1952).

Larva.—Final feeding stage (pl. XVIII, 220-227). Length, 7.8 mm. Slightly dorsoventrally flattened. White; head capsule, notal and sternal plates of thorax, and sternal plates of first three abdominal segments light brown. Dorsum and venter of body with fine microscopic setae, larger and more abundant around prolegs of abdominal segments 7 and 8 and on ventral swellings of abdominal segment 9.

Head dorsoventrally flattened; prognathous. Labrum slightly emarginated, two setae on dorsal surface; epipharynx with six stout spines on each half; right mandible with three ventral and two dorsal teeth; left mandible with three ventral and three dorsal teeth; one seta on outer basal angle of each mandible; maxillary palpus four-segmented; galea conical, subequal in length to maxillary palpus; lacinia with three stout spines; three setae on palpifer; labial palpus three-segmented, conspicuous; mentum with four setae.

Pronotal plate present; large prosternal plate and small mesosternal and metasternal plates present. Mesonotum and metanotum with dorsal swellings lacking microscopic setae. Thoracic legs five-segmented, small; tarsal claw minute, hooklike, appearing absent but visible under high magnification.

Venter of abdominal segments 1 to 4 with small light central plates. Prolegs present on segments 2 to 8, each with numerous microscopic spines about them; swellings on venter of segment 9

with concentration of small spines.

Healy (1869a), Slingerland (1905), Yuasa (1922), and Lorenz and Kraus (1957) also described the larva of ulmi. Maxwell (1955) described the internal larval anatomy.

Holotypes.—M. alsia MacGillivray, a female labeled "Ithaca, N.Y., 16 May 97," is at the Illinois Natural History Survey. The

location of Sundevall's type is not known.

Distribution.—Northeastern United States and southeastern Canada. The following records are from those specimens that I have examined. Massachusetts: Danvers, V-15-26, Ulmus. Michigan: East Lansing, May 15, 17, 1963. New York: Ithaca, June 6, 1904, V-22-1926. V-17-1963. June 2, 1904; Van Natta's Dam, Ithaca, May 23, 1937; Inlet Valley, Ithaca, V-3-1913; New Rochelle, May 9, 1925; Geneva, May 17, 1954, V-18-15; Delmar, May 15, 1906; Albany, April 5, 1903. Ontario: Weston, May 29, 1934, Quebec: St. Hillaire, 24-V-22.

Host.-Leaf miner of elm (Ulmus spp.). Felt (1898a) recorded it from American elm. However, Slingerland (1905) recorded it from English and Scotch elms in New York and stated that the American elms were immune from attack even where the

different host species were growing in close proximity.

Biology.—Slingerland (1905) reported on the biology of this species. The adults fly from the middle of May to the first of June. The eggs are usually inserted near the midrib of the leaf, sometimes as many as 25 per leaf. The larvae feed for about 3 weeks, and, as they develop, the mines may coalesce to give a blisterlike appearance to the leaf. About mid-July the larvae drop to the

ground and form thin papery cocoons in the soil where they over-

winter. There is one generation a year.

**Discussion.**—The surface sculpture of the mesonotum and abdomen, the position of vein 2r in the forewing, and the lobelike serrulae of the lancet will separate this species from other **Fenusa** species.

This is probably an introduced species that was brought to America with the imported elms. The first record for North America is that by Felt (1898a), who recorded it from New York as being a pest of elms. Slingerland (1905) first found this species in New York in 1899.

The approved common name for this species is the elm leaf miner.

#### Genus FENELLA Westwood

Fenella Westwood, 1840, p. 54; Cameron, 1882, p. 288; Konow, 1886b, p. 271;
Dalla Torre, 1894, p. 155; Ashmead, 1898, p. 250; Konow, 1905, p. 90;
Enslin, 1914, p. 308; Benson, 1936, p. 623; Benson, 1941, p. 87; Berland, 1947, p. 267; Benson, 1952, p. 109; Benson, 1953, p. 136; Lorenz and Kraus, 1957, p. 138; Smith, 1967, p. 281.
Type-species: Fenella nigrita Westwood. Monotypic.

Description.—Antenna with 11 or 12 segments; second segment longer than broad; third segment longer than fourth segment; segments beyond third subequal in length (pl. III, 34). Clypeus truncate; malar space less than diameter of front ocellus; postgenal carina absent. Tarsal claw simple, basal lobe absent (pl. III, 21); fourth segment of back tarsus not conspicuously produced apically (pl. III, 28). Forewing with vein 2A and 3A curved up and joining 1A forming small basal anal cell. Hindwing with radial cell open; anal cell absent; cells Rs and M absent (similar to pl. III, 19, 20).

Fenella is close to Fenusa, but it may be separated by the longer antenna and the less apically produced fourth segment of the back tarsus. Also, members of this genus are associated with herbs, whereas members of Fenusa are associated with trees.

This is a small genus of about six species, only one of which has been found in North America. Benson (1953) treated five species in a revision of this genus.

# Description of Fenella Species

# Fenella nigrita Westwood

Fenella nigrita Westwood, 1840, p. 54; Thomson, 1870, p. 270; Thomson, 1871, p. 180; Cameron, 1876a, p. 15; André, 1880, p. 233; Cameron, 1882, p. 288; Brischke and Zaddach, 1883, p. 257; Kenow, 1886b, p. 271; Konow, 1887, p. 276; Konow, 1890, p. 249; Dalla Torre, 1894, p. 155; Ghigi, 1904, p. 16; Enslin, 1914, p. 308; Dovnar-Zapolsky, 1929, p. 39; Dovnar-Zapolsky, 1931, p. 55; Conde, 1934, p. 186; Stritt, 1935, p. 187; Grandi, 1936, p. 250; Kvicala, 1938, p. 151; Benson, 1940, p. 213; Benson, 1941, p. 89; Buhr, 1941, p. 908; Wahlgren, 1944, p. 148; Berland, 1947, p. 268; Wahlgren, 1951, p. 76; Benson, 1952, p. 109; Benson, 1953, p. 136; Lorenz and Kraus, 1957, p. 140; Ermolenko, 1961, p. 94; Smith, 1967, p. 281.

Female.—Average length, 3.0 mm. Black with labrum, extreme apex of each femur, each tibia, and each tarsus brownish. Wings

uniformly, moderately infuscated.

Sheath short, rounded, without scopa at apex (pl. IV, 54). Basal serrulae of lancet pointed, with no anterior and seven or eight fine posterior subbasal teeth; apical serrulae flattened, each with seven or eight fine teeth on apical flattened margin (pl. VII, 82).

Male.—Unknown. Parthenogenetic.

Larva.—Final feeding stage (pl. XVII, 213-219). Length, 6.0 mm. Dorsoventrally flattened. White; head capsule, pronotal plate, large prosternal plate, and small mesosternal and metasternal plates light brown; dorsum of 10th abdominal segment very light brown.

Head dorsoventrally flattened, prognathous; oblong in dorsal view. Antenna conical, indistinctly three-segmented. Labrum slightly emarginated with two setae on dorsum; epipharynx with row of five short spines on each half; each mandible with three teeth; one seta on outer basal angle of each mandible; maxillary paipus four-segmented, palpifer with two setae; galea long, subequal in length to maxillary palpus; lacinia with two long stout spines; labial palpus three-segmented, long and distinct; mentum with four setae.

Pronotal plate present; large prosternal and small mesosternal and metasternal plates present. Thoracic legs five-segmented, short,

directed laterally; tarsal claw present.

Prolegs on abdominal segments 2 to 8, indistinct on 10. Abdominal segments 1 to 8 indistinctly two-annulate. Small plate on sternum of first abdominal segment. Dorsum of 10th abdominal segment slightly darkened, with numerous setae.

Lorenz and Kraus (1957) described the larva of this species.

Holotype.-Westwood's types are in the Hope Museum, Oxford,

England,

Distribution.—Northeastern United States and southeastern Canada; Europe. Connecticut: Canaan, 15-VIII-65, A. Stone. Maine: Penobscot Co., 3 mi. N. Passadumkeag, May 20, 1966, D. R. Smith. Michigan: E. Lansing, May 17, 1963, F. E. Giles, May 18, 1962, G. Eickwort. Ontario: Ottawa, ex Potentilla, July 20, 1964, W. R. Richards.

Host.—Leaf miner of Potentilla sp. In Britain it is found on Agrimonia eupatoria L. and Potentilla reptans L. (Benson, 1952).

Biology.—There is no North American literature on the life history of this species. In Europe Wahlgren (1944, 1951) and Lorenz and Kraus (1957) published some life-history notes.

*Discussion.*—This small species was first collected in North America at Ottawa, Ontario. Since then several specimens have been found from other localities. It is identical to the European specimens of *nigrita* and may be an introduced species.

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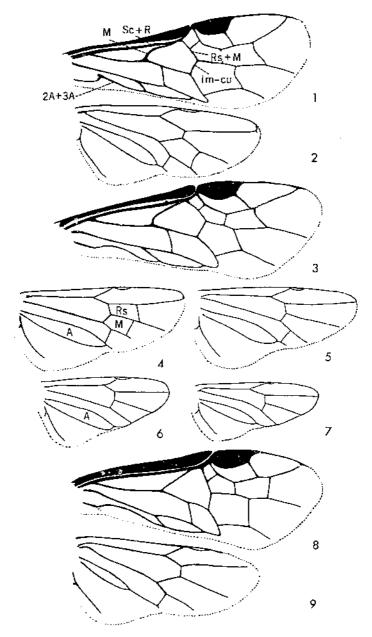
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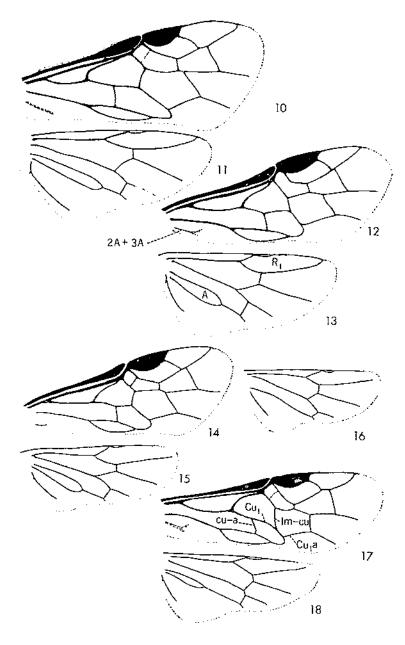
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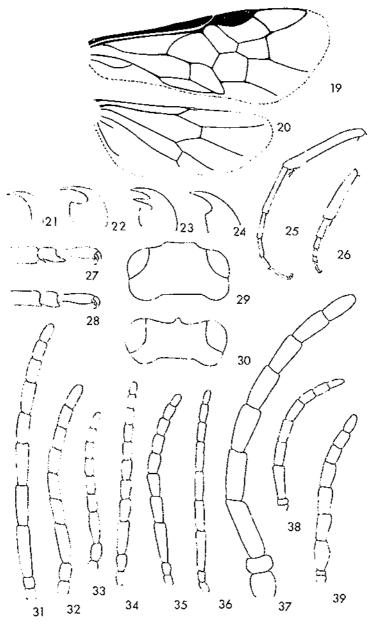
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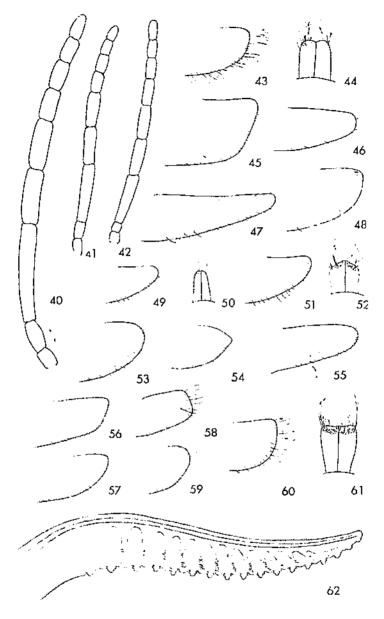
Forewing (1) and hindwing (2) of Endelonyia aethiops; forewing (3) and hindwing (4) of Caliroa cerasi; hindwing of C. lorata female (5), C. fasciata male (6), and C. lorata male (7); forewing (8) and hindwing (9) of Heterarthrus nemoratus. [A = anal cell; 2A + 3A = second and third anal vein; M = media, medial cell; 1m-cu = first mediocubital crossvein; Rs = radial sector, radial sector cell; Rs + M = radial sector and media; Sc + R = subcosta and radius.]



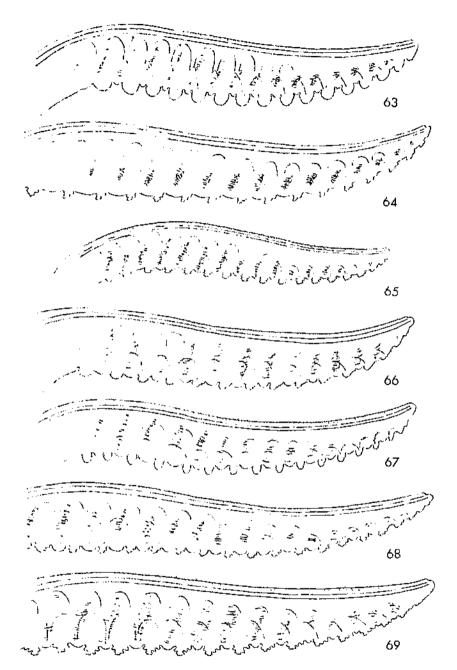
Forewing (10) and hindwing (11) of Metallus rohweri; forewing (12) and hindwing; (13) of Messa leucostoma; forewing (14) and hindwing (15) of Profenusa canadensis; hindwing of P. thomsoni (16); forewing (17) and hindwing (18) of Billightus platani. [A = anal cell; 2A + 3A = second and third anal vein;  $Cu_0 = \text{cubital anal vein}; cu_0 = \text{cubital anal vein}; cu_0 = \text{cubital anal cell}; 1m-cu = \text{first mediocubital crossvein}; <math>R_1 = \text{radial cell}.$ 



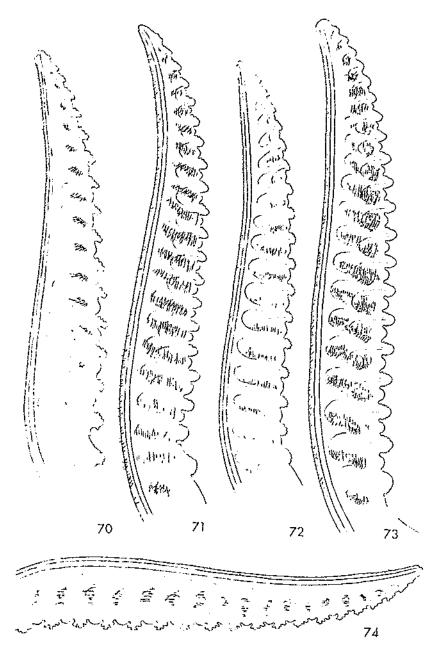
Forewing (19) and hindwing (20) of Fennsa pusilla; tarsal claws of F. pusilla (21), Nefusa ambigua (22), Bidigitus platani (23), and Setabara histrionica (24); front legs of Prolatus artus (25) and Fenusa pusilla (26); apex of hindtarsus of F. pusilla (27) and Fencila nigrita (28); hend, dorsal, of Caliroa cerasi (29) and Heterarthrus nemoratus (30); antennae of H. nemoratus (31), Fenusa dohrnii (32), F. pusilla (33), Fenella nigrita (34), Messa hortulana (35), M. nana (36), Prolatus artus (37), Metallus rohweri female (38), and M. rohweri male (39).



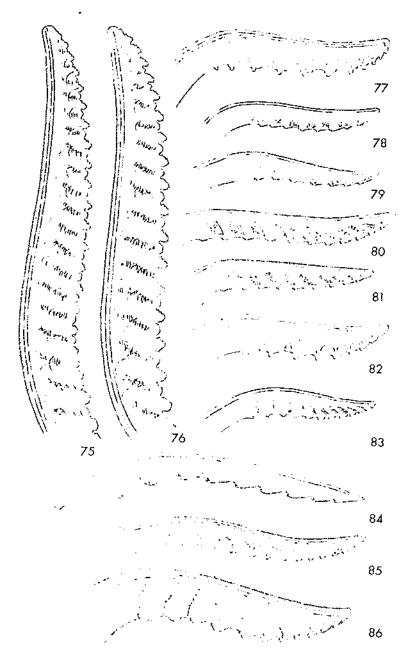
Antennae of Caliroa fasciata (40), C. liturata (41), and Endelonyia aethiops (42). Female sheaths: Lateral (43) and dorsal (44) of Heterarthrus nemoratus; lateral of Metallus rohweri (45), M. capitalis (46), Bidigitus platani (47), and Nefusa ambigua (48); lateral (49) and dorsal (50) of Messa nana; lateral (51) and dorsal (52) of M. hortulana; lateral of Profesusa canadensis (53), Fenella nigrita (54), Prolatus artus (55), Profesusa alumna (56), P. thomsoni (57), Fenusa ulmi (58), and F. pusilla (59); lateral (60) and dorsal (61) of Setabara histrionica. Female lancet of Euclelonyia aethiops (62).



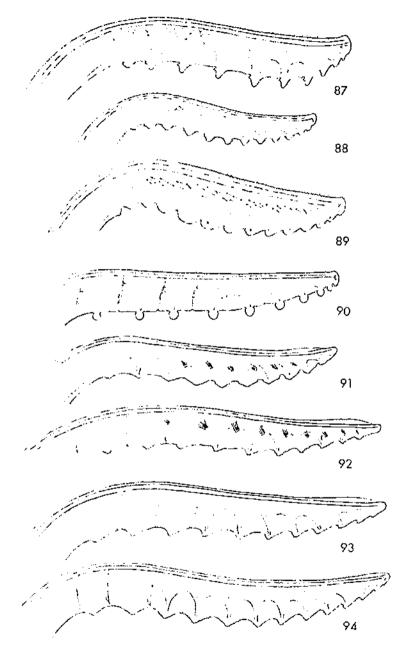
Female lancets of Caliroa distincta (63), C. hyalina (64), C. liturata (65), C. petiolata (66), C. lunata (67), C. lorata (68), and C. labrata (69).



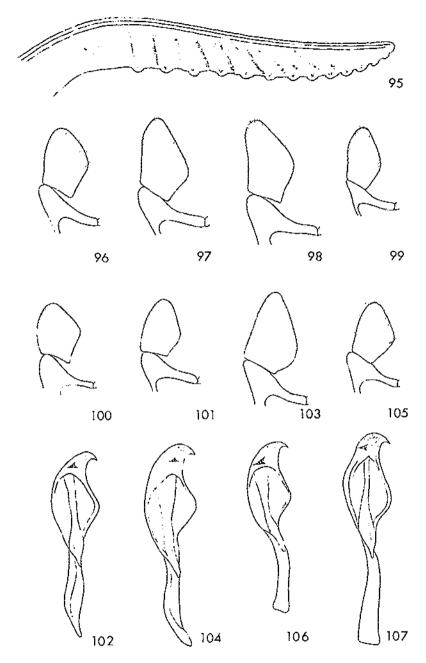
Female lancets of Caliroa cerasi (70), C. floridana (71), C. nyssae (72), C. lobata (73), and C. obsoleta (74).



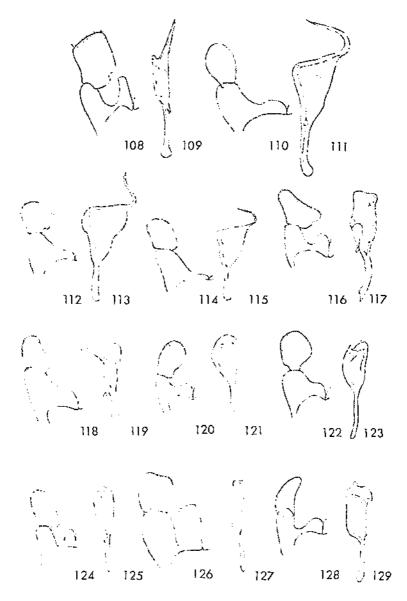
Female lancets of Caliroa fasciata (75), C. querouscoccineae (76), Fenusa ulmi (77), F. dohrnii (78), F. pusilla (79), Messa populifoliella (80), M. wuestneii (81), Fenella nigrita (82), Setabara histrionica (83), Prolatus artus (84), Bidigitus platani (85), and Nafusa ambigna (86).



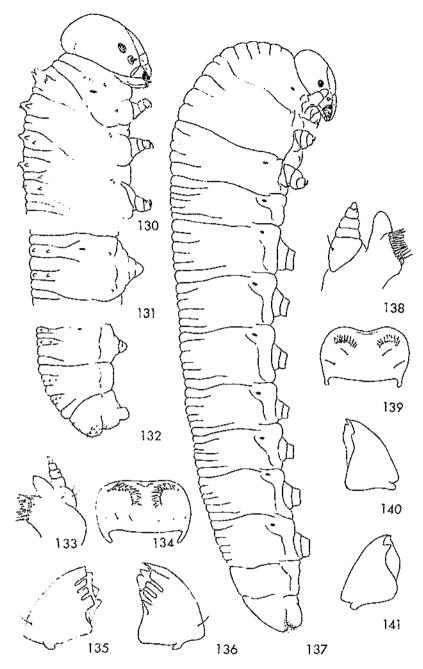
Female lancets of Metallus bensoni (87), M. capitalis (88), M. rohweri (89), Profenusa canadensis (90), P. thomsoni (91), P. alumna (92), P. inspirata (93), and P. lucifex (94).



Female lancet of Messa nana (95); male harpe and parapenis of Caliroa hyalina (96), C. labrata (97), C. lorala (98), C. obsoleta (99), C. petiolata (100); harpe and parapenis (101) and penis valve (102) of C. liturata; harpe and parapenis (103) and penis valve (104) of C. fasciata; harpe and parapenis (105) and penis valve (106) of C. nyssae; penis valve of C. lobata (107).

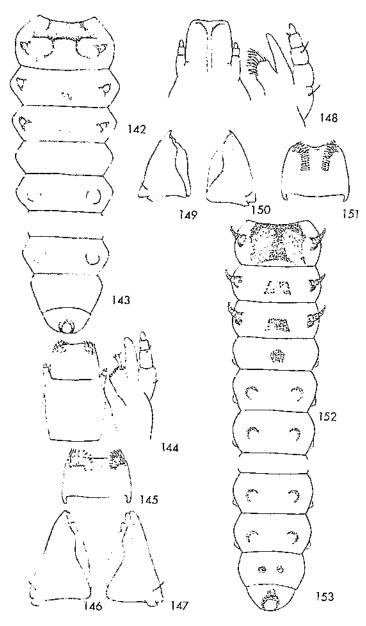


Male harpe and parapenis (108) and penis valve (109) of Metallus rohweri; harpe and parapenis (110) and penis valve (111) of Messa populifoliella; harpe and parapenis (112) and penis valve (113) of M. leucostoma; harpe and parapenis (114) and penis valve (115) of M. wuestneii; harpe and parapenis (116) and penis valve (117) of Setabara histrionica; harpe and parapenis (118) and penis valve (119) of Professa canadensis; harpe and parapenis (120) and penis valve (121) of P. alumna; harpe and parapenis (122) and penis valve (123) of P. inspirata; harpe and parapenis (124) and penis valve (125) of Fenusa pusilla; harpe and parapenis (126) and penis valve (127) of Nefusa ambigua; harpe and parapenis (128) and penis valve (129) of Bidigitus platani.

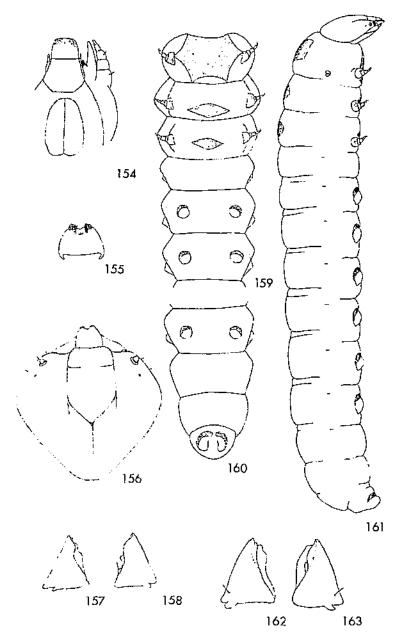


Endelomyia aethiops larva: Head and thorax (130); third abdominal segment (131); eighth, ninth, and 10th abdominal segments (132); maxilla (133); epipharynx (134); right (135) and left (136) mandibles, ventral. Caliroa cerasi larva: Lateral (137); maxilla (138); epipharynx (139); left (140) and right (141) mandibles, ventral.

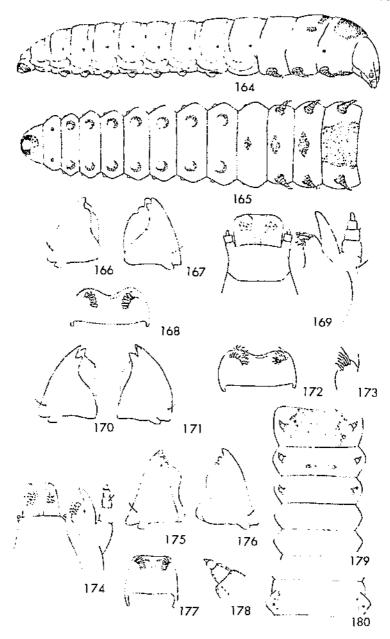
### TEA1420 (1971) USON TECHNOCHE BUCLETINS OFDATA NERCIIC SAWELIES III HETERARTHRINAE ADULTS AND LARVAE



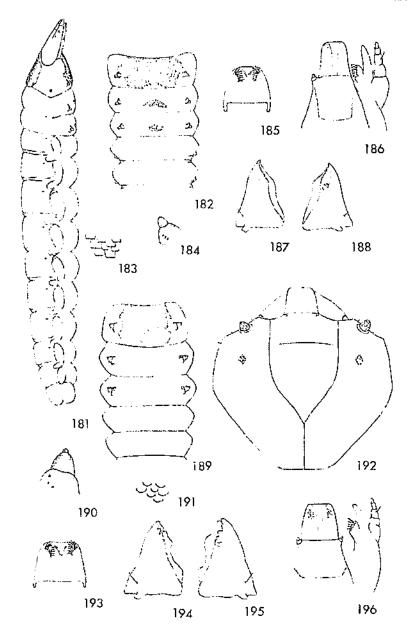
Heterarthrus nemoratus larva: Thorax and first two abdominal segments, ventral (142): eighth, ninth, and 10th abdominal segments, ventral (143); labium and maxilla, ventral (144); epipharynx (145); right (146) and left (147) mandibles, ventral. Messa nana larva: Labium and maxilla, ventral (148); right (149) and left (150) mandibles, ventral; epipharynx (151); thorax and first three abdominal segments, ventral (152); abdominal segments 7 to 10, ventral (153).



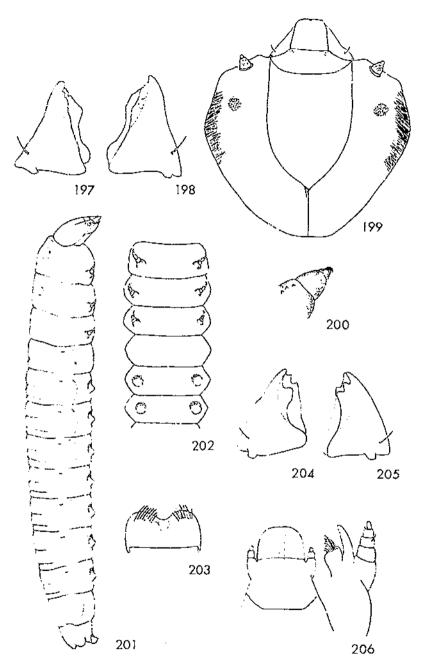
Metallus rohweri larva: Labium and maxilla, ventral (154); epipharynx (155); head, dorsal (156); right (157) and left (158) mandibles, ventral; thorax and first three abdominal segments, ventral (159); abdominal segments 7 to 10, ventral (160); entire larva, lateral (161). Metallus capitalis larva: Right (162) and left (163) mandibles, ventral.



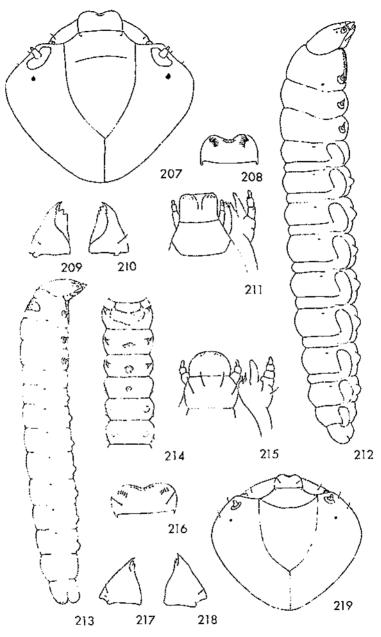
Messa sp. larva: Entire larva, lateral (164); thorax and abdomen, ventral (165); right (166) and left (167) mandibles, ventral; epipharynx (168); labium and maxilla, ventral (169). Messa sp. larva: Right (170) and left (171) mandibles, ventral; epipharynx (172); lacinia (173). Profesusa inspirata larva: Labium and maxilla, ventral (174); right (175) and left (176) mandibles, ventral; epipharynx (177); thoracic leg (178); thorax and first two abdominal segments, ventral (179); third abdominal segment, dorsal (180).



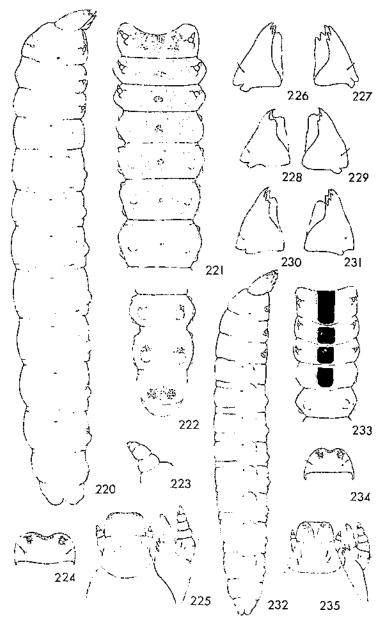
Profenusa thomsoni larva: Entire larva, lateral (181); thorax and first two abdominal segments, ventral (182); spines of integument (183); thoracic leg (184); epipharynx (185); labium and maxilla, ventral (186); right (187) and left (188) mandibles, ventral. Profenusa alumna larva: Thorax and first two abdominal segments, ventral (189); thoracic leg (190); spines of integument (191); head, dorsal (192); epipharynx (193); right (194) and left (195) mandibles, ventral; labium and maxilla, ventral (196).



Profesusa lucifex larva: Right (197) and left (198) mandibles, ventral; head, dorsal (199); thoracic leg (200). Profesusa canadensis larva: Entire larva, lateral (201); thorax and first three abdominal segments, ventral (202); epipharynx (203); right (204) and left (205) mandibles, ventral; labium and maxilla, ventral (206).



Bidigitus platani larva: Head, dorsal (207); epipharynx (208); right (209) and left (210) mandibles, ventral; labium and maxilla, ventral (211); entire larva, lateral (212). Fenella nigrita larva: Entire larva, lateral (213); thorax and first three abdominal segments, ventral (214); labium and maxilla, ventral (215); epipharynx (216); right (217) and left (218) mandibles, ventral; head, dorsal (219).



Fenusa ulmi larva: Entire larva, lateral (220); thorax and first four abdom'al segments, ventral (221); apical four abdominal segments, ventral (224); thoracic leg (223); epipharynx (224); labium and maxilla, ventral (225); right (226) and left (227) mandibles, ventral. Fenusa dohrnii larva: Right (228) and left (229) mandibles, ventral. Fenusa pusilla larva: Right (230) and left (231) mandibles, ventral; entire larva, lateral (232); thorax and first two abdominal segments, ventral (233); epipharynx (234); labium and maxilla (235).

# III NI